

SHORT REPORT

Open Access

Dermanyssus gallinae in layer farms in Kosovo: A high risk for *Salmonella* prevalence

Afrim Hamidi^{1*}, Kurtesh Sherifi¹, Skender Muji¹, Behlul Behluli¹, Fatgzim Latifi¹, Avni Robaj¹, Rezart Postoli², Claudia Hess³, Michael Hess³ and Olivier Sparagano⁴

Abstract

Background: The poultry red mite (PRM), *Dermanyssus gallinae* (*D.g.*) is a serious ectoparasitic pest of poultry and potential pathogen vector. The prevalence of *D. g.* and the prevalence of *Salmonella* spp. within mites on infested laying poultry farms were investigated in Kosovo.

Findings: In total, 14 populated layer farms located in the Southern Kosovo were assessed for *D. g.* presence. Another two farms in this region were investigated 6 months after depopulation. Investigated flocks were all maintained in cages, a common housing system in Kosovo. A total of eight farms were found to be infested with *D. g.* (50%) at varying levels, including the two depopulated farms. The detection of *Salmonella* spp. from *D. g.* was carried out using PCR. Out of the eight layer farms infested with *D. g.*, *Salmonella* spp. was present in mites on three farms (37.5%).

Conclusions: This study confirms the high prevalence of *D. g.* in layer flocks in Kosovo and demonstrates the link between this mite and the presence of *Salmonella* spp. on infested farms.

Background

The PRM *D. g.* is one of the most serious and widespread pests of poultry production. It is a worldwide ectoparasitic pest [1], showing genetic variation between the UK, France and Italy [2]. *D. g.* hides in cracks and crevices in poultry units and infests the birds only briefly for blood meals, mainly at night [3]. This haematophagous mite feeds rapidly from its host (in comparison with ticks for example) and can survive several months without a blood-meal. The mites enter poultry houses through open wall fans, wall inlets and air chimneys [4] or are brought by staff, bird cages, crates or wild birds. In recent years, the frequency of *D. g.* infestations in laying hens has increased in Europe, as has its pest status.

D. g. can have a significant economic impact in poultry production by causing a reduction in egg production, loss in body weight of birds, and a reduction of welfare of laying hens [5]. *D. g.* may also cause dermatitis in humans [6,7]. Infestation with *D. g.* can cause irritation and restlessness in affected hens and can even result in the death of birds [8]. It is reported that the death rate among the

hens can rise from 1 to 4%, with a reduction in laying performance of up to 10%, as a result of infestation. Downgrading of egg quality in poultry affected by *D. g.* has also been observed [9,10].

In addition to causing 'direct' losses in poultry production systems, *D. g.* has also been described as a potential vector of several bacteria and viruses of concern to poultry, among them several food borne pathogens [11-14].

In order to assess levels of *D. g.* infestation on farms different mite traps have been evaluated and used as indicators to inform control decisions [15,16]. Effective options available for control of *D. g.* are limited [17]. The use of inadequate, ineffective, or even illegal chemicals have been responsible for increases in infestation rates due to the spread of acaricide resistance now common in many countries [1]. Nevertheless, some products are still being developed and used with some success. For example, [18] found that phoxim 50% is an effective acaricide against *D. g.* with [19] showing the same to be true of spinosad.

D. g. is widespread among farms in Kosovo and might cause high economic losses within the national poultry industry.

* Correspondence: afrimhamidi@hotmail.com

¹Faculty of Agriculture and Veterinary, University of Prishtina, Kosovo
Full list of author information is available at the end of the article

Infestation rate

Fourteen populated layer farms (6,000 - 20,000 birds, from 6 to 12 months of age) and two de-populated farms located in Southern Kosovo were investigated for the prevalence of *D. g.* (Table 1). The 16 farms all had generally poor biosecurity measures, such as lack of traffic control and personal hygiene.

D. g. prevalence was determined using cardboard traps, which were placed into cages, but out of reach of the birds. Traps were left in place for 48 h, after which they were removed and evaluated for the presence of *D.g.* Ten cardboard traps were used at each farm and combined as a single pooled sample on the day of their collection. Traps, which were positive for *D. g.*, were frozen at - 20°C to kill any mites present. Mite numbers were assessed and the number of individuals (adults, larvae and nymphs) per trap were estimated to the nearest 50.

Incidence of *Salmonella* spp

Pooled samples of *D. g.* adults, nymphs and larvae (of between 50 and 100 mites) from each infested farm (n = 8), were selected in the laboratory and placed into plastic tubes and stored at -20°C. All samples (n = 80) of *D. g.* were examined for the presence of *Salmonella* using PCR.

Bacterial DNA was extracted using Dneasy Tissue Kit (QIAGEN) according to the manufacturer's instructions. Extracted DNA (25 µl/pooled mite sample) was stored at - 20°C. The PCR reaction mixture (25 µl) contained: 5 µl DNA sample, 14.05 µl distilled H₂O, 2.5 µl 10xPCR Buffer, 0.75 µl MgCl₂ (50 mM), 0.5 µl dNTP (10 mM), 1 µM of each primer and 1.25 U Taq DNA Polymerase. DNA was amplified according to [20]. The 16S rDNA primer sequences were: (forward)5'-TGT TGT GGT TAA TAA CCG CA-3' and (reverse)5'-CAC AAA TCC ATC TCT GGA-3'. PCR amplifications were performed in thermal cycler with a cycling program consisting of a 10 min denaturation step 94 C, followed by 35 cycles of denaturation (1 min, 94°C), annealing (45 s, 55°C) and extension (1 min 30 s, 72°C) and final extension step of 10 min at 72°C [21]. After 35 cycles the amplification product was expected to be 574 base pairs.

Amplification products, including their size, were determined by electrophoresis using 2% agarose gels.

Results and discussion

In total 8 out of 16 farms (50%) were infested with *D. g.* to varying degrees, where mites were present in both populated and unpopulated units. Out of the eight infested farms, pooled mite samples from three premises were positive for the presence of *Salmonella* spp. (37.5%). In two cases *Salmonella* was found in mites 6 months after the removal of layers (Farms 1 and 8). The other *Salmonella*-positive case was from mites trapped in a populated unit (Farm 2). Results are shown in Table 1.

D. g. represents a major problem for the layer industry in Kosovo and across much of the globe. Data from the present study showed that 50% of Kosovan cage rearing systems were infested with *D. g.*, which is comparable to prevalence figures in other countries with free range rearing systems, e.g. France (56%), UK (60%), and Denmark (68%) [1]. A high *Salmonella* spp. prevalence on infested farms was also observed, which demonstrates that *D. g.* may at least carry this pathogen potentially serving as vector for its spread in poultry. Such a vectorial role has already been observed under laboratory conditions [22].

As mites tested positive for *Salmonella* spp. in depopulated units, the role that *D. g.* can play in between-flock transmission of pathogens was also demonstrated by the present study. *Salmonella* was found within *D. g.*, 6 months after the removal of birds from the infested farm. This further suggests that transovarial transmission of *Salmonella* by *D. g.* is a possibility considering that mites do not survive for so long they must have passed the infection to their offspring. These same results show that *D. g.* can survive long periods of fasting, suggesting that these mites may be capable of pathogen transmission between flocks. Finally, where *Salmonella* was found in mites present on a populated farm, no signs of clinical Salmonellosis were observed in the birds. This could pose a risk to public health through *Salmonella*-positive mites being squashed on eggs or when such mites would infect birds.

Table 1 Data on *D. g.* samples from poultry farms in Kosovo

Farm	1	2	3	4	5	6	7	8
nr. of samples	10	10	10	10	10	10	10	10
nr. of PRM/sample	50	50-100	100	50	50	50	50-100	50-100
layers on the farm	No	Yes	Yes	Yes	Yes	Yes	Yes	No
nr. of layers on the farm	0*	6,000	12,000	15,000	20,000	18,000	16,000	0
<i>Salmonella</i> PCR	+	+	-	-	-	-	-	+

*Depopulated more than 6 months before sampling

The transfer of *D. g.* in Kosovo seems to occur via egg boxes which are exchanged at supermarkets, originating from different egg supply companies. In general all farms infested with *D. g.* in the present study demonstrated poor biosecurity measures. The emerging potential role of *D. g.* as a vector for *Salmonella* spp. supported by the results presented in this paper it should encourage the poultry industry to initiate strict biosecurity programs to control both *D. g.* and any pathogens it may foster, carry or transmit. Assuming that these results are not specific to Kosovo, they could explain why salmonellosis outbreaks are still observed in other countries, such as the UK. Although all birds are vaccinated against *Salmonella* the prevalence of *D. g.* is very high in the UK, which could explain why *Salmonella* transmission can overcome the vaccination effect as different serovars exist.

List of Abbreviations

D.g.: *Dermanyssus gallinae*; PRM: poultry red mite.

Acknowledgements and Funding

We thank the ASSO (Austrian Scientific Support Office) and Austrian Federal Ministry for Education for funding of this scientific activity.

Author details

¹Faculty of Agriculture and Veterinary, University of Prishtina, Kosovo.

²Faculty of Veterinary Medicine, Tirana, Albania. ³Clinic for Poultry, Fish and Reptiles, Veterinary Medical University Vienna, Austria. ⁴Northumbria University, School of Life Sciences, Newcastle upon Tyne, UK.

Authors' contributions

AH worked on study design, sampling, data analysis and drafting the paper. KS participated in sampling, carried analysis together with CH at the clinic of Poultry, Reptiles and Fishes at the Veterinary Medical University in Vienna. MH worked on study design, data analysis and drafting the paper. SM, BB, FL, AR and RP participated in sampling of *D. g.* OS worked on the data analysis and drafted the paper with AH. All authors read and approved the final version of the manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 2 June 2011 Accepted: 15 July 2011 Published: 15 July 2011

References

1. Sparagano O, Pavlićević A, Murano T, Camarda A, Sahibi H, Kilpinen O, Mul M, van Emous R, le Bouquin S, Hoel K, Cafiero MA: **Prevalence and key figures for the poultry red mite *Dermanyssus gallinae* infections in poultry farm systems.** *Exp Appl Acarol* 2009, **48**:3-10.
2. Marangi M, Cafiero MA, Capelli G, Camarda A, Sparagano OAE, Giangaspero A: **Evaluation of the poultry red mite, *Dermanyssus gallinae* (Acari: Dermanyssidae) susceptibility to some acaricides in field populations from Italy.** *Exp Appl Acarol* 2009, **48**:11-18.
3. Fiddes MD, Le Gresley S, Parsons DG, Epe C, Coles GC, Stafford KA: **Prevalence of the poultry red mite (*Dermanyssus gallinae*) in England.** *Vet Record* 2005, **157**:233-235.
4. Meyer-Kühling B, Heine J, Müller-Lindloff J, Pfister K: **Epidemiology of *Dermanyssus gallinae* and acaricidal efficacy of Phoxim 50% in alternative housing systems during the laying period of hens.** *Parasitol Res* 2007, **101**:S1-S12.
5. Chauve C: **The poultry red mite *Dermanyssus gallinae* (De Geer, 1778): current situation and future prospects for control.** *Vet Parasitol* 1998, **79**:239-245.

6. Cafiero MA, Camarda A, Circella E: **Pseudoscabies caused by *Dermanyssus gallinae* in Italian city dwellers: a new setting for an old dermatitis.** *J European Acad Dermatol Venerol* 2008, **22**:1382-1383.
7. Cafiero MA, Camarda A, Circella E, Galante D, Lomuto M: **An urban outbreak of red mite dermatitis in Italy.** *Internat J Dermatol* 2009, **48**:1119-1121.
8. Eckert J, Friedhoff KT, Zahner H, Deplazes P: **Lehrbuch der Parasitologie für die Tiermedizin.** Enke Verlag, Stuttgart; 2005, 369-371.
9. Kirkwood AC: **Anaemia in poultry infested with the red mite *Dermanyssus gallinae*.** *Vet Records* 1967, **80**:514-516.
10. Van Emous RA, Fiks-Van Niekerk TGCM, Mul MF: **€ 11 million damage for the sector: enquiry into the cost of mites to the poultry industry.** *De pluimveehouderij* 2006, **35**:8-9.
11. Chirico J, Eriksson H, Fossum O, Jansson D: **The poultry Red Mite, *Dermanyssus gallinae*, a potential vector of *Erysipelothrix rhusiopathiae* causing erysipelas in hens.** *Med Vet Entomol* 2003, **17**:232-234.
12. De Luna CJ, Arkle S, Harrington D, George D, Guy J, Sparagano OAE: **The poultry red mite (*Dermanyssus gallinae*) as a potential carrier of vector-borne diseases.** *Ann New-York Acad Sc* 2008, **1149**:255-258.
13. Valiente Moro C, De Luna CJ, Tod A, Guy JH, Sparagano OAE, Zenner L: **The poultry red mite (*Dermanyssus gallinae*): a potential vector of pathogenic agents.** *Exp Appl Acarol* 2009, **48**:93-104.
14. Valiente-Moro C, Chauve C, Zenner L: **Vectorial role of some dermanyssoid mites (Acari, Mesostigmata, Dermanyssidae).** *Parasite* 2005, **12**:99-109.
15. Nordenfors H, Chirico J: **Evaluation of a sampling trap for *Dermanyssus gallinae*.** *J Vet Entomol* 2001, **94**:1617-1621.
16. Lundh J, Wiktelius D, Chirico J: **Azadirachtin-impregnated traps for the control of *Dermanyssus gallinae*.** *Vet Parasitol* 2005, **130**:337-342.
17. Siegmann O, Neumann U: **Lehrbuch Kompendium der Geflügelkrankheiten.** Schlütersche, Hannover, 6, aktualisierte erweiterte Auflage; 2005, 314-316.
18. Meyer-Kühling B, Pfister K, Müller-Lindloff J, Heine J: **Field efficacy of phoxim 50% (ByeMite) against the poultry red mite *Dermanyssus gallinae* in battery cages stocked with laying hens.** *Vet Parasitol* 2007, **147**:289-296.
19. George D, Guy J, Appleby WGC, Knox A, Shiel RS: ***In vitro* and *in vivo* acaricidal activity and residual toxicity of spinosad to the poultry red mite, *Dermanyssus gallinae*.** *Vet Parasitol* 2010, **173**:307-3161.
20. Lin C, Tsen H: **Use of two 16S DNA targeted oligonucleotides as PCR primers for the specific detection of *Salmonella* in foods.** *J Appl Bacteriol* 1996, **80**:659-666.
21. Desloire S, Valiente-Moro C, Chauve C, Zenner L: **Comparison of four methods of extracting DNA from *D. gallinae* (Acari: Dermanyssidae).** *Vet Res* 2006, **37**:725-732.
22. Valiente Moro C, Chauve C, Zenner L: **Experimental infection of *Salmonella enteritidis* by the poultry red mite, *Dermanyssus gallinae*.** *Vet Parasitol* 2007, **146**:329-336.

doi:10.1186/1756-3305-4-136

Cite this article as: Hamidi et al: *Dermanyssus gallinae* in layer farms in Kosovo: A high risk for *Salmonella* prevalence. *Parasites & Vectors* 2011 **4**:136.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

