

RESEARCH

Open Access

Free-living ciliates as potential reservoirs for eukaryotic parasites: occurrence of a trypanosomatid in the macronucleus of *Euplotes encysticus*

Sergei I Fokin^{1,2}, Martina Schrällhammer^{3,4*}, Carolina Chiellini^{1,5}, Franco Verni¹ and Giulio Petroni¹

Abstract

Background: Flagellates of the family Trypanosomatidae are obligate endoparasites, which can be found in various hosts. Several genera infect insects and occur as monoxenous parasites especially in representatives of Diptera and Hemiptera. These trypanosomatid flagellates probably share the worldwide distribution of their hosts, which are often infested by large numbers of endoparasites. Traditionally, their taxonomy was based on morphology, host origin, and life cycle. Here we report the characterization of a trypanosomatid infection detected in a protozoan, a ciliate collected from a polluted freshwater pond in a suburb of New Delhi (India).

Methods: Live observations and morphological studies applying light, fluorescence and transmission electron microscopy were conducted. Molecular analyses of host and parasite were performed and used for phylogenetic reconstructions and species (host) or genus level (parasite) identification.

Results: Although the morphological characteristics were not revealing, a high similarity of the trypanosomatids 18S rRNA gene sequence to *Herpetomonas ztiplika* and *Herpetomonas trimorpha* (Kinetoplastida, Trypanosomatidae), both parasites of biting midges (*Culicoides kibunensis* and *Culicoides truncorum*, respectively) allowed the assignment to this genus. The majority of the host population displayed a heavy infection that significantly affected the shape of the host macronucleus, which was the main site of parasite localization. In addition, the growth rate of host cultures, identified as *Euplotes encysticus* according to cell morphology and 18S rRNA gene sequence, was severely impacted by the infection.

Conclusions: The host-parasite system described here represents a recent example of free-living protists acting as environmental reservoirs for parasitic eukaryotic microorganisms.

Keywords: Ciliophora, *Euplotes encysticus*, Endosymbiont, *Herpetomonas*, Host range, *Leptomonas*, Macronucleus, Parasite, *Phytomonas*, Protozoa

Background

Kinetoplastid flagellates belonging to the family Trypanosomatidae are obligate endoparasites of different hosts including vertebrates (*Trypanosoma*, *Leishmania*, *Endotrypanum*), plants (*Phytomonas*) and invertebrates [1]. Several genera are frequently found as monoxenous

parasites of insects and are especially abundant in members of the orders Diptera and Hemiptera [2-5]. They can be found in high prevalence and apparently share the worldwide distribution of their hosts [4]. Traditionally, their taxonomy was based on morphology, host origin, and life cycle: criteria that were often found to be unreliable. Some trypanosomatid flagellates were isolated from different host species and other hosts were found harboring more than a single species [3,6,7]. These data may suggest a not very explicit host-specificity together with a high physiological plasticity, which could facilitate

* Correspondence: martina.schraellhammer@biologie.uni-freiburg.de

³Microbiology, Institute of Biology II, University of Freiburg, Schänzlestraße 1, Freiburg 79104, Germany

⁴Institute of Hydrobiology, Technische Universität Dresden, Zellescher Weg 40, Dresden 01217, Germany

Full list of author information is available at the end of the article

the establishment of new host-parasite systems [2,8]. Furthermore, it was shown that some of these parasites can exhibit extremely diverse morphologies which may even differ depending on the growth environment, i.e. host or culture medium [1,8,9]. Molecular phylogenetics, based on the 18S rRNA, glycosomal glyceraldehyde phosphate dehydrogenase (gGAPDH) and spliced leader (SL) RNA genes are nowadays considered the method of choice to assess taxonomy and phylogenetic relationships of kinetoplastid flagellates [3-5,10-12].

Herpetomonas was one of the first taxonomic divisions within the trypanosomatid flagellates (after *Angomonas* and *Strigomonas* [12]) which was redescribed following a combined approach including molecular and microscopical analyses [5]. Also new genera of insect trypanosomatids, i.e. *Sergeia* and *Blechomonas*, have been established based on 18S rRNA and gGAPDH phylogenetic analyses complemented by description of morphological features [10,13].

The genus *Herpetomonas* was previously considered to be polyphyletic. After careful revision, it now represents a monophyletic assemblage comprising 13 species, some of which exhibit extreme morphological polymorphisms. In general, *Herpetomonas* are found as parasites of dipterans, but they have been reported also from predator hemipterans [14], plants [15,16], rats [6], and immunocompromised humans [17].

Transmission pathways of monoxenous insect trypanosomatids to other hosts are still unclear. Hemipterans could acquire the parasites by predation and cannibalism. Possible additional transmission routes are coprophagy and necrophagy [4]. Isolates from plants probably derive either from infection or contamination by insect feces.

There are only a few reports of free-living protists hosting eukaryotic microorganisms evolutionary closely related to pathogens [18-24]. So far, the involved endoparasites have not been molecularly characterized except for the microsporidian infecting *Euplotes woodruffi* [19]. Thus, according to our best knowledge, this characterization of a trypanosomatid flagellate hosted by a ciliated protist belonging to the genus *Euplotes* (Ciliophora, Hypotrichia) is the first molecular description of a member of Trypanosomatidae infecting another protist. Both host and parasite were studied on a morphological and molecular level.

Methods

Ciliate isolation and cultivation

Euplotes population Ind3 was collected from a heavily eutrophicated freshwater pond near the Yamuna River in the south-east suburb of New Delhi, India (N 28°56'/E 77°29'), in February 2007. Most of the cells harbored an unknown trypanosomatid flagellate localized in the macronucleus (Ma). About 100 cells of infected *Euplotes*

sp. were collected and grown as a "laboratory population" in lettuce medium [25] inoculated with *Enterobacter aerogenes*. The establishment of monoclonal cultures from this population failed. Thus, all subsequent investigations were carried out with cells from the laboratory population.

Light microscopy

Living observations of both host cells and trypanosomatid symbionts were carried out using an Orthoplan Leitz microscope (Hexagon Metrology GmbH Leitz Division, Wetzlar, Germany), equipped with differential interference contrast (DIC), and a Leica DMR microscope (Leica Microsystems GmbH, Wetzlar, Germany). The latter was also used for fluorescence microscopy. Pictures were taken with a Leica DC200 camera.

Fixation, staining, and electron microscopy

Feulgen staining after fixation in Bouin's solution [26] was used to reveal the nuclear apparatus of infected *Euplotes* sp. and its trypanosomatid parasites. The flagellate infection in living ciliate cells was observed using ethidium bromide (0.005%) staining followed by fluorescence microscopy. Silver nitrate impregnation for host identification was performed after fixation with Champy's solution [27]. For electron microscopy, infected cells were processed as described elsewhere [28].

DNA extraction, amplification of 18S rRNA genes and sequencing

Total DNA extraction was carried out from approximately 50 host cells with NucleoSpin™ Plant DNA Extraction Kit (Macherey-Nagel GmbH & Co., Düren, Germany), following the protocol for mycelium. The almost full-length host and parasite 18S rRNA genes were amplified using specific primer combinations (Additional file 1: Table S1) and Ex Taq PCR reagents (Takara Bio Inc., Otsu, Japan) according to the manufacturer's instructions.

Sequence availability and phylogenetic analysis

Characterized 18S rRNA gene sequences are available from the EMBL European Nucleotide Archive (the host population Ind3 [EMBL: HG425175] and its trypanosomatid parasite Ind3 [EMBL: HG425174]).

Phylogenetic analyses were carried out on a subset of 26 *Herpetomonas* (according to [5]) and *Phytomonas* sequences, aligned with the sequence aligner included in the ARB program package [29]. A filter was generated on the used selection to trim them to the same length and retain only positions conserved in at least 50% of the selected sequences. The resulting alignment comprised 1,904 columns.

Tree reconstructions were performed applying Maximum likelihood (PHYML program [30] included in the ARB package) and Bayesian Inference (MrBayes [31])

analysis. For the latter, three different Markov Chain Monte Carlo runs with one cold and three heated chains were performed, running for 1,000,000 generations. The stability of the Maximum likelihood tree was warranted by bootstrap analysis (1,000 pseudoreplicates). A similarity matrix was calculated in ARB.

In addition, the host sequence was aligned in ARB and a similarity matrix of the closest related *Euplotes* sequences was computed.

Results

Host identification and effects of parasite infection

The host population was identified as *E. encysticus*, a ciliate morphospecies occurring in freshwater bodies and so far described only from Japan and China [32-34] by classical taxonomic methods of nuclear and cortex preparations (Figure 1. 1-6). According to our best knowledge, this work represents the first report of *E. encysticus* from the Indian subcontinent [35-37]. Phylogenetic analysis supported this morphological identification. The 18S rRNA gene of the Ind3 population showed 100% sequence identity with that of *E. encysticus* EF535728 [34], and 99.94% with *E. encysticus* FJ346569 [38].

The presence of the parasite apparently impaired the growth rate of the host. Attempts to establish monoclonal lines of the infected ciliate failed. Isolated host cells died after a maximum of two divisions. The established laboratory population of *E. encysticus* became extinct

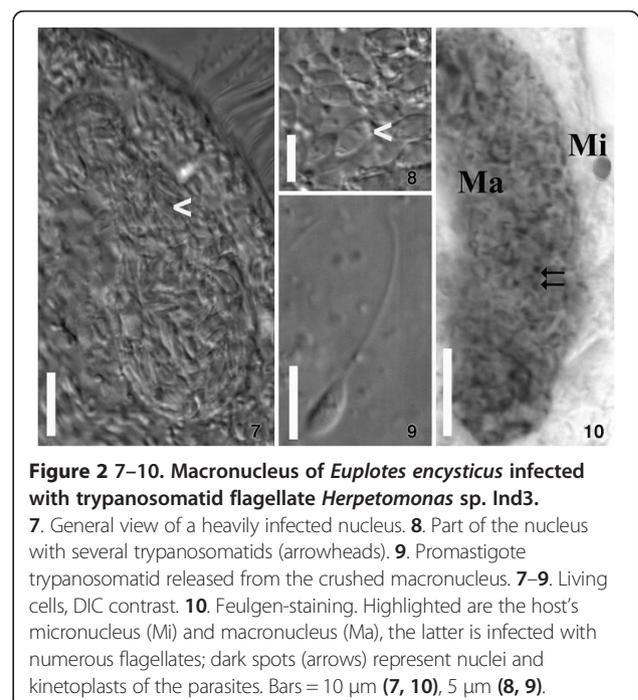
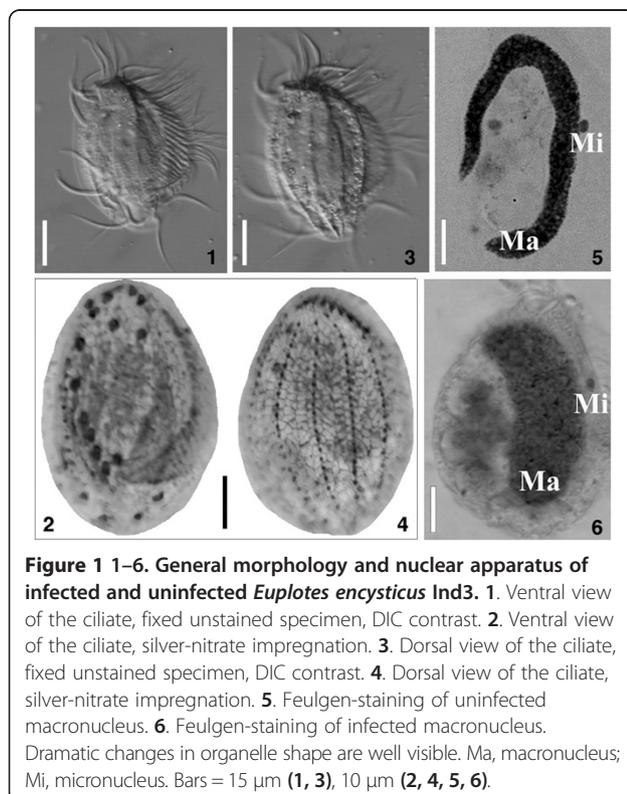
after two weeks of investigation. *Euplotes aediculatus* and members of the *Paramecium aurelia* species complex, additionally present in the original water sample, were not infected with the trypanosomatid parasite according to the controlled subset of specimens (for each group: 10 cells examined as living specimen and additional 10-15 cells observed after Feulgen staining).

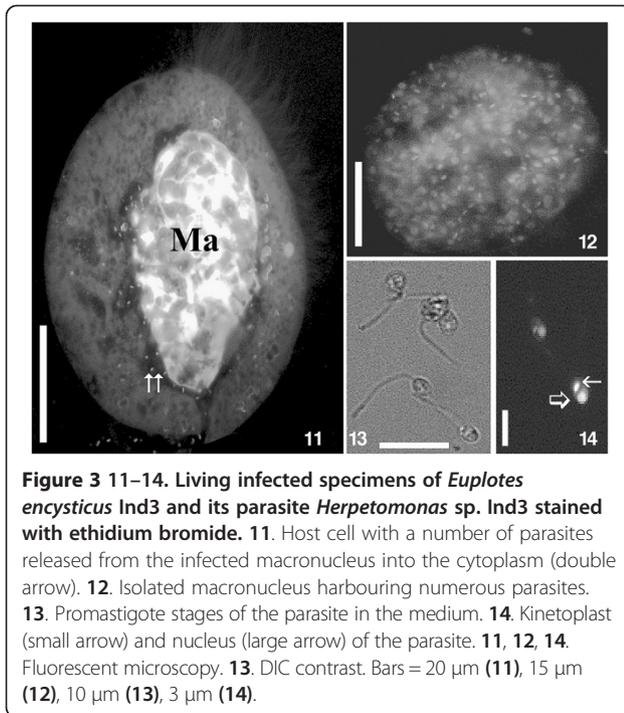
E. encysticus revealed an infection prevalence of 90% in the studied laboratory population. All infected cells featured macronuclei of unusual shape and size. Instead of the normally occurring relatively slim and C-shaped organelle, the macronucleus of parasitized cells formed a large oval body occupying the main part of the protozoan cytoplasm (Figures 1. 6, 2. 7-10, 3. 11). Each infected macronucleus harbored roughly more than one hundred flagellates (Figures 2. 7-10, 3. 12). In consequence of the infection level, only a small amount of macronuclear chromatin could be observed in the Feulgen stained ciliates (Figures 1. 6, 2. 10).

Inside the host organelle, the flagellates were immotile. After rupture of the macronucleus several parasites started to move vigorously in the culture medium. In a few host cells, low numbers (up to 10-15) of parasites were recorded in cytoplasmic vacuoles (Figure 4. 15-18), some of those were observed after recently completed cell division (Figure 4. 17).

Morphological and molecular characterization of the trypanosomatid flagellate

Promastigote forms of the trypanosomatid inside the host cell were characterized by a slightly elongated body





(Table 1 and Figures 2, 9, 3, 13, 4, 15). The flagellum originated near the kinetoplast and emerged from the flagellar pocket at the anterior end (Figures 2, 9, 3, 13, 4, 15-19). It was supported by a paraflagellar rod which could be seen in a cross-sectioned flagellum even before it exits the flagellar pocket (Figure 4, 16). The oval nucleus, which was over proportionally large, was situated at the posterior part of the cell, always close to the

kinetoplast (Table 1, Figure 4, 14, 15, 17-19). The latter, classically disc-shaped, was located next to the bottom of a deep flagellar pocket. It may occupy a significant part of the cell body (Figure 4, 15, 18, 19) and reached a thickness of $0.30 \pm 0.05 \mu\text{m}$ (Table 1, Figure 4, 15, 18, 19). The flagellate's plasmalemma was underlaid by subpellicular microtubules with regular spacing (Figure 4, 15, 16). No bacterial endosymbionts were found in the cytoplasm of investigated trypanosomatids.

The phylogenetic reconstructions presented here are in accordance with the recent redescription of the genus *Herpetomonas* [5]. All 13 *Herpetomonas* species were recovered with some minor differences in the branching order of the computed tree (Figure 5, 20) in respect to published results [5].

A similarity value (Additional file 2: Table S2) of 99.42% and 99.84%, respectively, is shared among the trypanosomatid 18S rRNA gene sequence described here and *Herpetomonas ztiplika* AY308760 [8] and *Herpetomonas trimorpha* EU179326 [9]. All three sequences formed a highly stable association (Figure 5, 20), but the affiliations within the group are less well supported.

Discussion

The role of free-living protists as reservoir hosts or even vectors for pathogens has been underestimated for a long time. Their significance in the genesis of several bacterial pathologies has been re-evaluated after disease outbreaks caused by protist-born microorganisms, i.e. *Legionella pneumophila*, the etiological agent of the Legionnaires' disease, and other amoebae-hosted pathogens [39,40]. In the last decade, many associations between

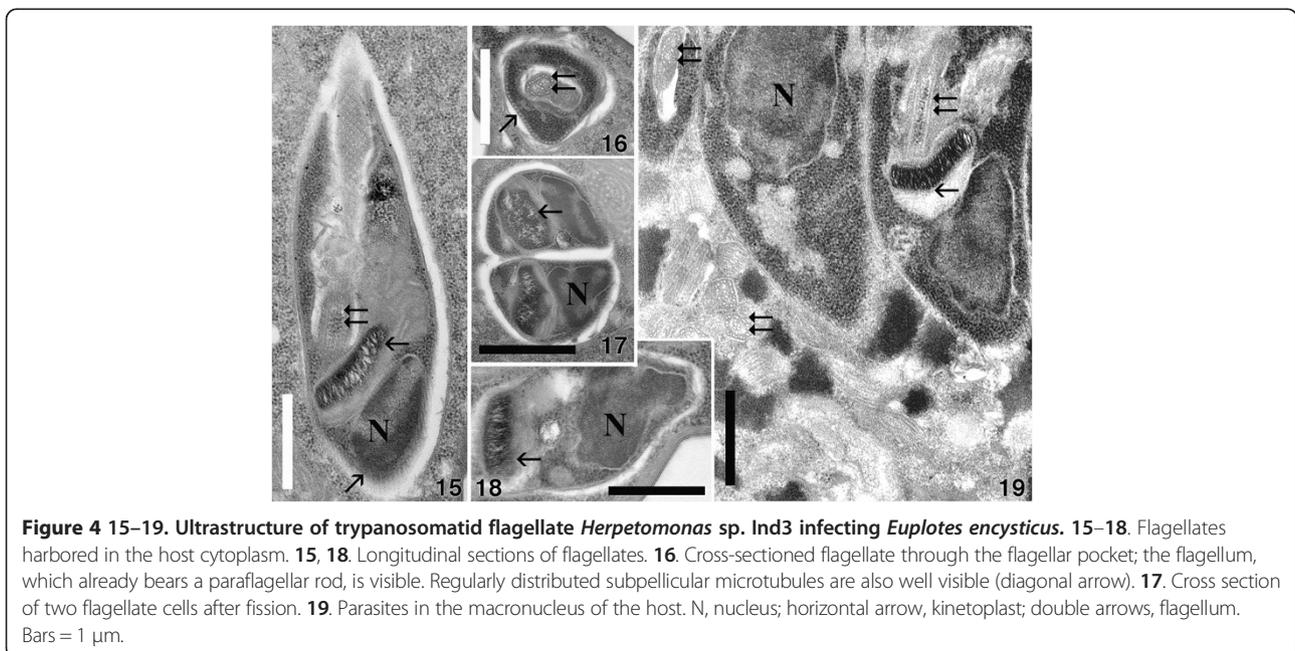


Table 1 Morphological features and localization within the host organism for some *Herpetomonas* and *Leptomonas* species

Feature	<i>Herpetomonas</i> sp. Ind3	<i>H. ztiplika</i> ^a	<i>H. trimorpha</i> ^a	<i>Leptomonas</i> sp.	<i>L. ciliatorum</i>
Host species	<i>Euplotes encysticus</i>	<i>Culicoides kibunensis</i> , <i>Culicoides truncorum</i>	<i>Culicoides truncorum</i>	<i>Euplotes</i> sp.	<i>Paraholosticha sterkii</i>
Location	Mainly macronucleus	Hindgut and Malpighian tubules	Malpighian tubules	Only macronucleus	Only macronucleus
Cell length	4.0 ± 0.2	8.9 ± 0.4	8.2 ± 0.3	5	6-8
Cell width	1.3 ± 0.7	2.1 ± 0.1	2.7 ± 1.2	1.7	2-3
Flagellum length	10.2 ± 0.8	10.7 ± 1.1	10.2 ± 2.3	12	10-14
Kinetoplast width	0.30 ± 0.05	0.16 ± 0.07 ^b	0.125 ± 0.025 ^b	0.18 + 0.0 ^c	0.4 ^c
Kinetoplast position	Closer to posterior end	Closer to anterior end ^b	Anterior end (mic p, l p) ^b ; central (s p) ^b ;	Not reported	Nearly central
Nucleus position	Closer to posterior end	Nearly central ^b	Anterior end (mic p) ^b ; central (s p, l p) ^c ;	Not reported	Close to posterior
Kinetoplast/nucleus position	Close each other	With some distance ^b	Close (mic p, s p) ^b ; distant (l p) ^b ;	Close each other	Close each other
Posterior end shape	Rounded	Fine pointed ^b	Pointed (mic p) ^b	Rounded	Slightly conical
References	Present study	[8,13]	[9]	[21]	[22]

All measurements given in µm; ^adata referred to *Herpetomonas* spp. harboured by insect hosts; ^bdata only available from cells in culture medium; ^cdata inferred from published figures; microflagellate (mic), small (s) and long (l) promastigote (p).

free-living protists and bacteria phylogenetically closely related to human or animal pathogens have been described. The most compelling examples refer to the families *Francisellaceae* and *Rickettsiaceae*. A novel *Francisella* subspecies harbored by a ciliate [41] and many “basal rickettsiae”

from amoebae [42], several ciliates [43-46] and green algae [47] have been described.

Despite the common capabilities of many free-living protists to host other eukaryotic microorganisms like many different kinds of eukaryotic “algae” [48-50], there

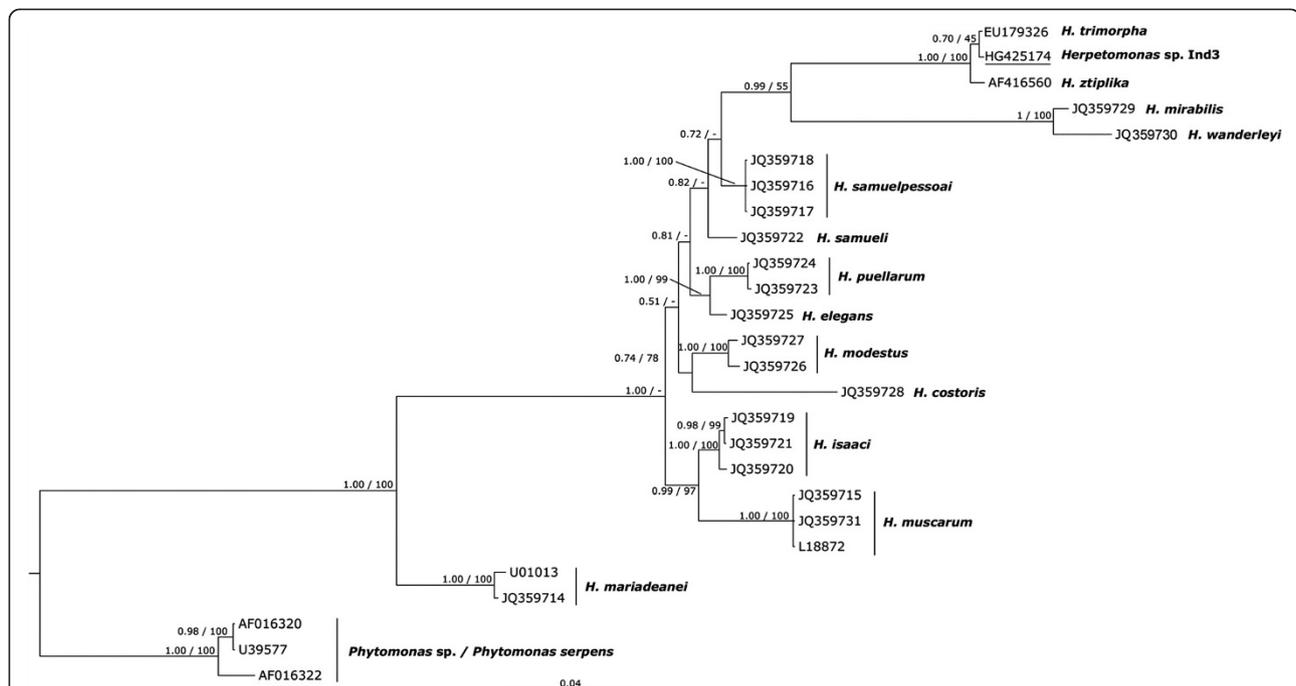


Figure 5 20. Bayesian tree of members of *Herpetomonas* and *Phytomonas*. The tree was built on the unmodified character matrix of 23 *Herpetomonas* and 3 *Phytomonas* sequences (1,904 columns) employing the GTR + I + G model. Numbers associated to each node correspond to posterior probability and Maximum Likelihood bootstrap values. The scale bar represents an estimated sequence divergence of 4%. Underlined is the sequence of the trypanosomatid flagellate of *Euplotes encysticus* Ind3.

are only a few reports of free-living ciliates hosting eukaryotic microorganisms belonging to evolutionary lineages that comprehend also human and/or animal pathogens. These include the microsporidians *Ciliatosporidium platyophryae* in *Platyophrya terricola* [18], and *Euplotespora binucleata* in *Euplotes woodruffi* [19], as well as different trypanosomatid species described in the macronuclei of *Paramecium trichium* (the younger synonym of *Paramecium putrinum*) [20], *Euplotes* sp. [21] and *Paraholosticha sterkii* [22].

None of these organisms have been characterized at the molecular level with the exception of *Euplotespora binucleata* [19]. These neglected reports [18-22] and the characterization presented here suggest that at least some free-living protists may act as a reservoir for parasitic ones. The possible capability of the latter to also parasitize higher organisms is a matter that, in our opinion, should deserve further attention. In this context, the extremely limited 18S rRNA distance (less than 0.58%) among *Herpetomonas* strains retrieved from midges, i.e. *H. ztiplika* and *H. trimorpha*, and the one characterized in the present report is particularly intriguing. Various transmission routes (e.g. predation, cannibalism, coprophagy and fecal contamination) for monoxenous insect trypanosomatids have been reported [4]. The general mechanism is based on uptake of infected organisms or material. It remains unclear how ciliates can acquire an infection but a plausible scenario involves the aquatic larval stage of infected insect hosts. The host flies of *H. ztiplika* and *H. trimorpha* (family Ceratopogonidae) comprise a semi-aquatic life style with their larval stages developing in bodies of freshwater. In these habitats also free living protozoans including ciliates do occur. Thus, an encounter of e.g. *Euplotes* with infected larvae is possible. Like many other ciliates, *Euplotes* is a filter feeder and might haphazardly ingest infected organic material or released trypanosomatid flagellates.

It has already been shown that the existence of extensive size and shape polymorphism within a single trypanosomatid species according to cultivation conditions strongly impairs the resolution power of those characters for identification and taxonomy [1,8,9,15]. The consequential conflict between traditional taxonomy and molecular phylogeny is well known also for the genus *Herpetomonas* and resulted in the recent redescription of this taxon, based mainly on molecular data [5]. For the affiliation of the newly found trypanosomatid from *E. encysticus*, application of morphological features was not feasible. According to morphology, the novel trypanosomatid shares similarities with some leptomonads described from other ciliates (size and shape of promastigote, kinetoplast width and position; Table 1) rather than with members of *Herpetomonas*. In particular *H. ztiplika* or *H. trimorpha*, which are its nearest relatives

according to molecular analyses (Figure 5. 20), share few morphological characteristics with the trypanosomatid described here (Table 1).

In contrast to morphology, molecular data demonstrate a strong affiliation of the newly characterized trypanosomatid to the genus *Herpetomonas* and, in particular, to *H. trimorpha* and *H. ztiplika* species; indeed the three organisms show an 18S rRNA gene sequence identity higher than 99.4% among themselves. Nevertheless, as far as no additional phylogenetic marker e.g. the gGAPDH gene sequence was obtained, an unambiguous affiliation to one of the mentioned *Herpetomonas* species or the establishment of a new species is not feasible.

Interestingly, the isolates of *H. ztiplika* [8] and *H. trimorpha* have the same geographic origin. Both have been recovered from biting midges collected in the Milovický forest in the vicinity of Mikulov, Czech Republic, whereas the host of the endoparasite described in this study was collected in a suburb of New Delhi, India. This observation adds to other reports of cosmopolitan trypanosomatid species [1,4,5,7] and the limited value of geographic and host origin for trypanosomatid and especially *Herpetomonas* taxonomy.

Morphological similarities between the newly described *Herpetomonas* sp. Ind3 and previously described ciliate-born leptomonads can be explained either as adaptation to similar host organisms (ciliated protist) and cell compartments (macronucleus) or to the misclassification of previously described ciliate-born trypanosomatids; only the molecular characterization of other trypanosomatids from ciliates will help to clarify this point.

Considering the present and previously reported cases, it is quite possible that some of the ciliates (eventually with the exception of *Paraholosticha sterkii*) are not the main host for the flagellates. This view is supported by the close phylogenetic relationship of the here described *Herpetomonas* sp. Ind3 to monoxenous parasites of dipterans and the severely impaired growth rate of infected *E. encysticus* Ind3.

Conclusions

Based on the results presented here we demonstrate that free-living protists can act at least as temporary environmental reservoirs for parasitic eukaryotic microorganisms. The capacity (or susceptibility) of ciliates for infections with "arthropod-specific" parasites has recently been reported for members of the obligate intracellular *Rickettsiaceae* (*Alphaproteobacteria*) [51-53]. Especially noteworthy is the bacterium '*Candidatus* Megaira polyxenophila' [43] which has been frequently found in a broad range of diverse protist hosts.

Extensive screening efforts revealed that at least some trypanosomatid species may infect a wide range of hosts,

in some cases even spanning different insect orders [6], which lead to the abolishment of the “one host – one parasite” paradigm. So far, trypanosomatid flagellates have been regarded as typical dipteran and hemipteran parasites. This pattern might change when more infected ciliates are described in the future.

It has been stated that trypanosomatid flagellates exhibit a low host specificity which might be caused by a high physiological plasticity and allow the establishment of new host-parasite systems [2,8]. In the case of *Herpetomonas*, this plasticity apparently allows a host range encompassing not only of different insect orders but also unicellular host organisms. Furthermore, it permits an endoparasitic as well as an intracellular, intramacronuclear lifestyle.

Additional files

Additional file 1: Table S1. Oligonucleotides used in the present work.

Additional file 2: Table S2. 18S rRNA similarity matrix of *Herpetomonas* species.

Competing interests

The authors declare that they have no competing interests.

Author's contributions

SIF and GP designed the study; sampling and isolation were carried out by SIF as well as the microscopical analysis, VF contributed to the microscopical analysis; primer design was done by GP; MS performed the phylogenetic reconstructions; SIF, MS and CC conducted the literature review and drafted the manuscript. All authors read and approved the final manuscript.

Acknowledgements

The authors wish to thank two anonymous reviewers for their constructive suggestions. This research was supported by the grant protocol 2012A4F828_002 from the Italian Ministry of University and Research (Ministero Italiano dell'Università e della Ricerca), the European FP7 program IRSES (CINAR-PATHOBACTER 247658), and by the BMBS COST Action BM1102. The article processing charge was funded by the German Research Foundation (DFG) and the University of Freiburg in the funding programme Open Access Publishing. S. Gabrielli is gratefully acknowledged for his technical assistance in photographic artwork, Dr. F. Ferrantini for his assistance in PCR and Dr. R. Gupta (University of Delhi) for her kind help in the collection of samples.

Author details

¹Department of Biology, Protistology-Zoology Unit, University of Pisa, Via A. Volta 4/6, Pisa 56126, Italy. ²Department of Invertebrate Zoology, St. Petersburg State University, Universitetskaya emb. 7/9, St. Petersburg 198034, Russia. ³Microbiology, Institute of Biology II, University of Freiburg, Schänzlestraße 1, Freiburg 79104, Germany. ⁴Institute of Hydrobiology, Technische Universität Dresden, Zellescher Weg 40, Dresden 01217, Germany. ⁵Consiglio per la Ricerca e la Sperimentazione in Agricoltura, Centro di Ricerca per l'Agrobiologia e Pedologia, Piazza D'Azeglio 30, Florence 50121, Italy.

Received: 26 November 2013 Accepted: 13 April 2014

Published: 28 April 2014

References

- Maslov DA, Votýpka J, Yurchenko V, Lukeš J: Diversity and phylogeny of insect trypanosomatids: all that is hidden shall be revealed. *Trends Parasitol* 2013, **29**:43–52.
- Podlipaev SA: Insect trypanosomatids: the need to know more. *Mem Inst Oswaldo Cruz* 2000, **95**:517–522.
- Votýpka J, Maslov DA, Yurchenko V, Jirků M, Kment P, Lun Z-R, Lukeš J: Probing into the diversity of trypanosomatid flagellates parasitizing insect hosts in South-West China reveals both endemism and global dispersal. *Mol Phylogenet Evol* 2010, **54**:243–253.
- Týč J, Votýpka J, Klepetková H, Suláková H, Jirků M, Lukeš J: Growing diversity of trypanosomatid parasites of flies (Diptera: Brachycera): Frequent cosmopolitanism and moderate host specificity. *Mol Phylogenet Evol* 2013, **69**:255–264.
- Borghesan TC, Ferreira RC, Takata CSA, Campaner M, Borda CC, Paiva F, Milder RV, Teixeira MMG, Camargo EP: Molecular phylogenetic redefinition of *Herpetomonas* (Kinetoplastea, Trypanosomatidae), a genus of insect parasites associated with flies. *Protist* 2013, **164**:129–152.
- Podlipaev SA, Sturm NR, Fiala I, Fernandes O, Westenberger S, Dollet M, Campbell D, Lukes J: Diversity of insect trypanosomatids assessed from the spliced leader RNA and 5S rRNA genes and intergenic regions. *J Eukaryot Microbiol* 2004, **51**:283–290.
- Votýpka J, Klepetková H, Yurchenko VY, Horák A, Lukeš J, Maslov DA: Cosmopolitan distribution of a trypanosomatid *Leptomonas pyrrocoris*. *Protist* 2012, **163**:616–631.
- Podlipaev SA, Votýpka J, Jirků M, Svobodová M, Lukes J: *Herpetomonas ztiplika* n. sp. (Kinetoplastida: Trypanosomatidae): a parasite of the blood-sucking biting midge *Culicoides kibunensis* Tokunaga, 1937 (Diptera: Ceratopogonidae). *J Parasitol* 2004, **90**:342–347.
- Zidková L, Cepicka I, Votýpka J, Svobodová M: *Herpetomonas trimorpha* sp. nov. (Trypanosomatidae, Kinetoplastida), a parasite of the biting midge *Culicoides truncorum* (Ceratopogonidae, Diptera). *Int J Syst Evol Microbiol* 2010, **60**:2236–2246.
- Votýpka J, Suková E, Kraeva N, Ishemgulova A, Du` I, Yurchenko V, Luke J: Diversity of Trypanosomatids (Kinetoplastea: Trypanosomatidae) parasitizing fleas (Insecta: Siphonaptera) and description of a new genus *Blechomonas* gen. n. *Proti* 2013, **164**:763–781.
- Hamilton PB, Stevens JR, Gidley J, Holz P, Gibson WC: A new lineage of trypanosomes from Australian vertebrates and terrestrial bloodsucking leeches (Haemadipsidae). *Int J Parasitol* 2005, **35**:431–443.
- Teixeira MMG, Borghesan TC, Ferreira RC, Santos MA, Takata CSA, Campaner M, Nunes VLB, Milder RV, de Souza W, Camargo EP: Phylogenetic validation of the genera *Angomonas* and *Strigomonas* of trypanosomatids harboring bacterial endosymbionts with the description of new species of trypanosomatids and of proteobacterial symbionts. *Protist* 2011, **162**:503–524.
- Svobodová M, Zidková L, Cepicka I, Oborník M, Lukes J, Votýpka J: *Sergeia podlipaevi* gen. nov., sp. nov. (Trypanosomatidae, Kinetoplastida), a parasite of biting midges (Ceratopogonidae, Diptera). *Int J Syst Evol Microbiol* 2007, **57**(Pt 2):423–432.
- Roitman I, Brener Z, Roitman C, Kitajima E: Demonstration that *Leptomonas pessoai* Galvão, Oliveira, Carvalho & Veiga, 1970, is a *Herpetomonas*. *J Protozool* 1976, **23**:291–293.
- Fiorini J, Takata C, Teofilo V, Nascimento L, Faria-e-Silva P, Soares M, Teixeira M, De Souza W: Morphological, biochemical and molecular characterization of *Herpetomonas samuelpeessoai* n. subsp., a trypanosomatid isolated from the flower of the squash *Cucurbita moschata*. *J Eukaryot Microbiol* 2001, **48**:62–69.
- Marín C, Fabre S, Sánchez-Moreno M, Dollet M: *Herpetomonas* spp. isolated from tomato fruits (*Lycopersicon esculentum*) in southern Spain. *Exp Parasitol* 2007, **116**:88–90.
- Morio F, Reynes J, Dollet M, Pratloug F, Dedet J-P, Ravel C: Isolation of a protozoan parasite genetically related to the insect trypanosomatid *Herpetomonas samuelpeessoai* from a human immunodeficiency virus-positive patient. *J Clin Microbiol* 2008, **46**:3845–3847.
- Foissner I, Foissner W: *Ciliatosporidium platyphryae* nov. gen., nov. spec. (*Micropora incerta sedis*), a parasite of *Platyphrya terricola* (Ciliophora, Colpodea). *Eur J Protistol* 1995, **31**:248–259.
- Fokin SI, Di Giuseppe G, Erra F, Dini F: *Euplotespora binucleata* n. gen., n. sp. (Protozoa: Microsporidia), a parasite infecting the hypotrichous ciliate *Euplotes woodruffi*, with observations on microsporidian infections in ciliophora. *J Eukaryot Microbiol* 2008, **55**:214–228.
- Gillies C, Hanson ED: A new species of *Leptomonas* parasitizing the macronucleus of *Paramecium trichium*. *J Protozool* 1963, **10**:467–473.
- Wille JJ, Weidner EJ, Steffens WL: Intracellular parasitism of the ciliate *Euplotes* by a trypanosomatid flagellate. *J Protozool* 1981, **28**:223–227.

22. Görtz H-D, Dieckmann J: *Leptomonas ciliatorum* n. sp. (Kinetoplastida, Trypanosomatidae) in the macronucleus of a hypotrichous ciliate. *J Protozool* 1987, **34**:259–263.
23. Dyková I, Fiala I, Lukes J: *Perkinsiella* amoebae-like endosymbionts of *Neoparamoeba* spp., relatives of the kinetoplastid *Ichthyobodo*. *Eur J Protistol* 2003, **52**:37–52.
24. Tanifuji G, Kim E, Onodera NT, Gibeault R, Dlutek M, Cawthorn RJ, Fiala I, Lukes J, Greenwood SJ, Archibald JM: Genomic characterization of *Neoparamoeba pemaquidensis* (Amoebozoa) and its kinetoplastid endosymbiont. *Eukaryot Cell* 2011, **10**:1143–1146.
25. Wichterman R: *The Biology of Paramecium*. New York: The Blakiston Company, Inc; 1953:1–527.
26. Fokin SI: Bacterial endobionts of the ciliate *Paramecium woodruffi*. II. Endobionts of the perinuclear space. *Tsitologiya* 1989, **31**:845.
27. Corliss JO: Silver impregnation of ciliated protozoa by the Chatton-Lwoff technique. *Stain Technol* 1953, **28**:97–100.
28. Fokin SI, Görtz H-D: “*Caedibacter macronucleorum*” sp. nov., a bacterium inhabiting the macronucleus of *Paramecium duboscqui*. *Arch Protistenkd* 1993, **143**:319–324.
29. Ludwig W, Strunk O, Westram R, Richter L, Meier H, Yadhukumar, Buchner A, Lai T, Steppi S, Jobb G, Förster W, Brettske I, Gerber S, Ginhart AW, Gross O, Grumann S, Hermann S, Jost R, König A, Liss T, Lüssmann R, May M, Nonhoff B, Reichel B, Strehlow R, Stamatakis A, Stuckmann N, Vilbig A, Lenke M, Ludwig T, et al: ARB: a software environment for sequence data. *Nucleic Acids Res* 2004, **32**:1363–1371.
30. Guindon S, Gascuel O: A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. *Syst Biol* 2003, **52**:696–704.
31. Huelsenbeck JP, Ronquist F: MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics* 2001, **17**:754–755.
32. Fan X, Huang J, Lin X, Li J, Al-Rasheid KAS, Hu X: Morphological and molecular characterization of *Euplotes encysticus* (Protozoa: Ciliophora: Euplotida). *J Mar Biol Assoc United Kingdom* 2010, **90**:1411–1416.
33. Yonezawa F: New hypotrichous ciliate *Euplotes encysticus* sp. nov. *J Sci Hiroshima Univ Ser B, Div 1* 1985, **32**:35–46.
34. Li Y, Niu Y, Liu L: Phylogenetic studies of four species of ciliate inferred from 16S-like small subunit rRNA gene sequences. *J For Res* 2008, **19**:119–124.
35. Bhatia BL: Protozoa: Ciliophora. In *The fauna of British India including Ceylon and Burma*, Volume 1. Edited by Sewell RBS. London: Taylor and Francis Ltd; 1936:1–548.
36. Das AK, Mandal AK, Sarkar NC: Protozoa. 1. Free-living Protozoa. Fauna of West Bengal. In *Zoological Survey of India*. New Alipore: M-Block; 1993:1–133.
37. Kalavati C, Raman AV: Taxonomy and ecology of ciliated Protozoa from marginal marine environments of east coast of India. In *Zoological Survey of India*. New Alipore: M-Block; 2008:1–136.
38. Yi Z, Weibo S, Clamp JC, Chen Z, Gao S, Zhang Q: Reconsideration of systematic relationships within the order Euplotida (Protista, Ciliophora) using new sequences of the gene coding for small-subunit rRNA and testing the use of combined data sets to construct phylogenies of the *Diophrys*-complex. *Mol Phylogenet Evol* 2009, **50**:599–607.
39. Lamoth F, Greub G: Amoebal pathogens as emerging causal agents of pneumonia. *FEMS Microbiol Rev* 2010, **34**:260–80.
40. Thomas V, McDonnell G, Denyer SP, Maillard J-Y: Free-living amoebae and their intracellular pathogenic microorganisms: risks for water quality. *FEMS Microbiol Rev* 2010, **34**:231–59.
41. Schrällhammer M, Schweikert M, Vallesi A, Verni F, Petroni G: Detection of a novel subspecies of *Francisella noatunensis* as endosymbiont of the ciliate *Euplotes raikovi*. *Microb Ecol* 2011, **61**:455–64.
42. Dyková I, Veverková M, Fiala I, Macháčková B, Pecková H: *Nuclearia pattersoni* sp. n. (Filosea), a new species of amphizoic amoeba isolated from gills of roach (*Rutilus rutilus*), and its rickettsial endosymbiont. *Folia Parasitol (Praha)* 2003, **50**:161–170.
43. Schrällhammer M, Ferrantini F, Vannini C, Galati S, Schweikert M, Görtz H-D, Verni F, Petroni G: “*Candidatus Megaira polyxenophila*” gen. nov., sp. nov.: Considerations on evolutionary history, host range and shift of early divergent Rickettsiales. *PLoS One* 2013, **8**:e72581.
44. Vannini C, Petroni G, Verni F, Rosati G: A bacterium belonging to the *Rickettsiaceae* family inhabits the cytoplasm of the marine ciliate *Diophrys appendiculata* (Ciliophora, Hypotrichia). *Microb Ecol* 2005, **49**:434–442.
45. Sun HY, Noe J, Barber J, Coyne RS, Cassidy-Hanley D, Clark TG, Findly RC, Dickerson HW: Endosymbiotic bacteria in the parasitic ciliate *Ichthyophthirius multifiliis*. *Appl Env Microbiol* 2009, **75**:7445–7452.
46. Ferrantini F, Fokin SI, Modeo L, Andreoli I, Dini F, Görtz H-D, Verni F, Petroni G: “*Candidatus Cryptoprodotis polytropus*”, a novel *Rickettsia*-like organism in the ciliated protist *Pseudomicrothorax dubius* (Ciliophora, Nassophorea). *J Eukaryot Microbiol* 2009, **56**:119–129.
47. Kawafune K, Hongoh Y, Hamaji T, Nozaki H: Molecular identification of rickettsial endosymbionts in the non-phagotrophic volvoclean green algae. *PLoS One* 2012, **7**:e31749.
48. Summerer M, Sonntag B, Sommaruga R: Ciliate-symbiont specificity of freshwater endosymbiotic *Chlorella* (Trebouxiophyceae, Chlorophyta). *J Phycol* 2008, **44**:77–84.
49. Kodama Y, Fujishima M: Secondary symbiosis between *Paramecium* and *Chlorella* cells. In *Int Rev Cell Mol Biol. Volume 2010*, **279**:33–77.
50. Nowack ECM, Melkonian M: Endosymbiotic associations within protists. *Philos Trans R Soc Lond B Biol Sci* 2010, **365**:699–712.
51. Montagna M, Sasser D, Epis S, Bazzocchi C, Vannini C, Lo N, Sacchi L, Fukatsu T, Petroni G, Bandi C: “*Candidatus Midichloriaceae*” fam. nov. (*Rickettsiales*), an ecologically widespread clade of intracellular Alphaproteobacteria. *Appl Entomol Zool* 2013, **79**:3241–3248.
52. Boscaro V, Petroni G, Ristori A, Verni F, Vannini C: “*Candidatus Defluviella procrastinata*” and “*Candidatus Cyrtobacter zanobii*”, two novel ciliate endosymbionts belonging to the “*Midichloria* clade”. *Microb Ecol* 2013, **65**:302–310.
53. Thepparit C, Sunyakumthorn P, Guillotte ML, Popov VL, Foil LD, Macaluso KR: Isolation of a rickettsial pathogen from a non-hematophagous arthropod. *PLoS One* 2011, **6**:e16396.

doi:10.1186/1756-3305-7-203

Cite this article as: Fokin et al.: Free-living ciliates as potential reservoirs for eukaryotic parasites: occurrence of a trypanosomatid in the macronucleus of *Euplotes encysticus*. *Parasites & Vectors* 2014 **7**:203.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

