RESEARCH Open Access

Synthesis of oxadiazole-2-oxide derivatives as potential drug candidates for schistosomiasis targeting SiTGR

Gongming Li^{1,3}, Qingqing Guo^{2,3†}, Chao Feng^{1,3}, Huan Chen¹, Wenjiao Zhao^{1,3}, Shu Li³, Yang Hong^{2*} and Degun Sun^{1*}

Abstract

Background: Schistosomiasis is a chronic parasitic disease that affects millions of people's health worldwide. Because of the increasing drug resistance to praziquantel (PZQ), which is the primary drug for schistosomiasis, developing new drugs to treat schistosomiasis is crucial. Oxadiazole-2-oxides have been identified as potential anti-schistosomiasis reagents targeting thioredoxin glutathione reductase (TGR).

Methods: In this work, one of the oxadiazole-2-oxides derivatives furoxan was used as the lead compound to exploit a series of novel furoxan derivatives for studying inhibitory activity against both recombinant *Schistosoma japonicum* TGR containing selenium (rS/TGR-Sec) and soluble worm antigen protein (SWAP) containing wild-type *Schistosoma japonicum* TGR (wtS/TGR), in order to develop a new leading compound for schistosomiasis. Thirty-nine novel derivatives were prepared to test their activity toward both enzymes. The docking method was used to detect the binding site between the active molecule and S/TGR. The structure—activity relationship (SAR) of these novel furoxan derivatives was preliminarily analyzed.

Results: It was found that several new derivatives, including compounds **6a–6d**, **9ab**, **9bd** and **9be**, demonstrated greater activity toward rS/TGR-Sec or SWAP containing wtS/TGR than did furoxan. Interestingly, all intermediates bearing hydroxy (**6a–6d**) showed excellent inhibitory activity against both enzymes. In particular, compound **6d** with trifluoromethyl on a pyridine ring was found to have much higher inhibition toward both rS/TGR-Sec (half-maximal inhibitory concentration, IC_{50} ,7.5nM) and SWAP containing wtS/TGR (IC_{50} ,55.8nM) than furoxan. Additionally, the docking method identified the possible matching sites between **6d** and *Schistosoma japonicum* TGR (S/TGR), which theoretically lends support to the inhibitory activity of **6d**.

Conclusion: The data obtained herein showed that **6d** with trifluoromethyl on a pyridine ring could be a valuable leading compound for further study.

Keywords: Anti-schistosomiasis, *Schistosoma japonicum*, S/TGR, Oxadiazole-2-oxides, Furoxan

Full list of author information is available at the end of the article

Background

Schistosomiasis is one of the most widespread parasitic diseases in the world, and is one of the main tropical endemic diseases in Asia, Africa and South America [1]. It is estimated that approximately 779 million individuals around the world are at risk of schistosomes infection [2]. Schistosomiasis is caused by five species of the trematode



© The Author(s) 2021. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

^{*}Correspondence: hongyang@shvri.ac.cn; dqsun@swust.edu.cn

[†]Gongming Li and Qingqing Guo contributed equally to this work

¹ School of Life Science and Engineering, Southwest University of Science and Technology, Mianyang 621010, China

² National Reference Laboratory of Animal Schistosomiasis, Key Laboratory of Animal Parasitology of Ministry of Agriculture, Shanghai Veterinary Research Institute, Chinese Academy of Agricultural Sciences, Shanghai 200241, China

Li et al. Parasites Vectors (2021) 14:225 Page 2 of 11

flatworm: Schistosoma japonicum (S. japonicum), Schistosoma mansoni (S. mansoni), Schistosoma haematobium, Schistosoma mekongi and Schistosoma intercalatum. When the human body is infected, the worms will persist in the liver, hepatic portal system or the urinary tract system, leading to inflammation and obstructive disease [3]. In China, schistosomiasis is caused mainly by Schistosoma japonicum, and more than 77 thousand people are infected [4].

The most effective drug for the treatment of schistosomiasis is praziquantel (PZQ) (1), which has been used extensively since the 1980s [5] in the clinic without any backup drugs. The overuse of praziquantel has caused and accelerated the emergence of drug resistance. While PZQ is very effective against the adult forms of all schistosome species, it is weak for juvenile forms of all schistosomes, which might be the reason that PZQ cannot cure schistosomiasis completely. Although R-PZQ was approved by the Food and Drug Administration (FDA) as an orphan drug in 2018, it is also at risk of resistance, and there is still no alternative drug against the migratory juvenile and subadult worms [6-9]. Thus, there is an urgent need to look for potential reagents to discover novel alternatives to PZQ for the treatment of schistosomiasis.

It has been reported [1, 10–13] that instead of separate thioredoxin reductase (TrxR) and glutathione reductase (GR) enzymes of mammalian hosts, a single multifunctional selenocysteine-containing flavoenzyme, thioredoxin glutathione reductase (TGR), plays a critical role in maintaining proper redox balance for parasite survival in both *S. mansoni* and *S. japonicum* (Fig. 1). The cellular redox system is important in the physiological function of an organism. Given the biochemical differences between the redox metabolism of schistosomes and host (mammalian), TGR is inferred to be a potential target for new drug design.

In previous studies [1, 2, 10–13], some oxadiazole-2-oxide analogues have been shown to effectively inhibit SmTGR and SjTGR, even exhibiting prominent worm killing activity in vivo. Importantly, some oxadiazole-2-oxides showed good juvenile and adult *S. japonicum* killing activity in vitro [2]. Modifications around an oxadiazole-2-oxide skeleton have been demonstrated as a valuable strategy for discovering potential antischistosomal agents. The application of bioisosteres is an effective strategy in drug design. In the design of compounds, previous studies [1, 2, 10–13] have mainly substituted on the benzene ring and changed the CN moiety, but did not pay attention to modification of the benzene ring and other moieties in the furoxan structure with corresponding bioisosteres. Therefore, in this work, the strategy of

applying bioisosteres was exploited to design novel oxadiazole-2-oxide derivatives.

On the other hand, the prokaryotic expression system has been used extensively for protein expression due to its rapid growth rate, capacity for continuous fermentation, and relatively low cost. Due to the lack of the posttranslational modifications of this expression system, the bioactivity, function, structure, solubility and so on will be affected to the expressed functional products. Although eukaryotic expression systems can resolve this problem, just like a two-edged sword, some disadvantages such as low yield, demanding culture conditions and higher cost cannot be totally avoided [14]. Thus, the special soluble worm antigen protein (SWAP) containing wtSjTGR extracted from worms might be a better alternative target for studying the bioactivity. It would reflect the actual situation in advance when Schistosoma japoni*cum* is treated with chemical compounds. Therefore, the target compounds herein were also applied to test the inhibition activity on SWAP containing wtSjTGR in addition to rSjTGR-Sec.

In this work, two series of novel furoxan derivatives (7 and 9) (Fig. 2) were obtained through two methods: either by replacing the phenyl moiety of furoxan with its bioisosteres pyridine or substituted pyridine, or by modifying the cyano moiety into an ester or amine group. Because the fluorine atom possesses a strong electron-withdrawing effect and high lipophilicity, appropriate introduction of fluorine atoms into molecules can improve their biofunction [15]. Therefore, several fluorine-containing derivatives were also intentionally synthesized. The target compounds (7 and 9) and their corresponding intermediates (6) (Fig. 2) were tested for their inhibitory activity on TrxR from rS/TGR-Sec and SWAP containing wtS/TGR.. Based on the result obtained, the SAR of these novel furoxan derivatives was preliminarily analyzed.

Methods

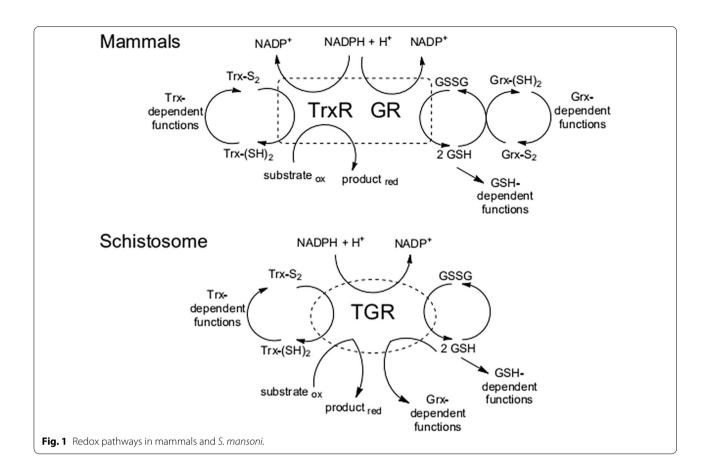
Reagent

All chemical reagents involved in synthetic procedures were purchased commercially from J&K Chemicals. The chemical reagents used in activity measurement, including dimethyl sulfoxide (DMSO), ethylene diamine tetraacetic acid (EDTA), dithionitrobenzoic acid (DTNB), and MTT, were purchased from Sigma-Aldrich Inc.

Synthesis of oxadiazole-2-oxide derivatives

The synthesis of compounds $6\mathbf{a}-\mathbf{d}$ commenced with the synthesis of α,β -unsaturated esters $(4\mathbf{a}-\mathbf{d})$ from substituted nicotinal dehydes $(3\mathbf{a}-\mathbf{d})$ by Wittig reaction [16]. Compounds $4\mathbf{a}-\mathbf{d}$ were then reduced using diisobuty-laluminum hydride (DIBAL-H) [17] to produce alcohols

Li et al. Parasites Vectors (2021) 14:225 Page 3 of 11



5a–d. Subsequent sodium nitrate-mediated cyclization [17] gave oxadiazole-2-oxides (**6a–d**) (Scheme 1).

In Scheme 2, compound 7 was readily prepared by esterification of alcohol 6a-d with various acyl chlorides using triethylamine and dimethylaminopyridine (DMAP) as base and catalyst, respectively. The synthesis of target compounds 9 began with the treatment of compounds 6a-d with benzenesulfonyl chloride to generate unstable intermediates 8a-d, which were aminated in situ to afford various target amines 9. As a result, a total of 39 novel oxadiazole-2-oxide derivatives were synthesized and their structures were determined by nuclear magnetic resonance (NMR) and high-resolution mass spectrometry (HRMS). The preparation process and spectrum of the compounds are detailed in Additional file 1.

Crystallography

Normally, two isomers **6** and **6'** were obtained (Scheme 1) and one of them was the main product after cyclization. In order to confirm the conformation of main product, its crystal was selected and was measured on a Super-Nova diffractometer (dual-source, Cu at zero, EosS2). The

crystal was kept at 150 K during data collection. Using Olex2 [18], the structure was solved with the Superflip structure solution program using charge-flipping and refined with the ShelXL refinement package using least squares minimization.

Li et al. Parasites Vectors (2021) 14:225 Page 4 of 11

Preparation of the rS/TGR-Sec and the SWAP containing wtS/TGR

The rS/TGR-Sec was expressed and harvested as described previously [13, 19]. Most of rS/TGR-Sec was expressed in *E. coli* as a soluble His-tagged fusion protein when bacterial growth occurred at 24 °C. The rS/TGR-Sec was purified by His Mag[™] Agarose beads according to manufacturer's instructions. The concentration of purified rS/TGR-Sec was determined using a Bradford assay kit (Sangon, China), and the protein was identified by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) and stored at 4 °C [20].

The SWAP containing wtS/TGR was prepared from adult schistosomes. Rabbits were infected with cercariae of *S. japonicum* by exposing their abdominal skin and were sacrificed at 42 days after infection to collect the adult schistosomes by perfusion. The worms were washed three times with phosphate-buffered saline (PBS). After sufficient grinding of the adult schistosomes in PBS, a suitable amount of benzoyl sulfonyl fluoride (PMSF) solution was added to obtain a final concentration of 1 mM. The homogenate was centrifuged at 4 °C, $12,000 \times g$ for 20 min [1, 10]. The concentration of supernatant containing wtS/TGR was determined by Bradford assay kit (Sangon, China).

Compound inhibition of rS/TGR-Sec and SWAP containing wtS/TGR

The novel furoxan derivatives in this work were dissolved in DMSO with a storage solution concentration of 64 millimoles per liter. The analysis of TrxR activity of SWAP containing wtSjTGR and rSjTGR-Sec was performed based on methods described previously [21]. Analysis of

the inhibitory activity of the compounds against the TrxR activity of SWAP containing wtSjTGR and rSjTGR-Sec described in this paper was also based on a previously described method [2]. The reaction was performed in 0.1 M potassium phosphate buffer (pH 7.0) containing 10 mM EDTA, 100 μ M nicotinamide adenine dinucleotide phosphate (NADPH), 0.2 μ g supernatant of SWAP containing wtSjTGR or 0.6 μ g rSjTGR-Sec, and the studied compounds with different concentrations at 25 °C in a final volume of 200 μ L, and was initiated by adding 3 mM DTNB. The increase of the absorbance at 412 nm was monitored, and the IC $_{50}$ values of the compounds were calculated by GraphPad Prism v6.0c software; The results are reported in Table 1. All assays were done in triplicate.

Docking study

The crystal structure of S/TGR is a homodimer, and we retained chain B for docking studies [22]. The docking was conducted by AutoDock 4, and the results are displayed in Figs. 4 and 5. The results were further processed and displayed by PyMOL2.3.2. The hydrophobic effects between inhibitors and protein were analyzed by LigPlot⁺v.2.1, and the result is shown in Fig. 5.

Results and discussion Synthesis and crystallography

In total, 39 novel oxadiazole-2-oxide derivatives were synthesized following Schemes 1 and 2. The structures of these compounds are detailed in Table 1. During synthesis of the important intermediate 6, two isomers were inevitably formed when the oxadiazole-N-oxide ring was constructed by sodium nitrate-mediated cyclization. Gasco and co-workers reported [23] that intramolecular

Li et al. Parasites Vectors (2021) 14:225 Page 5 of 11

$$R^{1}$$
:

 R^{1}
 R^{2}
 R^{3}
 R^{4}
 R^{4}
 R^{4}
 R^{4}
 R^{1} :

 R^{1}
 R^{2}
 R^{2}
 R^{3}
 R^{4}
 R^{4}

DCM, DMAP, r.t; (3) amine, K₂CO₃, Nal, MeCN, r.t. DCM, dichloromethane; DMAP, dimethylaminopyridine; r.t., room temperature.

hydrogen hand played a grueial rale on the formation of in Ta

hydrogen bond played a crucial role on the formation of the main isomer during cyclization and could stabilize the structure of the isomer a (Scheme 3).

For our case, two isomers, 6 and 6', were obtained (Scheme 1), and one of them was the main product after cyclization. To confirm the position of the N-oxide moiety, and whether the main product was 6 or 6, the crystal of the main product was obtained. It was interesting that the X-ray confirmed that the main product had a similar conformation with isomer a in Scheme 3, and specifically, the main product was 6a (Scheme 1). The crystal structure showed that there were two independent molecules in an asymmetric unit (Fig. 3a). One molecule's nitrogen atoms were labeled N1, N2 and N3, and the other molecule's nitrogen atoms were labeled N4, N5 and N6. However, intermolecular hydrogen bonds, instead of intramolecular hydrogen bonds, were found in the crystal of 6a. The molecule's nitrogen atoms labeled N1, N2 and N3 formed a hydrogen bond helix chain (Fig. 3b), while the nitrogen atoms labeled N4, N5 and N6 formed a hydrogen bond zigzag chain (Fig. 3c). This result also indicated that after the pyridine replaced the benzene, the intramolecular hydrogen bond shown in Scheme 3 might not be necessary for the formation of main isomer 6a-d.

TGR activity assessment

With furoxan as the positive control, oxadiazole-2-oxides (6, 7, 9) were evaluated for their inhibitory activity against both rSjTGR-Sec and SWAP containing wtSjTGR under different concentrations (Table 1). As shown

in Table 1, all intermediates bearing hydroxy (6a-6d) showed better inhibitory activity against both enzymes than furoxan. The pyridine ring in the structure of compound 6a had a stronger electron-withdrawing effect on the oxadiazole-2-oxide ring than compound 6b, and 6a (IC₅₀ 70nM of SWAP containing wtSjTGR, IC₅₀ 36nM of rSjTGR-Sec) had better inhibitory activity on both enzymes than 6b (IC₅₀ 0.14μM of SWAP containing wtSjTGR, IC₅₀ 0.39μM of rSjTGR-Sec). Compared with 6b, the electron-donating methoxy group at the ortho position of pyridine (like compound 6c) decreased the inhibitory activity (**6c**, IC₅₀ 1.1μM of SWAP containing wtSjTGR, IC₅₀ 2.4μM of rSjTGR-Sec). Notably, electronwithdrawing group trifluoromethyl made 6d exhibited excellent inhibitory effect (IC50 7.5nM of SWAP containing wtSjTGR, IC₅₀ 55.8nM of rSjTGR-Sec). These data indicated that the electron-withdrawing substituents very possibly provided enhanced potency.

To further investigate the relationship between the structure and bioactivity, modification of hydroxyl group into the ester group and amine, respectively, afforded compounds 7 and **9**. It was found that the ester group could not effectively improve the inhibitory effect, although the ethyl ester (7**ca**), chloracetyl ester (7**ae**, 7**be**), benzoyl ester (7**af**, 7**cf**) and acryloyl ester (7**bj**) exhibited inhibitory effects similar to furoxan. Only 7**ca** (IC₅₀ 34.77 μ M of SWAP containing wtS*j*TGR) showed inhibitory effects against SWAP containing wtS*j*TGR, but not as good as furoxan. Among chloracetyl esters, perhaps because the pyridine ring of 7**ae** had a stronger electron-withdrawing effect, 7**ae** (IC₅₀ 4.23 μ M of SWAP

Li et al. Parasites Vectors (2021) 14:225 Page 6 of 11

Table 1 Structures of oxadiazole-2-oxide derivatives and their inhibitory activity against the TrxR activity of SWAP containing wtS/TGR and rS/TGR-Sec

| Compounds | | R' | IC ₅₀ of SWAP containing wtSjTGR (μM) | IC ₅₀ of rS/TGR-Sec (μM) |
|----------------------------|---------|--|---|-------------------------------------|
| | | | | |
| CN + N-O | Furoxan | / | 4.00±0.42 | 5.87±1.37 |
| OH +N-0- N-0 | 6a | / | 0.07±0.01 | 0.036±0.002 |
| OH N-0 | 6b | / | 0.14±0.03 | 0.39±0.02 |
| OHOH | 6c | / | 1.12±0.27 | 2.38±0.42 |
| F ₃ C OH OH N-O | 6d | / | 0.00747±0.00095 | 0.05580±0.00318 |
| R' +N-O | 7aa | ~~ | >50 | >50 |
| | 7ab | ~\^ | >50 | >50 |
| | 7ac | ~~~ | >50 | >50 |
| | 7ad | | >50 | >50 |
| | 7ae | ~°Çcı | 4.23±0.54 | >50 |
| | 7af | | 8.87±0.46 | 3.23±0.05 |
| | 7ag | | >50 | >50 |
| | 7ah | F | >50 | >50 |
| | 7ai | CF ₃ | >50 | >50 |
| | 9aa | hn- | 21.57±1.44 | >50 |
| | 9ab | HN-\OCF3 | 0.62±0.03 | 1.52±0.20 |
| | 9ac | -N_N-_F | >50 | >50 |
| | 9ad | HN - OCF ₃ -N N - CF ₃ | >50 | >50 |
| R' +N-0 N-0' | 7ba | ~~ | >50 | >50 |
| | 7bb | ~ | >50 | >50 |
| | 7bc | | >50 | >50 |
| | 7bd | ~\ ~\ | >50 | >50 |

Li et al. Parasites Vectors (2021) 14:225 Page 7 of 11

Table 1 (continued)

| Compounds | | R' | IC ₅₀ of SWAP containing wtSjTGR (μM) | IC ₅₀ of rSjTGR-Sec (μM) |
|---|-----|----------------------------|---|-------------------------------------|
| | 7be | ~ 0 √ 01 | 16.40±0.67 | 14.82±3.67 |
| | 7bf | | >50 | >50 |
| | 7bg | , of F | >50 | >50 |
| | 7bh | , o , F , F | >50 | >50 |
| | 7bi | CF ₃ | >50 | >50 |
| | 7bj | ~°\ | 5.80±1.55 | >50 |
| | 7bk | , o C s | >50 | >50 |
| | 9ba | HN- | >50 | >50 |
| | 9bb | $HN \longrightarrow OCF_3$ | >50 | >50 |
| | 9bc | -N_N-_}F | >50 | >50 |
| | 9bd | -N_N-CF3 | 0.042 ± 0.01 | NA |
| | 9be | TBSQ N-Ts | 0.63±0.11 | 0.22±0.01 |
| MeO | 7ca | ~~ | 34.77±1.33 | >50 |
| | 7cf | | 12.51±0.68 | >50 |
| , | 9ca | HN- | >50 | >50 |
| F ₃ C N R' + N-O N-O | 7da | ~~ | >50 | >50 |
| | 7df | | >50 | >50 |
| | 9da | HN- | 11.62±1.93 | 3.73±1.67 |

IC50 values $> 50 \mu M$ signify lack of fitted curve through the dose-response data—that is, flat response within the range tested NA not applicable

containing wtS/TGR) showed a slight advantage over 7be (IC $_{50}$ 16.40µM of SWAP containing wtS/TGR, IC $_{50}$ 14.82µM of rS/TGR-Sec). However, 7ae did not show an effective inhibitory effect on rS/TGR-Sec. As for phenyl derivatives, 7af (IC $_{50}$ 8.87µM of SWAP containing wtS/TGR, IC $_{50}$ 3.23µM of rS/TGR-Sec) and 7cf (IC $_{50}$ 12.51µM of SWAP containing wtS/TGR) did not differ significantly in inhibiting SWAP containing wtS/TGR,

but 7**cf** did not show an inhibitory effect on rS/TGR-Sec. Among acrylate compounds, only 7**bj** showed inhibitory activity against SWAP containing wtS/TGR (IC₅₀ 5.80μ M).

Some amine derivatives (**9aa**, **9ab**, **9bd**, **9be**, **9da**) showed inhibitory activity, among which **9bd** had an inhibitory effect against SWAP containing wtSjTGR at the nanomolar level (IC₅₀ = 42nM). Compared with

Li et al. Parasites Vectors (2021) 14:225 Page 8 of 11

$$\begin{array}{c} \text{Ph} & \stackrel{\textstyle \bigwedge}{\longrightarrow} \text{OH} & \stackrel{\textstyle \bigvee}{\longrightarrow} \stackrel{\textstyle$$

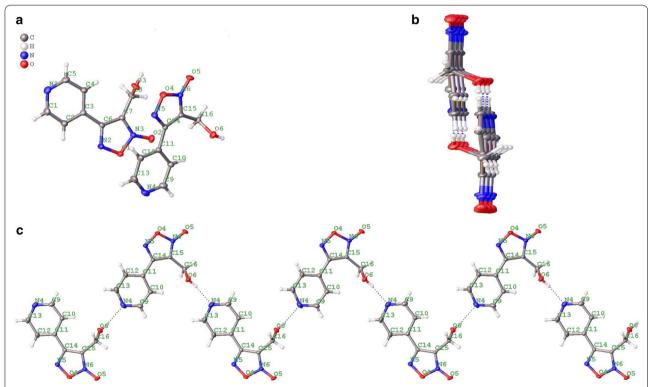


Fig. 3 a Crystal structure of 6a; b hydrogen bond helix chain formed by molecule's nitrogen atoms labeled N1, N2 and N3; c hydrogen bond zigzag chain formed by the molecule's nitrogen atoms labeled N4, N5 and N6.

phenyl amino derivative **9aa**, **9ba**, **9ca** and **9da**, the pyridine with strong electron-withdrawing ability improved the activity, such as compounds **9aa** (IC₅₀ 21.57μM to SWAP containing wtS/TGR) and **9da** (IC₅₀ 11.62μM to SWAP containing wtS/TGR, IC₅₀ 3.73μM to rS/TGR-Sec) showed better inhibition effects than **9ba** (IC₅₀ > 50μM) and **9ca** (IC₅₀ > 50μM). Similar to phenyl amino derivatives, 4-trifluoromethoxyphenyl amino derivative **9ab** (IC₅₀ 0.62μM to SWAP containing wtS/TGR, IC₅₀ 1.52μM to rS/TGR-Sec) showed better inhibitory effects than **9bb** (IC₅₀ > 50μM both to SWAP containing wtS/TGR and rS/TGR-Sec). However, among piperazine substituted derivatives (**9ac**, **9ad** and **9bd**), **9bd** exhibited

good inhibitory activity against SWAP containing wtS/TGR (IC $_{50}$ 0.042 μ M). Interestingly, compound **9be** is an intermediate of a synthetic target molecule, but it displayed much better inhibitory effects (IC $_{50}$ 0.63 μ M to SWAP containing wtS/TGR, IC $_{50}$ 0.22 μ M to rS/TGR-Sec) than furoxan. Perhaps the sulfamine or Si-O-N moieties in molecule **9be** has some influence on enzyme inhibition ability; this result might provide a new direction for the design of additional compounds.

Docking studies

To rationalize the obtained bioactivity data and to understand how the synthesized inhibitors interact with Li et al. Parasites Vectors (2021) 14:225 Page 9 of 11

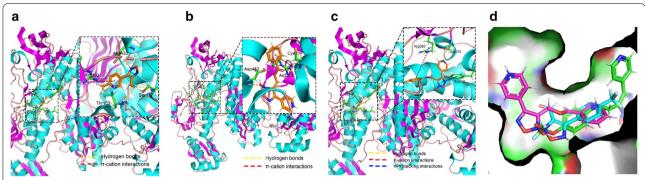


Fig. 4 In **a–c**, the protein (S_I TGR) is shown as ribbons, the synthesized inhibitors are shown as orange sticks, and the residues that can interact with inhibitors are shown as green sticks. **a** Compound **6d** forms a π -cation interaction (red dashed line) with Arg393 and hydrogen bonds (yellow dashed lines) with Ser117, Thr153 and Asp433. **b** Compound **7af** forms a π -cation interaction with Arg393 and hydrogen bonds with Cys154, and Asp433. **c** Compound **9ab** forms a π -cation interaction with Arg393, a π - π stacking interaction (blue dashed line) with Tyr296, and a hydrogen bond with Ser276. **d** Superposition of docking poses of compounds **6d** (blue), **7af** (purple), **9ab** (green) in the binding pocket of S_I TGR, S_I Chistosoma japonicum thioredoxin glutathione reductase.

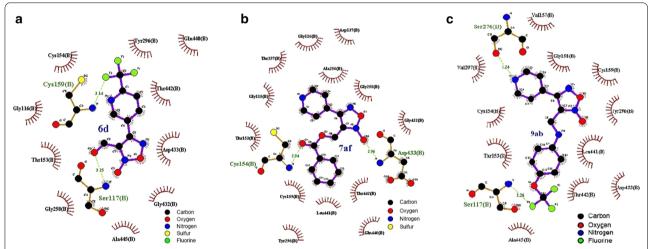


Fig. 5 LigPlot⁺ generated two-dimensional schematic overview of molecular interactions between S/TGR and compounds **6d**, **7af** and **9ab**. Hydrogen bonds are indicated by green dashed lines with corresponding distances between the atoms given in Å. Hydrophobic contacts are shown by red arcs with spokes radiating toward the atoms involved. S/TGR, Schistosoma japonicum thioredoxin glutathione reductase.

schistosomal proteins, the selected compounds with high activity **6d**, **7af** and **9ab** were docked to the available crystal structure of SjTGR (thioredoxin glutathione reductase from *Schistosoma japonicum*, PDB ID:4LA1). The crystal structure of SjTGR is a homodimer, and we retained chain B for docking studies. The results are displayed in Figs. **4** and **5**.

Compound **6d** formed four hydrogen bonds with Asp433, Ser117 and Thr153, and a π -cation interaction with Arg393 (Fig. 4a), and two strong hydrogen bonds existed between the hydroxyl moiety of **6d** and S*j*TGR. These results could explain the highest activity of **6d**. Compound **7af** formed two hydrogen bonds with Cys154 and Asp433, and a π -cation interaction with Arg393

(Fig. 4b), while the pyridine ring and oxadiazole ring did not form any type of interactions with the residues of Arg393. Compound **9ab** formed one hydrogen bond with Ser276, a π -cation interaction with Arg393 and a π - π stacking interaction with Tyr296. It was observed that the oxadiazole ring participated in the formation of chemical bonds, to constitute more interaction bonds. The distance between the oxadiazole ring and other ring-shaped conjugated structures should be taken into consideration. After superimposing three compounds (**6d**, **7af** and **9ab**) together (Fig. 4d), each of them formed one π -cation interaction with Arg393; therefore, it was speculated that Arg393 played an anchor role during the binding process.

Li et al. Parasites Vectors (2021) 14:225 Page 10 of 11

On the other hand, a hydrophobic effect was shown by red arcs with spokes radiating toward the atoms involved (Fig. 5). The numbers of the residues that had hydrophobic contacts with the inhibitors were negatively associated with enzyme inhibitory activity. Among three compounds (6d, 7af and 9ab), compound 7af had the most hydrophobic contacting residues and it had the lowest activity, while compound 6d had the least hydrophobic contacting residues and it exhibited the highest activity.

Conclusions

Using furoxan as the leading compound, a series of novel esters and amine derivatives have been synthesized by modifying the cyano moiety into an ester or amine group and replacing phenyl moiety with its bioisosteres pyridine or substituted pyridine. The structure of the key intermediate for target compounds was confirmed by crystallography. The inhibitory activity of all title compounds and key intermediates was evaluated against the TrxR activity of both rSjTGR-Sec and SWAP containing wtSjTGR for the first time. Several new derivatives, 6a-d, 9ab, **9bd** and **9be**, showed better activity toward rSiTGR-Sec or SWAP containing wtSiTGR than furoxan. The SAR of these novel furoxan derivatives was preliminarily analyzed. A pyridine ring with strong electron-withdrawing ability might be beneficial for enhancing the inhibitory effect. Docking studies showed that hydroxyl moiety can form two strong hydrogen bonds with SjTGR, which is of great help in enhancing the inhibitory activity of the compounds. Compound 6d with trifluoromethyl on the pyridine ring has the highest activity and is a good SiTGR inhibitor and could be a potent antischistosomal agent.

Abbreviations

PZQ: Praziquantel; TGR: Thioredoxin glutathione reductase; SfTGR: Schistosoma japonicum TGR; SmTGR: Schistosoma mansoni TGR; rSfTGR-Sec: Recombinant Schistosoma japonicum TGR containing selenium; SWAP: Soluble worm antigen; wtSfTGR: Wild-type SfTGR; SAR: Structure—activity relationship; TrxR: Thioredoxin reductase; GR: Glutathione reductase; NADPH: Nicotinamide adenine dinucleotide phosphate; NMR: Nuclear magnetic resonance; HRMS: High-resolution mass spectrometry; SDS-PAGE: Sodium dodecyl sulfate polyacrylamide gel electrophoresis; PBS: Phosphate-buffered saline; DMSO: Dimethyl sulfoxide; EDTA: Ethylene diamine tetraacetic acid; DTNB: Dithionitrobenzoic acid; DIBAL-H: Diisobutylaluminum hydride; DMAP: Dimethylaminopyridine; PMSF: Benzoyl sulfonyl fluoride.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s13071-021-04634-4.

Additional file 1. The preparation process and spectrum of compounds.

Acknowledgements

Not applicable.

Authors' contributions

DS, YH and GL designed the experiments and drafted the manuscript. GL, HC and WZ prepared and identified the compound. QG and SL performed the biological experiments and conducted the data analysis. CF performed the docking study. GL, QG and CF drafted the initial version of the manuscript. DS, YH and GL drafted the final version of the manuscript. All authors read and approved the final manuscript.

Funding

This work was supported by the National Natural Science Foundation of China (No. 81773560 and No. 31402192).

Availability of data and materials

Data supporting the conclusions of this article are included within the article and its additional file.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹School of Life Science and Engineering, Southwest University of Science and Technology, Mianyang 621010, China. ²National Reference Laboratory of Animal Schistosomiasis, Key Laboratory of Animal Parasitology of Ministry of Agriculture, Shanghai Veterinary Research Institute, Chinese Academy of Agricultural Sciences, Shanghai 200241, China. ³Marine College, Shandong University (Weihai), Weihai 264209, China.

Received: 27 July 2020 Accepted: 9 February 2021 Published online: 26 April 2021

References

- Song L, Li J, Xie S, Qian C, Wang J, Zhang W, et al. Thioredoxin glutathione reductase as a novel drug target evidence from Schistosoma japonicum. PLoS ONE. 2012;7:e31456.
- Song LJ, Luo H, Fan WH, Wang GP, Yin XR, Shen S, et al. Oxadiazole-2-oxides may have other functional targets, in addition to S/TGR, through which they cause mortality in Schistosoma japonicum. Parasites Vectors. 2016:9:1–12.
- Gryseels B, Polman K, Clerinx J, Kestens L. Human schistosomiasis. Lancet. 2006;368:1106–18.
- Zhang LJ, Xu ZM, Qian YJ, Dang H, Lv S, Xu J, et al. Endemic status of schistosomiasis in People's Republic of China in 2015. Chin J Schisto Control. 2016;28:611–7.
- Thetiot-Laurent SAL, Boissier J, Robert A, Meunier B. Schistosomiasis chemotherapy. Angew Chem Int Ed. 2013;52:7936–56.
- Cioli D, Pica-Mattoccia L, Basso A, Guidi A. Schistosomiasis control: praziquantel forever? Mol Biochem Parasitol. 2014:195:23–9.
- Guglielmo S, Cortese D, Vottero F, Rolando B, Kommer VP, Williams DL, et al. New praziquantel derivatives containing NO-donor furoxans and related furazans as active agents against Schistosoma mansoni. Eur J Med Chem. 2014:84:135–45.
- Sadhu PS, Kumar SN, Chandrasekharam M, Pica-Mattoccia L, Cioli D, Rao VJ. Synthesis of new praziquantel analogues: potential candidates for the treatment of schistosomiasis. Bioorg Med Chem Lett. 2012;22:1103–6.
- Zheng Y, Dong L, Hu C, Zhao B, Yang C, Xia C, et al. Development of chiral praziquantel analogues as potential drug candidates with activity to juvenile Schistosoma japonicum. Bioorg Med Chem Lett. 2014;24:4223–6.
- Sayed AA, Simeonov A, Thomas CJ, Inglese J, Austin CP, Williams DL. Identification of oxadiazoles as new drug leads for the control of schistosomiasis. Nat Med. 2008;14:407.
- 11. Rai G, Sayed AA, Lea WA, Luecke HF, Chakrapani H, Prast-Nielsen S, et al. Structure mechanism insights and the role of nitric oxide donation guide

Li et al. Parasites Vectors (2021) 14:225 Page 11 of 11

- the development of oxadiazole-2-oxides as therapeutic agents against schistosomiasis. J Med Chem. 2009;52:6474–83.
- Bonilla M, Denicola A, Novoselov SV, Turanov AA, Protasio A, Izmendi D, et al. Platyhelminth mitochondrial and cytosolic redox homeostasis is controlled by a single thioredoxin glutathione reductase and dependent on selenium and glutathione. J Biol Chem. 2008;283:17898–907.
- Kuntz AN, Davioud-Charvet E, Sayed AA, Califf LL, Dessolin J, Arnér ES, et al. Thioredoxin glutathione reductase from Schistosoma mansoni: an essential parasite enzyme and a key drug target. PLos Med. 2007;4:e206.
- 14. Yin J, Li G, Ren X, Herrler G. Select what you need: a comparative evaluation of the advantages and limitations of frequently used expression systems for foreign genes. J Biotechnol. 2007;127:335–47.
- Gillis EP, Eastman KJ, Hill MD, Donnelly DJ, Meanwell NA. Applications of fluorine in medicinal chemistry. J Med Chem. 2015;58:8315–59.
- Sano S, Yokoyama K, Fukushima M, Yagi T, Nagao Y. New reaction mode of the Horner–Wadsworth–Emmonsreaction using Sn(OSO2CF3)2 and N-ethylpiperidine. Chem Commun. 1997;28:559–60.
- Rai G, Thomas CJ, Leister W, Maloney DJ. Synthesis of oxadiazole-2-oxide analogues as potential antischistosomal agents. Tetrahedron Lett. 2009;50:1710–3.
- Dolomanov OV, Bourhis LJ, Gildea RJ, Howard JAK, Puschmann H. OLEX2: a complete structure solution, refinement and analysis program. J Appl Crystallogr. 2009;42:339–41.

- Rengby O, Johansson L, Carlson LA, Serini E, Vlamis-Gardikas A, Karsnas P, et al. Assessment of production conditions for efficient use of *Escherichia* coli in high-yield heterologous recombinant selenoprotein synthesis. Appl Environ Microbiol. 2004;70:5159–67.
- Alger HM, Williams DL. The disulfide redox system of Schistosoma mansoni and the importance of a multifunctional enzyme, thioredoxin glutathione reductase. Mol Biochem Parasitol. 2002;121:129–39.
- Han Y, Zhang M, Hong Y, Zhu Z, Li D, Li X, et al. Characterization of thioredoxin glutathione reductase in *Schiotosoma japonicum*. Parasitol Int. 2012;61:475–80.
- Peng Y, Wu Q, Huang F, Chen J, Li X, Zhou X, et al. Crystal structure of S/TGR complex with FAD. RCSB PDB. http://www.rcsb.org/structure/4LA1. Accessed 15 Oct 2019.
- Gasco AM, Fruttero R, Sorba G, Gasco A. Unsymmetrically substituted furoxans, XIII. Phenylfuroxancarbaldehydes and related compounds. Eur J Org Chem. 1991;11:1211–3.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- $\bullet\,$ thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

