

SHORT REPORT

Open Access



Trends in taxonomy of Triatomini (Hemiptera, Reduviidae, Triatominae): reproductive compatibility reinforces the synonymization of *Meccus* Stål, 1859 with *Triatoma* Laporte, 1832

Natália Regina Cesareto^{1†}, Jader de Oliveira^{2,3†}, Amanda Ravazi¹, Fernanda Fernandez Madeira⁴, Yago Visinho dos Reis¹, Ana Beatriz Bortolozo de Oliveira⁴, Roberto Dezan Vicente¹, Daniel Cesareto Cristal³, Cleber Galvão^{5*} Maria Tercília Vilela de Azeredo-Oliveira⁴, João Aristeu da Rosa³ and Kaio Cesar Chaboli Alevi^{1,3}

Abstract

Background: *Meccus'* taxonomy has been quite complex since the first species of this genus was described by Burmeister in 1835 as *Conorhinus phyllosoma*. In 1859 the species was transferred to the genus *Meccus* and in 1930 to *Triatoma*. However, in the twentieth century, the *Meccus* genus was revalidated (alteration corroborated by molecular studies) and, in the twenty-first century, through a comprehensive study including more sophisticated phylogenetic reconstruction methods, *Meccus* was again synonymous with *Triatoma*. Events of natural hybridization with production of fertile offspring have already been reported among sympatric species of the *T. phyllosoma* subcomplex, and experimental crosses demonstrated reproductive viability among practically all species of the *T. phyllosoma* subcomplex that were considered as belonging to the genus *Meccus*, as well as between these species and species of *Triatoma*. Based on the above, we carried out experimental crosses between *T. longipennis* (considered *M. longipennis* in some literature) and *T. mopan* (always considered as belonging to *Triatoma*) to evaluate the reproductive compatibility between species of the *T. phyllosoma* complex. In addition, we have grouped our results with information from the literature regarding crosses between species that were grouped in the genus *Meccus* with *Triatoma*, in order to discuss the importance of experimental crosses to confirm the generic reorganization of species.

Results: The crosses between *T. mopan* female and *T. longipennis* male resulted in viable offspring. The hatching of hybrids, even if only in one direction and/or at low frequency, demonstrates reproductive compatibility and homeology between the genomes of the parents.

*Correspondence: clebergalvao@gmail.com

†Natália Regina Cesareto and Jader de Oliveira contributed equally as first authors

⁵ Laboratório Nacional e Internacional de Referência em Taxonomia de Triatomíneos, Instituto Oswaldo Cruz (FIOCRUZ), Av. Brasil 4365, Pavilhão Rocha Lima, sala 505, Rio de Janeiro, RJ 21040-360, Brazil

Full list of author information is available at the end of the article



© The Author(s) 2021. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Conclusion: Considering that intergeneric crosses usually do not result in viable offspring in Triatominae, the reproductive compatibility observed between the *T. phyllosoma* subcomplex species considered in the *Meccus* genus with species of the *Triatoma* genus shows that there is "intergeneric" genomic compatibility, which corroborates the generic reorganization of *Meccus* in *Triatoma*.

Keywords: Chagas disease vector, Triatomines, *T. longipennis*, *T. mohan*, Experimental crosses

Background

Triatomines are hematophagous insects of great importance for public health, since they are considered the main form of transmission of the protozoan *Trypanosoma cruzi* (Chagas, 1909) (Kinetoplastida, Trypanosomatidae), the etiological agent of Chagas disease [1]. Currently, there are 8 million infected people worldwide and around 25 million living in an area at risk of infection [1], the control of vector populations being the main measure for the reduction of new chagasic patients [1].

Triatomines are part of the Hemiptera order, Heteroptera suborder, Reduviidae family and Triatominae subfamily [2]. There are 156 species in this subfamily, distributed in 18 genera and five tribes [3–6]. The Triatomini tribe is composed of nine genera, namely, *Dipetalogaster* Usinger, 1939, *Eratyrus* Stål, 1859, *Hermanlentia* Jurburg & Galvão, 1997, *Linchcosteus* Distant, 1904, *Mepraia* Mazza, Gajardo & Jörg, 1940, *Nesotriatoma* Usinger, 1944, *Panstrongylus* Berg, 1879, *Paratriatoma* Barber, 1938, and *Triatoma* Laporte, 1832 [3, 4]. However, during the taxonomic history within this tribe, several genera have already been considered valid: *Eutriatoma* Pinto, 1926, *Conorhinus* Laporte, 1833, *Callotriatoma* Usinger, 1939, *Cenaeus* Pinto, 1925, *Neotriatoma* Pinto, 1931, *Lamus* Stål, 1859, *Mestor* Kirkaldy, 1904, *Triatomptera* Neiva & Lent, 1940, and *Meccus* Stål, 1859 [7, 8]. *Eutriatoma*, *Conorhinus*, *Neotriatoma* and *Meccus* were the genera synonymous with *Triatoma* [7, 8].

Meccus' taxonomy has been quite complex, since the first species of this genus was described by Burmeister [9] as *Conorhinus phyllosoma* Burmeister, 1835; in 1859 the species was transferred to the genus *Meccus* [10] and in 1930 to *Triatoma* [11]. However, in the twentieth century, Carcavallo et al. [12] proposed the revalidation of the *Meccus* genus based on morphological data (alteration corroborated by Hypsa et al. [13] through molecular studies). Finally, in the twenty-first century, Justi et al. [8], through a comprehensive study including more sophisticated phylogenetic reconstruction methods, again synonymized *Meccus* with *Triatoma*.

The six species initially considered as *Meccus* [*T. bas-solsae* Aguilar, Torres, Jiménez, Jurberg, Galvão & Carcavallo, 1999, *T. longipennis* Usinger, 1939, *T. mazzottii* Usinger, 1941, *T. pallidipennis* Stål, 1872, *T. phyl-lo-somus* (Burmeister, 1835), and *T. picturatus* Usinger,

1939], together with *T. bolivari* Carcavallo, Martínez & Pelaez, 1987, *T. mexicana* (Herrich-Schaeffer, 1848) and *T. ryckmani* Zeledón & Ponce, 1972, form the *T. phyllosoma* subcomplex [3]. This subcomplex, together with the *T. dimidiata* subcomplex [*T. dimidiata* (Latreille, 1811), *T. hegneri* Mazzotti, 1940, *T. huehuetenanguensis* Lima-Cordón et al., 2019, *T. mohan* Dorn et al., 2018, *T. brailovskyi* Martínez, Carcavallo & Pelaez, 1984, and *T. gomeznunezi* Martínez, Carcavallo & Jurberg, 1994], form the *T. phyllosoma* complex [3, 14, 15].

Events of natural hybridization with production of fertile offspring have already been reported among sympatric species of the *T. phyllosoma* subcomplex [16]. Experimental crosses demonstrated reproductive viability among practically all species of the *T. phyllosoma* subcomplex that were considered as belonging to genus *Meccus* in some literature [17, 18]. In addition, experimental crosses between these species and species of *Triatoma* from the *T. phyllosoma* subcomplex (*T. mexicana*) and the *T. lecticularia* complex [*T. recurva* (Stål, 1868)] also resulted in the production of hybrids [19, 20].

The study of reproductive barriers by experimental crossings was used in taxonomy (with emphasis on description, revalidation, and synonymization of species [5, 21, 22]) and systematics (with emphasis on the evolutionary relationship between species [23]) of Triatominae. Based on the above, we carried out experimental crosses between *Triatoma* species of the *T. phyllosoma* (*T. longipennis*) and *T. dimidiata* (*T. mohan*) subcomplexes, to evaluate the reproductive compatibility between species of the *T. phyllosoma* complex. In addition, we have grouped our results with information from the literature regarding crosses between species that were initially grouped in the genus *Meccus* with *Triatoma*, in order to discuss the importance of experimental crosses to confirm the generic reorganization of species.

Methods

Reciprocal experimental crosses were conducted between *T. longipennis* and *T. mohan*. These two species were chosen because both belong to the *T. phyllosoma* complex [3, 14, 15], and *T. mohan* has never been considered as belonging to *Meccus*, unlike *T. longipennis*. The insects used in the experiment came from colonies kept in the Triatominae insectary of the School

of Pharmaceutical Sciences, São Paulo State University (UNESP), Araraquara, São Paulo, Brazil. The experimental crosses were conducted in the Triatominae insectary, according to the experiments of Correia et al. [24] and Mendonça et al. [25]: the insects were sexed as fifth instar nymphs, and males and females were kept separately until they reached the adult stage to guarantee the virginity of the insects used in the crosses. For the experimental crosses, three couples from each set were placed in plastic jars (diameter 5 cm × height 10 cm) and kept at room temperature.

Results and discussion

As observed for the crosses between *T. recurva* and *T. phyllosoma* (as *M. phyllosomus*) [20] and between *T. mexicana* and *T. longipennis* [19], only the direction between *T. mopan* female and *T. longipennis* male resulted in viable offspring (Table 1). The hatching of hybrids, even if only in one direction and/or at low frequency (Table 1), demonstrates reproductive compatibility and homeology between the genomes of the parents.

Intergeneric crosses usually do not result in viable offspring in Triatominae (possibly due to pre-zygotic barriers, such as genomic incompatibility), as noted for the crossings between *Panstrongylus* and *Triatoma*, *Panstrongylus* and *Nesotriatoma*, *Rhodnius* Stål, 1859 and *Psammolestes* Bergroth, 1911 (KCCA, personal communication) and *Rhodnius* and *Triatoma* [26]. The

reproductive compatibility observed between the *T. phyllosoma* subcomplex species considered in the *Meccus* genus with species of the *Triatoma* genus (Table 1) shows that there is “intergeneric” genomic compatibility, which corroborates the regrouping of species in the same genus carried out by Justi et al. [8].

The genus *Triatoma* is a paraphyletic group comprising 82 species [3, 5, 8]. There are species of *Triatoma* related phylogenetically to the genera *Panstrongylus*, *Paratriatoma*, *Linshcosteus* and *Hermanlentia* [8], which justifies the paraphyly of the genus. The inclusion of the species of the genus *Meccus* in *Triatoma* rescues a discussion about the application of the morphological characteristics used for a long time as diagnoses for the genera of the subfamily Triatominae (as recently highlighted by Monteiro et al. [27]).

Taxonomy is a fundamental science for the entomological epidemiology of Chagas disease, because correctly classifying triatomines can assist in the activity of vector control programs [28]. Even though since 2014 the generic status of the species grouped in *Meccus* has been changed to *Triatoma*, several authors continued publishing articles using the *Meccus* nomenclature as valid [20, 29–46] and, quite mistakenly, as *Triatoma* (*Meccus*) *pallidipennis* [47–49]—since *Meccus* after the genus *Triatoma* (between parentheses) represents a subgenus and, so far, there are no valid subgenera in the subfamily Triatominae.

Table 1 Experimental crosses performed between *Triatoma* spp. and *Meccus* spp.

Crossing experiments					Number of eggs	Egg fertility
♀	<i>T. mopan</i>	x	<i>T. longipennis</i>	♂	161	98 (61%)
♀	<i>T. mazzottii</i>	x	<i>T. mexicana</i>	♂	18 ^a	12 ^a (67%)
♀	<i>T. mexicana</i>	x	<i>T. mazzottii</i>	♂	14 ^a	0 ^a (64%)
♀	<i>T. picturatus</i>	x	<i>T. mexicana</i>	♂	25 ^a	19 ^a (76%)
♀	<i>T. mexicana</i>	x	<i>T. picturatus</i>	♂	32 ^a	23 ^a (72%)
♀	<i>T. mexicana</i>	x	<i>T. longipennis</i>	♂	14 ^a	9 ^a (64%)
♀	<i>T. phyllosomus</i>	x	<i>T. mexicana</i>	♂	208 ^a	156 ^a (75%)
♀	<i>T. mexicana</i>	x	<i>T. phyllosomus</i>	♂	392 ^a	295 (75%)
♀	<i>T. recurva</i>	x	<i>T. longipennis</i>	♂	71.0 ± 78.3 ^b	6.0 ± 0 ^b (8.4%)
♀	<i>T. longipennis</i>	x	<i>T. recurva</i>	♂	74.8 ± 44.6 ^b	6.0 ± 0 ^b (8%)
♀	<i>T. recurva</i>	x	<i>T. picturatus</i>	♂	94.8 ± 39.6 ^b	5.7 ± 6.4 ^b (6%)
♀	<i>T. picturatus</i>	x	<i>T. recurva</i>	♂	136.0 ± 68.9 ^b	12.3 ± 15.4 ^b (8.8%)
♀	<i>T. recurva</i>	x	<i>T. pallidipennis</i>	♂	91.2 ± 77.3 ^b	5.0 ± 0 ^b (5.5%)
♀	<i>T. pallidipennis</i>	x	<i>T. recurva</i>	♂	54.0 ± 59.9 ^b	14.5 ± 13.4 ^b (26.8%)
♀	<i>T. recurva</i>	x	<i>T. mazzottii</i>	♂	92.7 ± 56.5 ^b	3.0 ± 1.3 ^b (3.2%)
♀	<i>T. mazzottii</i>	x	<i>T. recurva</i>	♂	119.8 ± 38.3 ^b	5.3 ± 0.6 ^b (4.4%)
♀	<i>T. recurva</i>	x	<i>T. phyllosomus</i>	♂	127.8 ± 88.1 ^b	26.0 ± 26.7 ^b (20%)

^a Martinez-Ibarra et al. [19]; ^b Martinez-Ibarra et al. [20]

Conclusion

Thus, through reproductive compatibility, we confirm the generic reorganization of *Meccus* in *Triatoma* proposed by Justi et al. [8]. In addition, we highlight the importance of the correct classification of the species of the *T. phyllosoma* subcomplex into this genus to avoid future misunderstandings by the scientific community and vector control programs.

Acknowledgements

We appreciate the à Pesquisa do Estado de São Paulo (FAPESP, Brazil), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES, Brazil) – Finance Code 001, and Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq, Brazil) for financial support.

Authors' contributions

NRC: conceptualization, methodology, investigation, writing—original draft preparation and writing—review & editing. JO: conceptualization, methodology, investigation, data curation, and writing—review & editing. AR: methodology, investigation, and data curation. FFM: methodology, investigation, and data curation. YVR: methodology, investigation, and data curation. ABO: methodology, investigation, and data curation. RDV: methodology, investigation, and data curation. DCC: methodology, investigation, and data curation. CG: conceptualization, writing—review & editing, and funding acquisition. MTVAO: conceptualization, funding acquisition, and writing—review & editing. JAR: conceptualization, resources, and writing—review & editing. KCCA: conceptualization, methodology, investigation, writing—original draft preparation, writing—review & editing, supervision, project administration, and funding acquisition. All authors read and approved the final manuscript.

Funding

The study was supported by Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP, Brazil), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES, Brazil) – Finance Code 001, and Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq, Brazil). JAR: CNPq, PQ-2, process 307 398/2018-8.

Availability of data and materials

The data supporting the conclusions of this article are included within the article.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Universidade Estadual Paulista “Júlio de Mesquita Filho” (UNESP), Instituto de Biociências, Rua Dr. Antônio Celso Wagner Zanin, 250, Distrito de Rubião Júnior, Botucatu, SP 18618-689, Brazil. ²Laboratório de Entomologia em Saúde Pública, Departamento de Epidemiologia, Faculdade de Saúde Pública, Universidade de São Paulo, Av. Dr. Arnaldo 715, São Paulo, SP, Brazil. ³Laboratório de Parasitologia, Universidade Estadual Paulista “Júlio de Mesquita Filho” (UNESP), Faculdade de Ciências Farmacêuticas, Rodovia Araraquara-Jaú km 1, Araraquara, SP 14801-902, Brazil. ⁴Laboratório de Biologia Celular, Universidade Estadual Paulista “Júlio de Mesquita Filho” (UNESP), Instituto de Biociências, Letras e Ciências Exatas, Rua Cristóvão Colombo 2265, São José Do Rio Preto, SP 15054-000, Brazil. ⁵Laboratório Nacional e Internacional de Referência em Taxonomia de Triatomíneos, Instituto Oswaldo Cruz (FIOTRIZ), Av. Brasil 4365, Pavilhão Rocha Lima, sala 505, Rio de Janeiro, RJ 21040-360, Brazil.

Received: 22 April 2021 Accepted: 11 June 2021

Published online: 26 June 2021

References

- World Health Organization. Chagas disease (American trypanosomiasis). [http://www.who.int/news-room/fact-sheets/detail/chagas-disease-\(american-trypanosomiasis\)](http://www.who.int/news-room/fact-sheets/detail/chagas-disease-(american-trypanosomiasis)). Accessed 12 Mar 2021.
- Galvão C. Vetores da doença de Chagas no Brasil. 1^a ed. Curitiba: Sociedade Brasileira de Zoologia; 2014.
- Justi SA, Galvão C. The evolutionary origin of diversity in Chagas disease vectors. Trends Parasitol. 2017;33:42–52.
- Galvão C. Taxonomia dos Vetores da Doença de Chagas da Forma à Molécula, quase três séculos de história. In: Oliveira J, Alevi KCC, Camargo LMA, Meneguetti DUO, editores. Atualidades em Medicina Tropical no Brasil: Vetores. São Paulo: Strictu Sensu Editora; 2020. p. 9–37.
- Alevi KCC, Oliveira J, Garcia ACC, Cristal DC, Delgado LMG, Bittinelli IF, et al. *Triatoma rosai* sp. nov. (Hemiptera, Triatominae): a new species of argentinian chagas disease vector described based on integrative taxonomy. Insects. 2020;11:830.
- Zhao Y, Galvão C, Cai W. *Rhodnius micki*, a new species of Triatominae (Hemiptera, Reduviidae) from Bolivia. ZooKeys. 2021;1012:71–93.
- Galvão C, Carcavallo R, Rocha DS, Jurberg J. A checklist of the current valid species of the subfamily Triatominae Jeannel, 1919 (Hemiptera, Reduviidae) and their geographical distribution, with nomenclatural and taxonomic notes. Zootaxa. 2003;202:1–36.
- Justi SA, Russo CAM, dos Mallet JR, Obara MT, Galvão C. Molecular phylogeny of Triatomini (Hemiptera: Reduviidae: Triatominae). Parasit Vect. 2014;7:149.
- Burmeister H. Handbuch der Entomologie. Berlin: Enslin; 1835.
- Stål C. Monographie der Gattung *Conorhinus* und Verwandten. Berl Entomol Zeitsch. 1859;3:99–117.
- Del Ponte E. Catálogo descriptivo de los géneros *Triatoma* Lap., *Rhodnius* Stål, e *Eratyrus* Stål. Rev. Instituto Bacteriol. Dep. Nac. Hig. 1930; 5: 855–937.
- Carcavallo RU, Jurberg J, Lent H, Noireau F, Galvão C. Phylogeny of the Triatominae (Hemiptera, Reduviidae). Proposals for taxonomic arrangements Entom Vect. 2000;7:1–99.
- Hypsia V, Tietz D, Zrzavy J, Rego RO, Galvão C, Jurberg J. Phylogeny and biogeography of Triatominae (Hemiptera, Reduviidae): molecular evidence of a New World origin of the Asiatic clade. Mol Phlog Evol. 2012;23:447–57.
- Dorn PL, Justi AS, Dale C, Stevens L, Galvão C, Cordon RL, Monroy C. Description of *Triatoma mopan* sp. n. (Hemiptera, Reduviidae, Triatominae) from a cave in Belize. Zookeys. 2018;775:69–95.
- Lima-Cordón RA, Monroy MC, Stevens L, Rodas A, Rodas GA, Dorn PL, Justi AS. Description of *Triatoma huehuetenangensis* sp. n., a potential Chagas disease vector (Hemiptera, Reduviidae, Triatominae). Zookeys. 2019;820:51–70.
- Martínez-Hernandez F, Martínez-Ibarra JA, Catalá S, Villalobos G, de la Torre P, Laclette J, Alejandre-Aguilar R, Espinoza B. Natural crossbreeding between sympatric species of the *Phyllosoma* complex (Insecta: Hemiptera: Reduviidae) indicate the existence of only one species with morphologic and genetic variations. Am J Trop Med Hyg. 2010;82:74–82.
- Martínez-Ibarra JA, Ventura-Rodríguez LV, Meillón-Isáis K, Barajas-Martínez HM, Alejandre-Aguilar P, Lupercio-Coronel R, Rocha-Chávez G, Nogueda-Torres B. Biological and genetic aspects of crosses between species of the *Phyllosoma* complex (Hemiptera: Reduviidae Triatominae). Mem Inst Oswaldo Cruz. 2008;103:236–43.
- Martínez-Ibarra JA, Grant-Guillén Y, Ventura-Rodríguez LV, Osorio-Pelayo PD, Macías-Amezcuza MD, Meillón-Isáis K, Alejandre-Aguilar R, Rodríguez-Bataz E, Nogueda-Torres B. Biological and genetic aspects of crosses between species of the genus *Meccus* (Hemiptera: Reduviidae Triatominae). Mem Inst Oswaldo Cruz. 2011;106:293–300.
- Martínez-Ibarra JA, Grant-Guillén Y, Delgadillo-Aceves IN, Zumaya-Estrada FA, Rocha-Chávez G, Salazar-Schettino PM, Alejandre-Aguilar R, Villalobos G, Nogueda-Torres B. Biological and genetic aspects of crosses between phylogenetically close species of Mexican Triatomines (Hemiptera: Reduviidae). J Med Entomol. 2011;48:705–7.

20. Martínez-Ibarra JA, Nogueda-Torres B, Licón-Trillo Á, Alejandre-Aguilar R, Salazar-Schettino PM, Vences-Blanco MO. Biological aspects of crosses between *Triatoma recurva* (Stål), 1868 (Hemiptera: Reduviidae: Triatominae) and other members of the *Phyllosoma* complex. J Vec Ecol. 2015;40:117–22.
21. Mendonça VJ, Alevi KCC, Pinotti H, Gurgel-Gonçalves R, Pita S, Guerra AL, Panzera F, Araújo RF, Azeredo-Oliveira MTV, Rosa JA. Revalidation of *Triatoma bahiensis* Sherlock & Serafim, 1967 (Hemiptera: Reduviidae) and phylogeny of the *T. brasiliensis* species complex. Zootaxa. 2016;4107:239–54.
22. Nascimento JD, Ravazi A, Alevi KCC, Pardo-Díaz C, Salgado-Roa FC, da Rosa JA, de Azeredo Oliveira MTV, de Oliveira J, Hernández C, Salazar C, Ramírez JD. Taxonomical over splitting in the *Rhodnius prolixus* (Insecta: Hemiptera: Reduviidae) clade: are *R. taquarussuensis* (da Rosa et al., 2017) and *R. neglectus* (Lent, 1954) the same species? PLoS ONE. 2019;14:e0211285.
23. Neves SJM, Sousa PS, Oliveira J, Ravazi A, Madeira FF, Reis YV, et al. Prezygotic isolation confirms the exclusion of *Triatoma melanocephala*, *T. vitticeps* and *T. tibiamaculata* of the *T. brasiliensis* subcomplex (Hemiptera: Triatominae). Infect Genet Evol. 2020;79:104149.
24. Correia N, Almeida CE, Lima-Neiva V, Gumié M, Lima MM, Medeiros LMO, Mendonça VJ, Rosa JA, Costa J. Crossing experiments confirm *Triatoma sherlocki* as a member of the *Triatoma brasiliensis* species complex. Acta Trop. 2013;128:162–7.
25. Mendonça VJ, Alevi KCC, Medeiros LM, Nascimento JD, Azeredo-Oliveira MTV, Rosa JA. Cytogenetic and morphologic approaches of hybrids from experimental crosses between *Triatoma lenti* Sherlock & Serafim, 1967 and *T. sherlocki* Papa et al. 2002 (Hemiptera: Reduviidae). Infect Genet Evol. 2014;26:123–31.
26. Perlwagora-Szumlewics A, Correia MV. Induction of male sterility manipulation of genetic mechanisms present in vector species of Chagas disease (remarks on integrating sterile-male release with insecticidal control measures against vectors of Chagas disease). Rev Inst Med Trop São Paulo. 1972;14:360–71.
27. Monteiro FA, Weirauch C, Felix F, Lazoski C, Abad-Franch F. Evolution, Systematics, and Biogeography of the Triatominae Vectors of Chagas Disease. Adv Parasitol. 2018;99:265–344.
28. Dujardin JP, Costa J, Bustamante D, Jaramillo N, Catala S. Deciphering morphology in Triatominae: the evolutionary signals. Acta Trop. 2009;110:101–11.
29. Martínez-Ibarra JA, Nogueda-Torres B, Vargas-Llamas V, García-Benavides G, Bustos-Saldaña R, Villagrán ME, de Diego-Cabrera JA, Tapia-González JM. Biological characteristics of geographically isolated populations of *Meccus mazzottii* (Hemiptera: Reduviidae) in southern Mexico. J Ins Sc. 2014;14:1–5.
30. Martínez-Ibarra JA, Nogueda-Torres B, la Cruz MÁ, Villagrán ME, Diego-Cabrera JA, Bustos-Saldaña R. Biological parameters of interbreeding subspecies of *Meccus phyllotomus* (Hemiptera: Reduviidae: Triatominae) in western Mexico. Bull Entomol Res. 2015;105:763–70.
31. Martínez-Ibarra JA, Nogueda-Torres B, del Toro-González AK, Ventura-Anacleto LÁ, Montañez-Valdez OD. Geographic variation on biological parameters of *Meccus picturatus* (Usinger), 1939 (Hemiptera: Reduviidae: Triatominae) under laboratory conditions. J Vec Ecol. 2015;40:66–70.
32. Martínez-Ibarra JA, Nogueda-Torres B, Salazar-Schettino PM, Cabrera-Bravo M, Vences-Blanco MO, Rocha-Chavez G. Transmission Capacity of *Trypanosoma cruzi* (Trypanosomatida: Trypanosomatidae) by Three Subspecies of *Meccus phyllotomus* (Heteroptera: Reduviidae) and Their Hybrids. Med Veter Entom. 2016;53:928–34.
33. Martínez-Ibarra JA, Nogueda-Torres B, García-Lin JC, Arroyo-Reyes D, Salazar-Montaño LF, Hernández-Navarro JA, Diaz-Sánchez CG, Toro-Arreola ES, Rocha-Chávez G. Importance of Hybrids of *Meccus phyllotomus mazzottii*, and *M. p. pallidipennis*, and *M. p. phyllotomus* to the Transmission of *Trypanosoma cruzi* in Mexico. Jap J Infect Dis. 2016;69:202–6.
34. Martínez-Ibarra JA, Nogueda-Torres B, Salazar-Montaño LF, García-Lin JC, Arroyo-Reyes D, Hernández-Navarro JA. Comparison of biological fitness in crosses between subspecies of *Meccus phyllotomus* (Hemiptera: Reduviidae: Triatominae) in southern Mexico. Ins Sc. 2017;24:114–21.
35. Valenzuela-Campos R, González-Rangel N, Nogueda-Torres B, Goicochea-Del Rosal G, Martínez-Ibarra JA. Biological characteristics of *Meccus phyllotomus pallidipennis* (Hemiptera: Reduviidae: Triatominae) fed on two different hosts. Rev Soc Bras Med Trop. 2019;52:e20190020.
36. Rivas N, Sánchez-Cordero V, Camacho AD, Alejandre-Aguilar R. External female genitalia of six species of the genus *Meccus* (Hemiptera: Reduviidae: Triatominae). J Vector Ecol. 2017;42:271–8.
37. Díaz L, Covarrubias K, Licón A, Astorga M, Moreno Y, Martínez JA. Parámetros biológicos de *Meccus phyllotomus phyllotomus* (Burmeister) 1835, de *Triatoma recurva* (Stål) 1868 (Hemiptera, Reduviidae) y de sus híbridos de laboratorio. Biomedica. 2017;37:77–82.
38. Favila-Ruiz G, Jiménez-Cortés JG, Córdoba-Aguilar A, Salazar-Schettino PM, Gutiérrez-Cabrera AE, Pérez-Torres A, De Fuentes-Vicente JA, Vences-Blanco MO, Bucio-Torres MI, Flores-Villegas AL, Cabrera-Bravo M. Effects of *Trypanosoma cruzi* on the phenoloxidase and prophenoloxidase activity in the vector *Meccus pallidipennis* (Hemiptera: Reduviidae). Parasit Vectors. 2018;11:434.
39. Flores-Villegas AL, Cabrera-Bravo M, Torielo C, Bucio-Torres MI, Salazar-Schettino PM, Córdoba-Aguilar A. Survival and immune response of the Chagas vector *Meccus pallidipennis* (Hemiptera: Reduviidae) against two entomopathogenic fungi, *Metarhizium anisopliae* and *Isaria fumosorosea*. Parasit Vectors. 2016;9:176.
40. Flores-Villegas AL, Cabrera-Bravo M, Pérez-Torres A, Córdoba-Aguilar A, Salazar-Schettino PM, Hernández-Velázquez VM, Torielo C. Effects on *Meccus pallidipennis* (Hemiptera: Reduviidae) eggs exposed to entomopathogenic fungi: exploring alternatives to control Chagas disease. J Med Entomol. 2018;56:284–90.
41. Flores-Villegas AL, Cabrera-Bravo M, Fuentes-Vicente JA, Jiménez-Cortés JG, Salazar-Schettino M, Bucio-Torres MI, Córdoba-Aguilar A. Coinfection by *Trypanosoma cruzi* and a fungal pathogen increases survival of Chagasic bugs: advice against a fungal control strategy. B Entomol Res. 2020;110:363–9.
42. López-Vías FI, Vázquez-Chagoyán JC, Acosta-Díbarat JP, Medina-Torres I, Díaz-Albitore M, Fernández-Rosas P, Oca-Jiménez RM. Molecular Characterization of *Trypanosoma cruzi* in Infected *Meccus pallidipennis* in the Southern Region of the State of Mexico. Mexico Vector-Borne and Zoonotic Dis. 2018;18:683–9.
43. Murillo-Alonso KT, Hernández-Velázquez VM, Salazar-Schettino PM, Cabrera-Bravo M, Torielo C. Effects of *Metarhizium anisopliae* on *Meccus pallidipennis* (Hemiptera: Reduviidae) over different types of wall surfaces. Biocontrol Sci Technol. 2019;29:466–77.
44. Madeira FF, Lima ACC, Rosa JA, Azeredo-Oliveira MTV, Alevi KCC. Nucleolar persistence phenomenon during spermatogenesis in genus *Meccus* (Hemiptera, Triatominae). Genet Mol Res. 2016; gmr.15017427.
45. Madeira FF, Borsatto KC, Lima ACC, Ravazi A, Oliveira J, Rosa JA, Azeredo-Oliveira MTV, Alevi KCC. Nucleolar persistence: peculiar characteristic of spermatogenesis of the vectors of chagas disease (Hemiptera, Triatominae). Am J Trop Med Hyg. 2016;95:1118–20.
46. Alevi KCC, Oliveira J, Rosa JA, Azeredo-Oliveira MTV. Coloration of the testicular peritoneal sheath as a synapomorphy of triatomines (Hemiptera, Reduviidae). Biota Neotrop. 2014;14:1–3.
47. Díaz-Garrido P, Sepulveda-Robles O, Martínez-Martínez I, Espinoza B. Variability of defensin genes from a Mexican endemic Triatominae: *Triatoma* (*Meccus*) *pallidipennis* (Hemiptera: Reduviidae). Biosci Rep. 2018;38:1–11.
48. Gutiérrez-Cabrera AE, Alejandre-Aguilar R, Hernández-Martínez S, Espinoza B. Development and glycoprotein composition of the perimicrovillar membrane in *Triatoma* (*Meccus*) *pallidipennis* (Hemiptera: Reduviidae). Arthropod Struct Dev. 2014;43:571–8.
49. Gutiérrez-Cabrera AE, Zandberg WF, Zenteno E, Rodríguez MH, Espinoza B, Lowenberger C. Glycosylation on proteins of the intestine and perimicrovillar membrane of *Triatoma* (*Meccus*) *pallidipennis*, under different feeding conditions. Insect Sci. 2019;26:796–808.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.