

RESEARCH

Open Access



# Long-term study of *Borrelia* and *Babesia* prevalence and co-infection in *Ixodes ricinus* and *Dermacentor reticulatus* ticks removed from humans in Poland, 2016–2019

Agnieszka Pawelczyk<sup>1</sup>, Małgorzata Bednarska<sup>2</sup>, Adrianna Hamera<sup>2</sup>, Emilia Religa<sup>2</sup>, Milena Poryszewska<sup>2</sup>, Ewa J. Mierzejewska<sup>3</sup> and Renata Welc-Falęciak<sup>2\*</sup>

## Abstract

**Background:** Lyme borreliosis (LB) is the most common vector-borne disease in Europe. Monitoring changes in the prevalence of different *Borrelia* species in ticks may be an important indicator of risk assessment and of differences in pathogenicity in humans. The objective of our study was to assess the prevalence, co-infection and distribution of *Borrelia* and *Babesia* species in ticks removed from humans in a large sample collected during a study period of 4 years.

**Methods:** The ticks were collected throughout Poland from March to November over 4-year period from 2016 to 2019. All ticks ( $n = 1953$ ) were morphologically identified in terms of species and developmental stage. Molecular screening for *Borrelia* and *Babesia* by amplification of the flagellin gene (*flaB*) or 18S rRNA marker was performed. Pathogen identity was confirmed by Sanger sequencing or PCR–restriction fragment length polymorphism analysis.

**Results:** The ticks removed from humans in Poland during this study belonged to two species: *Ixodes ricinus* (97%) and *Dermacentor reticulatus* (3%). High *Borrelia* prevalence (25.3%), including *B. miyamotoi* (8.4%), was confirmed in *Ixodes ricinus* ticks removed from humans, as was the change in frequency of occurrence of *Borrelia* species during the 4-year study. Despite *Babesia* prevalence being relatively low (1.3%), the majority of tested isolates are considered to be pathogenic to humans. *Babesia* infection was observed more frequently among *Borrelia*-positive ticks (2.7%) than among ticks uninfected with *Borrelia* (0.8%). The most frequent dual co-infections were between *Borrelia afzelii* and *Babesia microti*. The presence of *Borrelia* was also confirmed in *D. reticulatus* (12.7%); however the role of these ticks in spirochete transmission to susceptible hosts is still unclear.

**Conclusions:** Although the overall risk of developing LB after a tick bite is low in Europe, knowledge of the prevalence and distribution of *Borrelia* and *Babesia* species in ticks might be an important indicator of the risk of both these tick-borne diseases.

**Keywords:** Lyme borreliosis, Babesiosis, Co-infection, Ticks, Tick-borne diseases, *Borrelia*, *Babesia*

## Background

With 85,000 cases reported annually, Lyme borreliosis (LB) is the most common vector-borne disease in Europe [1]. The estimated incidence of LB in Poland increased dramatically from 20.3 per 100,000 inhabitants in 2007 to

\*Correspondence: [rwelc@biol.uw.edu.pl](mailto:rwelc@biol.uw.edu.pl)

<sup>2</sup> Department of Parasitology, Faculty of Biology, University of Warsaw, 1 Miecznikowa Street, 02-096 Warsaw, Poland  
Full list of author information is available at the end of the article



© The Author(s) 2021. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

53.6 per 100,000 inhabitants in 2019 (an estimated average increase from 7735 cases per year in 2007 to 20,614 cases per year in 2019) [2]. However, the reliability of current incidence data on LB is uncertain due to diagnostic problems and limited reporting [3]. At least five species of *Borrelia*, namely *Borrelia burgdorferi* (s.s.), *B. garinii*, *B. afzelii*, *B. spielmanii* and *B. bavariensis*, are known to be pathogenic to humans, with each of these species believed to be associated with different clinical manifestations. The heterogeneity among *B. burgdorferi* (s.l.) species seems to be the main factor causing the regional differences in the clinical expression of human LB [4]. *Borrelia burgdorferi* (s.s.) is believed to be arthritogenic, *B. afzelii* causes mainly skin infections and *B. garinii* is especially neurotropic. Recently, *Borrelia miyamotoi* has been identified as a human pathogen related to relapsing fever in Europe [5] and little is known about its local impact on human health. *Borrelia miyamotoi* disease (BMD) has also been confirmed in an immunocompetent patient, and BMD concurrent with Lyme disease has also been described [6].

In Europe, cases of human babesiosis is reported sporadically, with about 60 confirmed cases caused mainly by *Babesia divergens* described to date in several European countries, including France, UK, Austria, the Czech Republic, Finland, Germany, Italy, Portugal, Switzerland and Poland (for review, see [7]). Non-specific clinical symptoms of human babesiosis, such as fever, flu-like symptoms, headaches, chills, sweats and myalgia, as well as the diagnostic difficulties have a key impact on their correct diagnosis and, consequently, effective treatment [8]. *Babesia microti* infections in immunocompetent individuals often have an asymptomatic but chronic course [9]. In terms of safe blood donation, this is of fundamental importance, especially if blood recipients are immunosuppressed. Transfusion-transmitted babesiosis is being increasingly described globally, mainly in the USA [10].

The tick species *Ixodes ricinus* is associated with deciduous and mixed forests, but the increasing numbers of *I. ricinus* observed over the past decades have allowed it to extend its range to northern areas of the continent and areas located at a higher altitude [11]. Cull et al. [12] have shown that *I. ricinus* typically constitute 90–100% of all ticks removed from humans in Europe and that nymphs are the most commonly detected life stage associated with zoonotic pathogen transmission. The increase in the density of ticks, also in urban areas, and the prolonged period of activity of these arachnids are probably the result of changes occurring in the environment, such as in land use in agriculture, forest management, changes in abundance and distribution of free living animals and climate change [13–16]. The observed phenomena translate

directly into an increase in the risk of transmission of pathogens vectored by ticks, which can be a significant health problem, especially for people with an impaired immune system, whose percentage in the general population is steadily increasing [17].

*Ixodes ricinus* ticks are competent vectors for many species of pathogenic viruses, bacteria and protozoa. An important problem in the epidemiology of tick-borne diseases is co-infection, i.e. simultaneous, multi-species infections; such infections are especially difficult to diagnose in humans [18]. Co-infection in humans and animals might enhance disease severity and may have significant consequences in terms of tick-borne disease treatment and diagnosis [18, 19].

Knowledge of *Borrelia* prevalence and species distribution is crucial to our understanding of the epidemiology as well as of the prevention and diagnosis of LB. There are only a limited number of studies on the prevalence of specific pathogenic species in ticks removed from humans, which mainly provided information on the *B. burgdorferi* (s.l.) complex. In Poland, most of the studies conducted to date were on questing ticks or ticks collected from animals [20–27]. However, the evaluation of pathogens in ticks parasitizing humans may provide specific information on the risk of human exposure to tick-borne infections. The aim of our study was to assess the prevalence and distribution of *Borrelia* and *Babesia* species in ticks removed from humans in Poland, in a large sample collected during a 4-year study period.

## Methods

### Tick collection and identification

The research reported here was conducted over 4-year period from 2016 to 2019. The ticks were collected throughout Poland from March to November of each year and then were delivered directly by patients or by a delivery company in a tightly-sealed, ethanol-filled container to the Diagnostic Laboratory of Parasitic Diseases and Zoonotic Infections AmerLab Ltd, a spin-off company of the University of Warsaw and Medical University of Warsaw, within 5 days after removal from the skin by a physician or the patients themselves. Ticks were morphologically identified in terms of species and developmental stage using a standard taxonomic key [28]. Specimens that could not be identified due to being extensively damaged when being removed from the skin were not included in the study.

### DNA extraction and PCR analysis

Individual larvae, nymph and adult ticks were washed in sterile ethanol and then in sterile water to avoid DNA contamination and then homogenized using sterile a mini-homogenizer with hand-held mixing motor drive

(EURx, Gdańsk, Poland). Genomic DNA from ticks was isolated using the DNeasy Blood & Tissue Kit (Qiagen, Hilden, Germany) according to the manufacturer's protocol. Genomic DNA was used for molecular screening for spirochetes through amplification of the flagellin gene (*flaB*) marker, using the primers reported in [29]. Initial PCR conditions were modified as follows: 95 °C, 5 min (initial denaturation); then 95 °C/30 s (denaturation), 52 °C/30 s (primer annealing), 72 °C/80 s (elongation), for 35 cycles; and a final elongation at 72 °C for 7 min. Nested PCR was performed with minor modification: 95 °C, 20 s (denaturation); then 55 °C/20 s (denaturation), 72 °C/60 s (elongation). For *B. miyamotoi* detection among positive samples, specific primers for the *flaB* marker were used [24]. *Babesia* spp. were detected and identified using GR2 and GF2 primers targeting the 18S rDNA fragment. The primers and thermal profiles used in this study are described in [30]. Negative controls were performed in the absence of template DNA. *Babesia microti* King's College strain DNA isolated from infected BALB/c mice blood and sequenced *Borrelia* DNA obtained from infected ticks [24] were used as positive controls. PCR products were visualized in 1.5% agarose gels stained with Midori Green Stain (Nippon Genetics Europe, Düren, Germany).

#### Identification of *Borrelia* and *Babesia* species

*Borrelia*-positive samples from ticks collected in 2016 and 2017 and *Babesia*-positive samples from ticks collected in 2016, 2017 and 2018 were sequenced by a private company (Genomed S.A., Warszawa, Poland) in both directions. Obtained nucleotide sequences were analyzed using BLAST NCBI and MEGA v. 7.0 software [31] for sequence alignment and species typing using sequences deposited in GenBank NCBI. The new nucleotide sequences have been deposited in the GenBank database under accession numbers MW791411–MW791420.

Restriction fragment length polymorphism (RFLP) was used to differentiate *Borrelia*-positive isolates at the species level obtained in 2018 and 2019. Positive amplicons after nested PCR were digested with the restriction enzyme HpyF3I (Thermo Fisher Scientific, Waltham, MA USA), which recognizes the 5'C↓TNAG3' sequence [29], according to the manufacturer's protocol. The digestion products were separated in a 2% agarose gel, visualized and archived in the GelDoc-It imaging system (Thermo Fisher Scientific). The obtained restriction patterns enabled recognition of species of the *B. burgdorferi* complex and *B. miyamotoi*.

#### Statistical analysis

Statistical analysis was performed using IBM SPSS Statistics v. 25.0 software (IBM Corp., Armonk, NY, USA).

Prevalence of *Borrelia* and *Babesia* infection (percentage of ticks infected) was analyzed using maximum likelihood techniques based on log-linear analysis of contingency tables (HILOGLINEAR). For analysis of the prevalence of *Borrelia* and *Babesia* in ticks, we fitted the prevalence of pathogens as a binary factor (infected = 1, uninfected = 0) and then by year (4 levels: 2016–2019 for *Borrelia*; 3 levels: 2016–2018 for *Babesia*) and tick stadium (larvae, nymphs, adults). *P* values < 0.05 were considered to be statistically significant.

## Results

#### *Ixodes ricinus* and *D. reticulatus* ticks

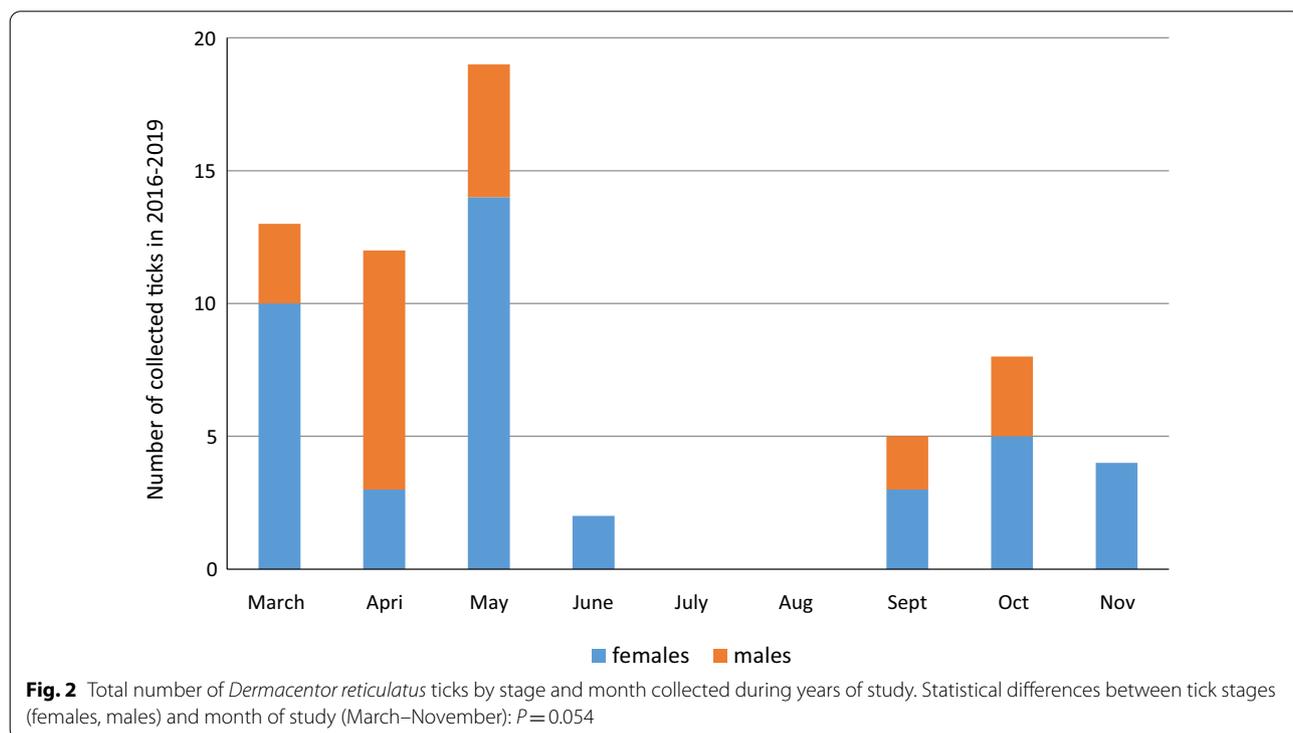
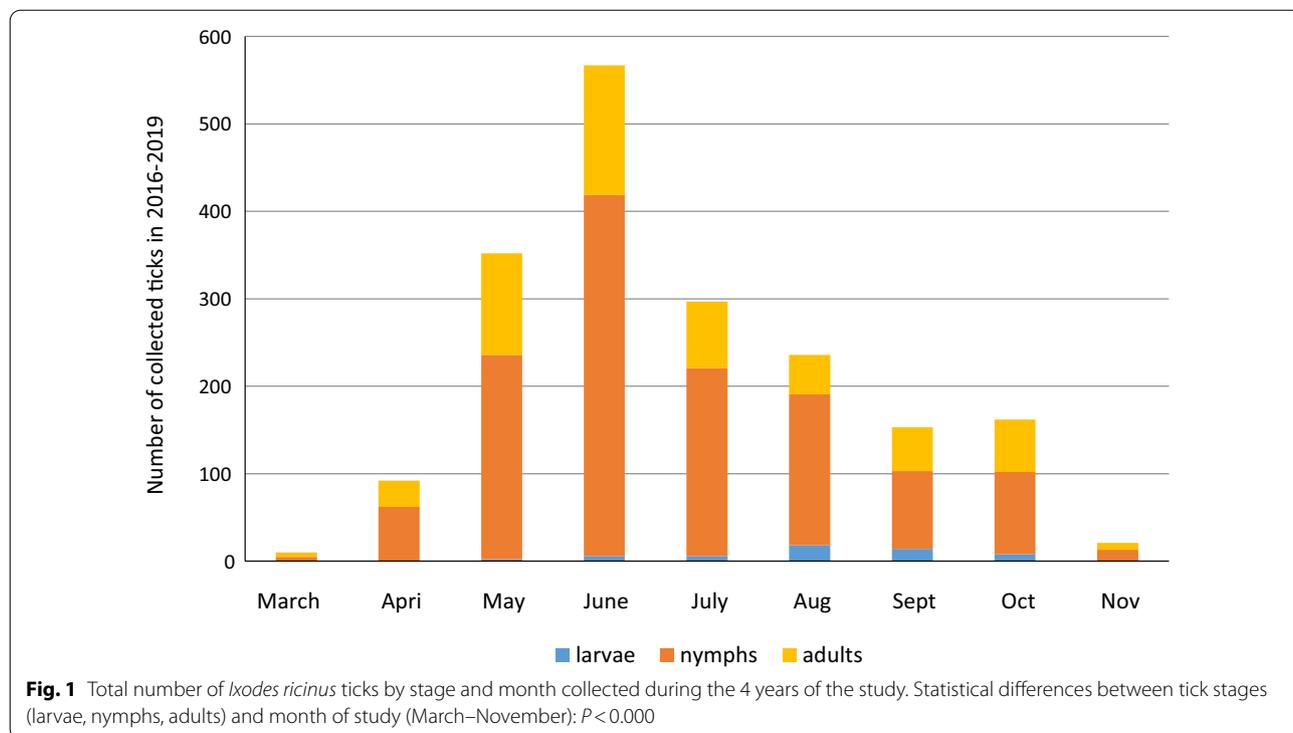
Almost all ticks collected from humans in during the study period (2016–2019) were *I. ricinus* (97%), with the other tick species identified being *D. reticulatus* (3%).

A total of 1890 *I. ricinus* ticks were collected from humans during the study period, of which 54 (2.9%) were larvae, 1298 (68.7%) were nymphs, 524 (27.7%) were females and 14 (0.7%) were males. Most of these ticks were collected in 2018–2019 ( $n=762$  and  $n=775$ , respectively), whereas in 2016–2017 only 353 ticks were analyzed ( $n=126$  and  $n=227$ , respectively). Two peaks of tick activity were observed: the first in June, followed by a second one in October; however, the mean number of ticks collected in October was almost fourfold lower than that in June (Fig. 1). The number of ticks in each stadium (larvae, nymphs and adults) removed from humans varied significantly between the months of study (month  $\times$  number of *I. ricinus* ticks in each stadium:  $\chi^2_{16} = 85.5$ ,  $P < 0.000$ ; Fig. 1; Additional File 1). Over the entire study period, the median number of larvae collected per month was six, with a minimum of two larvae (in May), a maximum of 18 larvae (in July) and no larvae collected in March–April and November. The highest number of nymphs ( $n=413$ ) and adults ( $n=148$ ) was noted in June, whereas the median number of ticks collected by month was 144 for nymphs and 60 for adults.

A total of 63 *D. reticulatus* ticks were collected from humans during the study period, of which 41 (65%) were females and 22 (35%) were males. Most *D. reticulatus* ticks were collected in 2018 and 2019 ( $n=54$ ; 85.7%). Over the entire study period, the median number of ticks collected monthly was seven; however, the highest number of ticks was noted from March to May (21 and 21, respectively; 30% of all collected ticks), and no *D. reticulatus* ticks were observed in July and August (month  $\times$  number of *D. reticulatus* ticks in each stadium:  $\chi^2_8 = 14.8$ ;  $P = 0.054$ ) (Fig. 2; Additional File 2).

#### *Borrelia* prevalence in *I. ricinus* and *D. reticulatus* ticks

Overall, the prevalence of *Borrelia* infection in the *I. ricinus* ticks removed from humans, as determined



by nested PCR, was 25.3% (479/1890; CI: 23.4–27.3%) (Table 1). Statistical analysis of prevalence over the long term (i.e. the entire study period) revealed a significant

decrease in *Borrelia* prevalence between 2016 (30.2%; CI: 22.7–38.6%) and 2019 (23.4%; CI: 20.5–26.4%) ( $\chi^2_3 = 7.58$ ;  $P = 0.051$ ; Table 1). A significant effect of tick stage was

**Table 1** Stadium and year distribution of *Borrelia*-infected ticks removed from humans between 2016 and 2019

Stadium of <i>Borrelia</i> -infected ticks	No. of tested ticks (n)	No of positive ticks <sup>a</sup>					P value
		2016	2017	2018	2019	Total	
<i>Borrelia</i> -positive <i>Ixodes ricinus</i> ticks <sup>b</sup>							
Larvae	54	2 (28.6; 6.5–64.8)	0 (0.0)	2 (8.7; 1.9–25.1)	1 (7.7; 0.8–30.7)	5 (9.3; 3.6–19.1)	=0.231
Nymphs	1298	22 (27.5; 18.6–38.0)	38 (29.2; 21.9–37.4)	129 (24.5; 21.0–28.3)	132 (23.6; 20.2–27.2)	321 (24.7; 22.5–27.2)	=0.550
Adults	538	14 (35.9; 22.3–51.5)	28 (32.6; 23.4–42.9)	63 (29.7; 23.9–36.1)	48 (23.8; 18.3–30.0)	153 (28.4; 24.7–32.3)	=0.247
Total	1890	38 (30.2; 22.7–38.6)	66 (29.1; 23.5–35.2)	194 (25.5; 22.5–28.6)	181 (23.4; 20.5–26.4)	479 (25.3; 23.4–27.3)	=0.051*
<i>Borrelia</i> -positive <i>Dermacentor reticulatus</i> ticks <sup>c</sup>							
Adults	63	1 (20; 2.3–62.9)	1 (25; 2.8–71.6)	2 (7.7; 1.6–22.5)	4 (14.3; 5.0–30.5)	8 (12.7; 6.1–22.2)	=0.740

\*Hierarchic logline analysis: year of study × *Borrelia* prevalence in *Ixodes ricinus* ticks:  $\chi^2_3 = 7.6; P = 0.051$

<sup>a</sup> Values for positive ticks are given as the percentage with the 95% confidence interval (CI) in parentheses

<sup>b</sup> Hierarchic logline analysis included: year of study × *I. ricinus* tick stages (larvae, nymphs, adults) × *Borrelia* prevalence:  $\chi^2_6 = 5.4; P = 0.500$

<sup>c</sup> Hierarchic logline analysis included: year of study × *D. reticulatus* tick stages (females, males) × *Borrelia* prevalence:  $\chi^2_3 = 2.2; P = 0.542$

also observed ( $\chi^2_2 = 11.9; P = 0.003$ ). *Borrelia* DNA was detected in 9.3% (5/54; CI: 3.6–19.1%) of *Ixodes* larvae, 24.7% (321/1298; CI: 22.5–27.2%) of *Ixodes* nymphs and 28.4% (153/538; CI: 24.7–32.3%) of adult *Ixodes* ticks (Table 1).

In total, 12.7% (8/63; CI: 6.1–22.2%) of the *D. reticulatus* ticks delivered to the laboratory during the 2016–2019 study period tested positive for *Borrelia* infections (Table 1). Prevalence of infection decreased from 20% (CI: 2.3–62.9%) in 2016 and 25% (CI: 2.8–71.6%) in 2017 to 7.7% (CI: 1.6–22.5%) in 2018 and 14.3% (CI: 5.0–30.5%) in 2019; however, only nine *D. reticulatus* ticks were tested within the first 2 years of the study (Table 1). Female ticks (9.8%; CI: 3.4–21.5%) were infected less often than males (18.2%; CI: 6.5–37.6%); however the differences between males and females were not statistically significant ( $P = 0.348$ ).

***Borrelia* species in *I. ricinus* and *D. reticulatus* ticks**

Species typing was conducted on the basis of sequencing of fragments of the *flaB* gene (540-bp product) or RFLP-PCR analysis. Species differentiation of *Borrelia*-infected ticks was successful in 251 of the 479 *Borrelia*-positive tick samples (52.4%) and, according to life stage, in 91 of 153 adults (59%), 157 of 321 nymphs (49%) and three of five larvae (60%). The most frequently detected *Borrelia* species was *B. afzelii* (65.3%; CI: 59.3–71.0%), followed by *B. burgdorferi* (10.8%; CI: 7.4–15.0%), *B. garinii* (8.8%; CI: 5.7–12.7%), *B. valaisiana* (5.2%; CI: 2.9–8.4%), *B. spielmanii* (1.2%; CI: 0.3–3.2%) and *B. lusitaniae* (0.4%; CI: 0.0–1.8%) (Table 2). The spirochete *B. miyamotoi*, which causes relapsing fever, was identified in 8.4% (CI: 5.4–12.3%) of analyzed ticks.

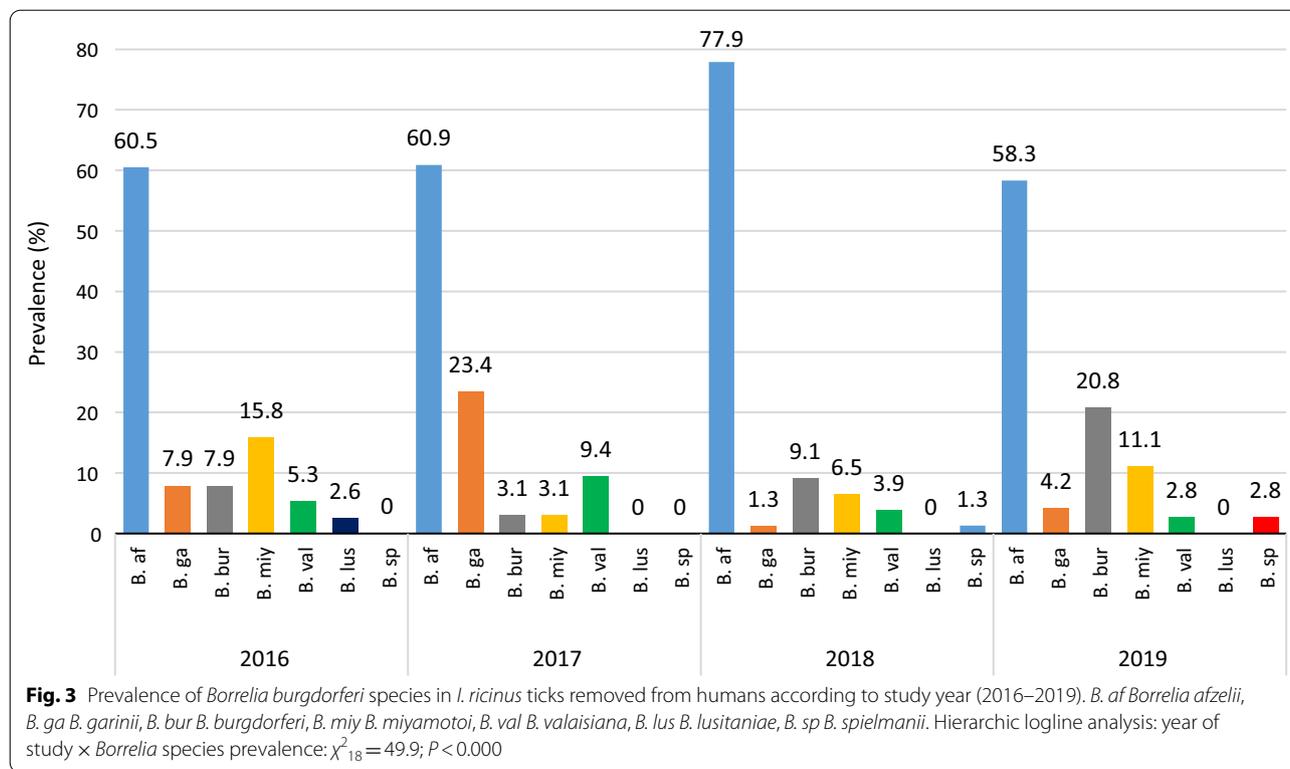
*Borrelia* species distribution showed no significant differences between tick stages ( $P = 0.231$ ) (Table 2). Adult ticks were more frequently infected with *B. afzelii* (62.6%, 57/91; CI: 52.4–72.1%) and *B. garinii* (12.1%, 11/91; CI:

**Table 2** *Borrelia* species distribution in infected *I. ricinus* ticks (n = 251) removed from humans

<i>Borrelia</i> species distribution	No of tested ticks	No of positive ticks <sup>a,b</sup>						
		<i>B. afzelii</i>	<i>B. garinii</i>	<i>B. burgdorferi</i>	<i>B. miyamotoi</i>	<i>B. valaisiana</i>	<i>B. lusitaniae</i>	<i>B. spielmanii</i>
Total	251	164 (65.3; 59.3–71.0)	22 (8.8; 5.7–12.7)	27 (10.8; 7.4–15.0)	21 (8.4; 5.4–12.3)	13 (5.2; 2.9–8.4)	1 (0.4; 0.0–1.8)	3 (1.2; 0.3–3.2)
Tick stage								
Larvae	3	2 (66.7; 17.7–96.1)	0	0	0	0	1 (33.3; 3.9–82.3)	0
Nymphs	157	105 (66.9; 59.3–73.9)	11 (7.0; 3.8–11.8)	18 (11.5; 7.2–17.1)	12 (7.6; 4.2–12.6)	8 (5.1; 2.4–9.4)	0	3 (1.9; 0.5–5.0)
Adults	91	57 (62.6; 52.4–72.1)	11 (12.1; 6.6–19.9)	9 (9.9; 5.0–17.3)	9 (9.9; 5.0–17.3)	5 (5.5; 2.1–11.6)	0	0

<sup>a</sup> Values for positive ticks are given as the percentage with the 95% CI in parentheses

<sup>b</sup> Hierarchic logline analysis included *Borrelia* species prevalence × tick stages (larvae, nymph, adults):  $\chi^2_{12} = 15.2; P = 0.231$



6.6–19.9%) than with other *Borrelia* species . In nymphs, the most commonly detected species were *B. afzelii* (66.9%, 105/157; CI: 59.3–73.9%) and *B. burgdorferi* (11.5%, 18/157; CI: 7.2–17.1%). Larvae were infected only with *B. afzelii* (66.7%, 2/3; CI: 17.7–96.1%) and *B. lusitaniae* (33.3%, 1/3; CI: 3.9–82.3%).

Species distribution in different sampling years is shown in Fig. 3 ( $\chi^2_{18} = 49.9$ ;  $P < 0.000$ ; Additional File 3). Throughout our 4-year study, the collected ticks were predominantly infected with *B. afzelii* [60.5 (CI: 44.7–74.8%), 60.9 (CI: 48.7–72.2%), 77.9 (CI: 67.7–86.1%) and 58.3% (CI: 46.8–69.2%) in 2016, 2017, 2018 and 2019, respectively]. In 2016, *B. miyamotoi* was detected in 15.8% (CI: 6.9–29.7%) of ticks; in 2017 and 2018 the number of *B. miyamotoi*-infected ticks decreased to 3.1 (CI: 0.7–9.6%) and 6.5%, (CI: 2.5–13.6%) respectively; this was followed by an increase to 11.1% (CI: 5.4–19.9%) in 2019. *Borrelia garinii* was the second most frequently noted species in 2017 (23.4%; CI: 14.4–34.8%); however, only 1.3 (CI: 0.1–5.9%) and 4.2% (CI: 1.2–10.7%) ticks were infected in 2018 and 2019. *Borrelia burgdorferi* was the most frequently identified species after *B. afzelii* in 2018 and 2019 [9.1 (CI: 4.2–17.0%) and 20.8% (CI: 12.7–31.2%)]—despite only 3.1% (CI: 0.7–9.6%) of ticks being infected in 2017.

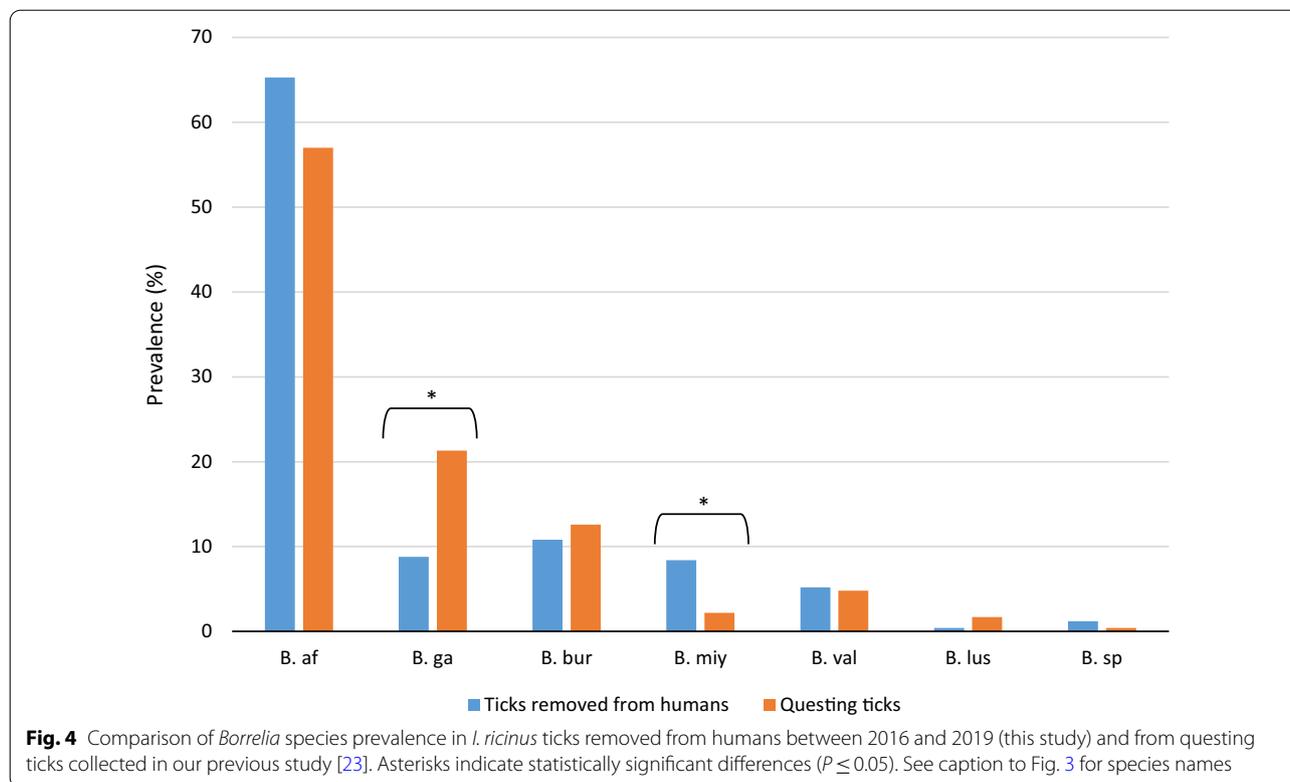
Analysis of co-infection in multiple infected ticks was performed only using RFLP-PCR in 2018 and 2019.

**Table 3** Co-infection in *I. ricinus* ticks removed from humans between 2016 and 2019

Tick stage	No. of specimens	Pathogen species
Female	1	<i>Borrelia afzelii</i> + <i>B. spielmanii</i>
Nymph	1	<i>Borrelia afzelii</i> + <i>B. burgdorferi</i> (s.s.)
Nymph	1	<i>Borrelia afzelii</i> + <i>B. miyamotoi</i>
Female	1	<i>Borrelia afzelii</i> + <i>B. burgdorferi</i> (s.s.) + <i>B. lusitaniae</i>
Female	1	<i>Borrelia valaisiana</i> + <i>Babesia microti</i>
Nymph	2	<i>Borrelia afzelii</i> + <i>Babesia microti</i>
Female	1	
Female	1	<i>Borrelia afzelii</i> + <i>Babesia canis</i>
Female	2	<i>Borrelia afzelii</i> + <i>Babesia venatorum</i>

Overall, 2.0% (3/149) of the analyzed *I. ricinus* ticks (two nymphs and female) carried two *Borrelia* species (*B. afzelii* with *B. burgdorferi*/*B. miyamotoi*/*B. spielmanii*), while triple infections were observed in only one (0.7%) female (*B. afzelii*/*B. burgdorferi*/*B. lusitaniae*) (Table 3).

Comparison of species distribution in *I. ricinus* ticks removed from humans with those from questing ticks in our previous study [24] showed that engorged ticks were by far more frequently infected with *B. miyamotoi* ( $P = 0.003$ ), whereas questing ticks were more commonly



infected with *B. garinii* ( $P=0.0001$ ). These results are shown in detail in Fig. 4.

Species differentiation of *Borrelia*-infected *D. reticulatus* ticks was successful in six of eight *Borrelia*-positive tick samples (75%). All *Borrelia* isolates were identified on the basis of RFLP-PCR analysis, as for *B. afzelii*.

**Babesia prevalence in collected ticks**

In total, 1.3% (15/1115; CI: 0.8–2.2%) of the *I. ricinus* ticks delivered to the laboratory during the study period tested positive for *Babesia* infections. No significant statistical differences between sex and stage of ticks and year of study were detected. The prevalence of *Babesia* infection ranged from 0.9% (2/227; CI: 0.2–2.8%) in 2017 to 2.4% (3/126; CI: 0.7–6.2%) in 2016. The higher *Babesia* prevalence of 2.4% (8/337; CI: 1.1–4.4%) was found in adult *I. ricinus* compared to nymphs (0.9%, 7/737; CI: 0.4–1.9%); no infected larvae were noted (Additional File 4).

Species typing was performed on the basis of sequencing of 18S rRNA gene fragment (530-bp product); all positive PCR samples were sequenced. Alignment and BLAST-NCBI analyses revealed the presence of three *Babesia* species. Of the 15 isolates, nine (60%) showed a high similarity (>99%) to the *B. microti* strain Jena

isolated originally from a human patient in Germany (EF413181 [32]). The nucleotide sequences of five isolates (33.3%) were identical to *B. venatorum* isolated from human patients in Italy and Austria (AY046575 [33]). One isolate was identified as *B. canis* with a similarity level of >99% to another Polish isolate (KT272401 [34]).

During 3 years of study (2016, 2017, 2018), one *D. reticulatus* tick (1/36, 2.8%) was infected with *B. canis* which showed a similarity level of >99% to another Polish isolate (KT272401; [34]).

**Borrelia and Babesia co-infection in I. ricinus ticks**

Statistical analysis of co-infection in *I. ricinus* ticks revealed significant differences among infected ticks ( $\chi^2_1=4.81$ ;  $P=0.028$ ). *Babesia*-positive *I. ricinus* ticks were more frequently observed among *Borrelia*-positive ticks (2.7%; 8/290) than among ticks uninfected with *Borrelia* (0.8%; 7/810). The most frequent dual co-infections were between *B. afzelii* and *B. microti* (3/7), followed by *B. afzelii* and *B. venatorum* (2/7; Table 3).

**Discussion**

We performed a long-term study of *Borrelia* and *Babesia* prevalence in two tick species removed from humans in Poland. The results of this study may have important implications in terms of public health since the

prevalence of *B. burgdorferi* (s.l.) spirochetes in ticks has been considered an essential element of risk assessment for LB [35].

The ticks collected in this study belonged to two species. While almost the whole of Europe is considered to be an endemic region for *I. ricinus*, the geographical range of *D. reticulatus* in Europe is discontinuous, with two main macroregions, and the spreading of *D. reticulatus* is believed to be associated with the loss of forest area [36]. The adult ticks collected in our study appeared to show a bimodal activity pattern, with the highest density in March–May and September–November, whereas no ticks were collected in the summer, which is typical for this tick species [37]. *Dermacentor reticulatus* represents the second genus of medical and veterinary importance after *Ixodes* in Europe. This tick species can bite humans [38] and is sporadically removed from patients in Germany, Belgium and Poland [39–41]. Several pathogens, including *B. burgdorferi*, have been detected in *D. reticulatus* ticks, suggesting a possible role of this tick species in the life-cycle and transmission of spirochetes, but it does not necessarily mean that they are capable of transmitting these bacteria to a susceptible host. Previous studies have shown that *Borrelia* prevalence in questing *D. reticulatus* ticks is significantly lower (by up to 4%) [34, 37, 42, 43] than that reported in our study. Nevertheless, the infection rates reported in engorged *D. reticulatus* ticks collected from dogs in a previous study [27] are similar to our results.

For *I. ricinus*, we observed peak activity in June, which is congruent with the results of our previous study on questing ticks [24] as well as with the results of other studies on the seasonality of *I. ricinus* bites on humans [12, 13, 44, 45]. The predominance of nymphs according to the month of the year observed in our study was similar to that found in other European studies on ticks collected from humans [39–41, 46–48]. We found that the activity of larvae was the highest in August and September; however, only 54 specimens in total were removed from humans. It is worth noting that the highest number of tick bites occurred during the summer period when people are more likely to be exposed to ticks by spending time outdoors, not only in natural areas.

Overall in Europe, including Poland, *Borrelia* prevalence in ticks removed from humans ranges from 5 to 29% [39–41, 46–53]. Surprisingly, between 2016 and 2019, annual *Borrelia* prevalence in ticks decreased significantly from 38 to 25%. At the same time, the number of LB cases in Poland decreased slightly from 21,220 in 2016 to 20,614 in 2019 [2]. Our previous studies have shown that annual *Borrelia* occurrence in questing *I. ricinus* ticks in Poland also varied from 8 to 15% between 2013 and 2014 [24]. These inter-annual fluctuations in

*Borrelia* prevalence may be due to climatic or other ecological factors affecting tick density or the abundance and, as a result, the availability of reservoir hosts, such as rodents or birds. It has been proven that the relative abundance of the white-footed mouse (*Peromyscus leucopus*) is positively associated with the prevalence of nymphal infection, which is regarded as the most important indicator of LB risk [54]. Interestingly, the *Borrelia* prevalence in our study differed significantly between *I. ricinus* ticks removed from humans and questing ticks [24]. Some results from other studies suggest that the abundance of spirochaetes in questing *Ixodes* ticks may be low (< 300 copies of bacteria) and, therefore, often undetectable, while blood repletion or simply the increased ambient temperature triggers bacteria growth and increases detectability, but possibly only within a short period (around 72 h after changing the conditions) [55, 56]. Understanding of this phenomenon is limited at the present time.

The observed significant lower *Borrelia* infection rates in *I. ricinus* larvae compared to nymphs and in nymphs compared to adults is in accordance with previous studies on questing as well as engorged ticks [24, 25, 39, 41, 47]. Since each tick stadium has only one blood meal from different hosts and the probability of acquiring pathogens increase with every blood meal, the highest prevalence of infection is noted in adults ticks. It is believed that transovarial transmission of *B. burgdorferi* (s.l.) is rare or non-existent and seems not to be essential for the circulation of spirochetes in Europe [57]. However, van Duijvendijk et al. [58] showed that flagged larvae can transmit *B. afzelii* and *B. miyamotoi* to rodents. Compared to other *Borrelia* species, a significantly higher efficiency of transovarial transmission from infected females to 90% of their larvae has been noted for *B. miyamotoi* [59]. Thus, in our study, larvae infected mainly with *B. miyamotoi* were expected. However, detection of the spirochetes in larvae in both the present study and other studies [40, 48, 60] strengthens the body of evidence for transovarial transmission of *Borrelia* under field conditions. Nonetheless, Faulde et al. [50] did not confirm the case of acquired LB after the bite of an infected *I. ricinus* larva. Hence, the hypothesis of *Borrelia* transmission from larvae to human needs further experimental studies.

Since different *Borrelia* species are involved in specific clinical manifestations, it is crucial to have accurate numbers on the prevalence of a particular species in order to be able to make precise assessments. The identification of different *Borrelia* species in our study revealed that *B. afzelii* was the most frequent species within the study period; these results are comparable to data on questing and engorged ticks from other European countries (reviewed in [61]). The low frequency of *B. spielmanii*

and *B. lusitaniae* could be explained by a relatively low abundance of the competent reservoir host for these species in Poland, mainly dormice (*Eliomys quercinus*, *Musccardinus avellanarius*) and lizards (*Lacerta agilis*) [62, 63]. Interestingly, in our study *I. ricinus* ticks removed from humans were more frequently infected with *B. miyamotoi* than was earlier reported for questing ticks [24], whereas the latter were significantly more often infected with *B. garinii*. We also observed that *B. afzelii* prevalence was higher in ticks removed from humans than in questing ticks. Similar results were obtained by Springer et al. [47] and Waindok et al. [39]. The differences in *Borrelia* prevalence in questing and engorged ticks might be the result of specific eco-epidemiological conditions within the habitats affecting the availability and abundance of reservoir hosts for ticks as well as for *Borrelia* spirochetes. Coipan et al. [64] also reported that *B. afzelii* and *B. bavariensis* were significantly more frequent in human cases than in questing ticks, which is related with the fact that both are rodent-associated *Borrelia* species. Rodents are the main reservoir hosts for these two *Borrelia* species, as well as for *I. ricinus* larvae and nymphs; therefore, this phenomenon might be the result of spatial overlap between habitats of rodents with areas of human activity where the risk of tick bites is significant [47, 64]. Nevertheless, no *B. bavariensis* isolates were observed in this study, likely due to using the single restriction enzyme *DdeI*, which is not able to distinguish the recently described *B. bavariensis* from *B. garinii* [21]. However, sequence analysis of *Borrelia* isolates collected in 2016 and 2017 did not confirm the presence of *B. bavariensis* species.

It is well-known that the distribution and prevalence of *Borrelia* spp. in questing and engorged ticks show significant temporal and spatial variations [10, 65] and that monitoring these changes might be an important indicator of risk assessment [66]. Surprisingly, in our study, the annual prevalence of *B. miyamotoi* was relatively high (up to 15.8% in 2016) compared to that reported in other European studies in questing as well as feeding ticks where the prevalence usually did not exceed 5–8% [24, 25, 47, 67–70]. Breuner et al. [70] showed that single *I. scapularis* nymphs effectively transmit *B. miyamotoi* while feeding and that transmission can occur within the first 24 h of nymphal attachment. Additionally, probably due to the overlap of endemic areas for *B. miyamotoi* with those for the *B. burgdorferi* (s.l.) complex, co-infections of *B. miyamotoi* with other spirochete species in *I. ricinus* ticks and humans have been observed [6, 24]. Taken together, these data indicate that the risk of *B. miyamotoi* infection in Poland should not be underestimated. So far, only one case of human *B. miyamotoi* infection has been diagnosed [71]. However, Fiecek et al. [71] suggested that

in the case of the patients who do not meet the criteria for neuroborreliosis (presence of *B. burgdorferi* antibodies only in serum, no antibodies in polymyalgia rheumatica [PMR]), *B. miyamotoi* disease should be considered. According to the National Institute of Public Health–National Institute of Hygiene in Poland [2], in 2013 only 14% of all reported cases with neurological symptoms ( $n = 1267$ ) met the clinical and laboratory criteria of neuroborreliosis (detection of antibodies in PMR) [71].

Co-infections in ticks are frequently reported, likely due to a large variety of animals from which they can ingest blood. Such co-infections may result in a combination of vertical (transovarial) and horizontal (blood meal) transmission of *Borrelia* species as well as a combination of systemic and co-feeding transmission [72]. Larval ticks taking multiple blood meals from different hosts (interrupted blood meals [59]) could also produce double infections of various host-specific species. The observed rate of co-infection prevalence in our study was significantly lower than that reported in feeding *I. ricinus* ticks in other European studies [39, 47]. Dual co-infection involving *B. afzelii*/*B. burgdorferi* or *B. afzelii*/*B. miyamotoi* were observed in nymphs, possibly a result of simultaneous infection during blood-feeding on rodents in the larval stadium. However, transovarial transmission of *B. miyamotoi* is also possible. Herrmann et al. [72] reported that *Borrelia* species which share the same vertebrate reservoir hosts frequently occur together and exhibit weak inhibition and even facilitation with respect to the spirochete load inside the nymphal tick. These authors also observed that rodents are relevant reservoir hosts for *B. miyamotoi* and *B. afzelii*, which explains the high number of double infections involving these two species. Alternatively, the co-occurrence of these two species in nymphs may be due to a combination of transovarial transmission of *B. miyamotoi* and horizontal transmission of *B. afzelii* [72].

Co-infections with *B. afzelii*/*B. spielmanii* and *B. afzelii*/*B. burgdorferi*/*B. lusitaniae* noted in *I. ricinus* females were likely the result of acquiring pathogens during successive blood meals on different reservoir hosts. Herrmann et al. [72] reported that *B. burgdorferi* (s.l.) species pairs that are specialized on different vertebrate reservoir hosts rarely occur together and that they exhibit strong inhibition with respect to spirochete load. This negative association between occurrence and spirochete load inhibition is likely caused by the vertebrate immune system (vertebrate complement system), which is present in the tick midgut and is capable of lysing spirochetes that are not adapted to that particular vertebrate host [72–74]. Nonetheless, the low prevalence of co-infection with different *Borrelia* species has suggested that the risk of this type co-infection in humans in Poland is low.

We found the prevalence of *Babesia* in the *I. ricinus* ticks removed from humans to be low and similar to that reported in other European studies on engorged as well as questing *I. ricinus* ticks [41, 75, 76]. In Europe, the majority of human babesiosis cases are caused by *Babesia divergens* [7]. However, in Poland so far only *B. microti* infections in humans have been noted [77–80]. Additionally, *B. microti* species occurred significantly more often than *B. divergens* in questing [81–83] and engorged (this study) *I. ricinus* ticks.

Recent studies concentrating on *Babesia microti* and *Borrelia burgdorferi* (s.s.) infections in rodents and ticks have indicated that co-infection with these pathogens is common in vectors and enzootic hosts, with a greater probability of co-infection than that predicted by chance, suggesting that co-infection provides a survival advantage to both pathogens; however, the molecular mechanism of these facilitation remain still unclear [18]. In our study, *Babesia*-positive *I. ricinus* ticks were observed significantly more often among *Borrelia*-positive ticks than among ticks not infected with *Borrelia*. However, it is difficult to conclude whether the observed findings are the result of positive interactions between these pathogens. Of the seven co-infections, three involved *B. afzelii* and *B. microti*, species that are associated with the same animal host (rodents); as such, the infections may have been acquired simultaneously, especially in nymphs. Diuk-Wasser et al. [18] discussed in their review that infection with Lyme spirochetes promotes the uptake of *B. microti* by ticks from the host (particularly reservoir host) and increases the suitability of the reservoir host for these parasites. Co-infections with other *Borrelia/Babesia* species in *I. ricinus* females are probably the consequences of sequential infection during feeding on various vertebrate hosts. Despite the fact that the *Borrelia/Babesia* species considered to be pathogenic for humans were involved in the majority of detected coinfections, its low prevalence suggests that the risk of this type of co-infection in humans in Poland is negligible.

## Conclusions

In conclusion, our study confirms a relatively high *Borrelia* prevalence in ticks removed from humans, with the spirochete species showing a significant annual variation. Although based on detected sequences, *B. afzelii* constitutes the majority pathogenic species, the risk of *B. miyamotoi* disease in humans should not be underestimated. Analysis of *Babesia* prevalence suggests that the risk of human babesiosis is negligible, which is consistent with the low number of babesiosis cases reported in Poland. Although the overall risk of developing LB after a tick bite in Europe is 4% [84], increased knowledge of the

prevalence and distribution of *Borrelia* and *Babesia* species in ticks may be an important indicator of both tick-borne disease risk assessment and varying pathogenicity in humans.

## Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s13071-021-04849-5>.

**Additional file 1:** The number of *I. ricinus* ticks by stage, month and year collected during four years of study.

**Additional file 2:** The number of *D.reticulatus* ticks by stage, month and year collected during the 4 years of the study

**Additional file 3:** *Borrelia* genospecies/species distribution in infected *I. ricinus* ticks ( $n = 251$ ) removed from humans between 2016 and 2019.

**Additional file 4:** Stadium and year distribution of *Babesia*-infected *I. ricinus* ticks removed from humans in 2016, 2017 and 2018.

## Acknowledgements

Special thanks to Dorota Kiewra for her valuable comments and suggestions and to Magdalena Szatan and Maciej Kowalec for their technical support in DNA isolation and PCR amplification in Laboratory AmerLab Ltd.

## Authors' contributions

AP: analysis and interpretation of data, writing—original draft, review & editing. AP, MB, AH, MP, ER and EM: methodology (tick collection and molecular analysis), visualization, analysis and interpretation of data; RWF: conceptualization, analysis and interpretation of data, statistical analysis, supervision, writing—original draft, review & editing. All authors read and approved the final manuscript.

## Funding

Not applicable.

## Availability of data and materials

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

## Declarations

### Ethics approval and consent to participate

The Internal Review Board of the Warsaw Medical University was informed about the study protocol (no. AKBE/73/2021). Informed consent was obtained from all individual participants included in the study in accordance with the Polish regulations. All experimental protocols were approved by Diagnostic Laboratory of Parasitic Diseases and Zoonotic Infections AmerLab Ltd, registered as medical entity in the National Chamber of Laboratory Diagnosticians (Poland).

### Consent for publication

Not applicable.

### Competing interests

The authors declare that they have no competing interests.

### Author details

<sup>1</sup>Department of Immunopathology of Infectious and Parasitic Diseases, Medical University of Warsaw, 3C Pawińskiego Street, 02-106 Warsaw, Poland. <sup>2</sup>Department of Parasitology, Faculty of Biology, University of Warsaw, 1 Miecznikowa Street, 02-096 Warsaw, Poland. <sup>3</sup>Wild Urban Evolution and Ecology Lab, Centre of New Technologies, Banacha 2c Street, 02-097 Warsaw, Poland.

Received: 26 March 2021 Accepted: 12 June 2021

Published online: 01 July 2021

## References

- Lindgren E, Jaenson TGT. Lyme borreliosis in Europe. Influences of climate and climate change, epidemiology, ecology and adaptation measures. Copenhagen: World Health Organization; 2006. [https://www.euro.who.int/\\_\\_data/assets/pdf\\_file/0006/96819/E89522.pdf](https://www.euro.who.int/__data/assets/pdf_file/0006/96819/E89522.pdf). Accessed Jan 2021
- National Institute of Public Health-National Institute of Hygiene. Epidemiological reports. <https://www.pzh.gov.pl/serwisy-tematyczne/meldunki-epidemiologiczne/>. Accessed Jan 2021
- Zajkowska J, Dunaj J. Lyme borreliosis. Challenges and difficulties of laboratory diagnosis. *Forum Zakażeń*. 2013;4(4):241–9.
- Margos G, Vollmer SA, Ogden NH, Fish D. Population genetics, taxonomy, phylogeny and evolution of *Borrelia burgdorferi* sensu lato. *Infect Genet Evol*. 2011;11(7):1545–63. <https://doi.org/10.1016/j.meegid.2011.07.022>.
- Tobudic S, Burgmann H, Stanek G, Winkler S, Schötta AM, Obermüller M, et al. Human *Borrelia miyamotoi* infection, Austria. *Emerg Infect Dis*. 2020;26(9):2201–4. <https://doi.org/10.3201/eid2609.191501>.
- Oda R, Kutsuna S, Sekikawa Y, Hongo I, Sato K, Ohnishi M, et al. The first case of imported *Borrelia miyamotoi* disease concurrent with Lyme disease. *J Infect Chemother*. 2017;23(5):333–5. <https://doi.org/10.1016/j.jiac.2016.12.015>.
- Hildebrandt A, Gray JS, Hunfeld KP. Human babesiosis in Europe: what clinicians need to know. *Infection*. 2013;41(6):1057–72. <https://doi.org/10.1007/s15010-013-0526-8>.
- Krause PJ. Human babesiosis. *Int J Parasitol*. 2019;49(2):165–74. <https://doi.org/10.1016/j.ijpara.2018.11.007>.
- Bloch EM, Kumar S, Krause PJ. Persistence of *Babesia microti* infection in humans. *Pathogens*. 2019;8(3):102. <https://doi.org/10.3390/pathogens8030102>.
- Ward SJ, Stramer SL, Szczepiorkowski ZM. Assessing the risk of Babesia to the United States blood supply using a risk-based decision-making approach: report of AABB's Ad Hoc Babesia Policy Working Group (original report). *Transfusion*. 2018;58(8):1916–23. <https://doi.org/10.1111/trf.14912>.
- Sormunen JJ, Kulha N, Klemola T, Mäkelä S, Vesilähti EM, Vesterinen EJ. Enhanced threat of tick-borne infections within cities? Assessing public health risks due to ticks in urban green spaces in Helsinki, Finland. *Zoonoses Public Health*. 2020;67(7):823–39. <https://doi.org/10.1111/zph.12767>.
- Cull B, Pietzsch ME, Gillingham EL, McGinley L, Medlock JM, Hansford KM. Seasonality and anatomical location of human tick bites in the United Kingdom. *Zoonoses Public Health*. 2020;67(2):112–21. <https://doi.org/10.1111/zph.12659>.
- Buczek A, Ciura D, Bartosik K, Zając Z, Kulisz J. Threat of attacks of *Ixodes ricinus* ticks (Ixodida: Ixodidae) and Lyme borreliosis within urban heat islands in south-western Poland. *Parasites Vectors*. 2014;7:562. <https://doi.org/10.1186/s13071-014-0562-y>.
- Léger E, Vourc'h G, Vial L, Chevillon C, McCoy KD. Changing distributions of ticks: causes and consequences. *Exp Appl Acarol*. 2013;59(1–2):219–44. <https://doi.org/10.1007/s10493-012-9615-0>.
- Medlock JM, Hansford KM, Bormane A, Derdakova M, Estrada-Peña A, George JC, et al. Driving forces for changes in geographical distribution of *Ixodes ricinus* ticks in Europe. *Parasit Vectors*. 2013;6:1. <https://doi.org/10.1186/1756-3305-6-1>.
- Pfäffle M, Littwin N, Mudders SV, Petney TN. The ecology of tick-borne diseases. *Int J Parasitol*. 2013;43(12–13):1059–77. <https://doi.org/10.1016/j.ijpara.2013.06.009>.
- Karp G, Schlaeffer F, Jotkowitz A, Riesenberg K. Syphilis and HIV co-infection. *Eur J Intern Med*. 2009;20:9–13. <https://doi.org/10.1016/j.ejim.2008.04.002>.
- Diuk-Wasser MA, Vannier E, Krause PJ. Coinfection by *Ixodes* tick-borne pathogens: ecological, epidemiological, and clinical consequences. *Trends Parasitol*. 2016;32(1):30–42. <https://doi.org/10.1016/j.pt.2015.09.008>.
- Krause PJ, Telford SR 3rd, Spielman A, Sikand V, Ryan R, Christianson D, et al. Concurrent Lyme disease and babesiosis. Evidence for increased severity and duration of illness. *JAMA*. 1996;275(21):1657–60.
- Dunaj J, Zajkowska JM, Kondrusik M, Gern L, Rais O, Moniuszko A, et al. *Borrelia burgdorferi* genospecies detection by RLB hybridization in *Ixodes ricinus* ticks from different sites of North-Eastern Poland. *Ann Agric Environ Med*. 2014;21(2):239–43. <https://doi.org/10.5604/1232-1966.1108583>.
- Kiewra D, Stańczak J, Richter M. *Ixodes ricinus* ticks (Acari, Ixodidae) as a vector of *Borrelia burgdorferi* sensu lato and *Borrelia miyamotoi* in Lower Silesia, Poland—preliminary study. *Ticks Tick Borne Dis*. 2014;5(6):892–7. <https://doi.org/10.1016/j.ttbdis.2014.07.004>.
- Gryczyńska A, Welc-Fałęciak R. Long-term study of the prevalence of *Borrelia burgdorferi* s.l. infection in ticks (*Ixodes ricinus*) feeding on blackbirds (*Turdus merula*) in NE Poland. *Exp Appl Acarol*. 2016;70(3):381–94. <https://doi.org/10.1007/s10493-016-0082-x>.
- Wójcik-Fatla A, Zając V, Sawczyn A, Sroka J, Cisak E, Dutkiewicz J. Infections and mixed infections with the selected species of *Borrelia burgdorferi* sensu lato complex in *Ixodes ricinus* ticks collected in eastern Poland: a significant increase in the course of 5 years. *Exp Appl Acarol*. 2016;68(2):197–212. <https://doi.org/10.1007/s10493-015-9990-4>.
- Kowalec M, Szewczyk T, Welc-Fałęciak R, Siński E, Karbowski G, Bajera A. Ticks and the city—are there any differences between city parks and natural forests in terms of tick abundance and prevalence of spirochaetes? *Parasites Vectors*. 2017;10(1):573. <https://doi.org/10.1186/s13071-017-2391-2>.
- Kubiak K, Dziekońska-Rynko J, Szymańska H, Kubiak D, Dmitryjuk M, Dzika E. Questing *Ixodes ricinus* ticks (Acari, Ixodidae) as a vector of *Borrelia burgdorferi* sensu lato and *Borrelia miyamotoi* in an urban area of north-eastern Poland. *Exp Appl Acarol*. 2019;78(1):113–26. <https://doi.org/10.1007/s10493-019-00379-z>.
- Michalik J, Wodecka B, Liberska J, Dabert M, Postawa T, Piksa K, et al. Diversity of *Borrelia burgdorferi* sensu lato species in *Ixodes* ticks (Acari: Ixodidae) associated with cave-dwelling bats from Poland and Romania. *Ticks Tick Borne Dis*. 2020;11(1): 101300. <https://doi.org/10.1016/j.ttbdis.2019.101300>.
- Michalski MM, Kubiak K, Szczotko M, Chajęcka M, Dmitryjuk M. Molecular detection of *Borrelia burgdorferi* sensu lato and *Anaplasma phagocytophilum* in ticks collected from dogs in urban areas of North-Eastern Poland. *Pathogens*. 2020;9(6):455. <https://doi.org/10.3390/pathogens9060455>.
- Hillyard PD. Ticks of North-West Europe. London: Backhuys Publishers; 1996.
- Wodecka B. *flaB* gene as a molecular marker for distinct identification of *Borrelia* species in environmental samples by the PCR-restriction fragment length polymorphism method. *Appl Environ Microbiol*. 2011;77(19):7088–92. <https://doi.org/10.1128/AEM.05437-11>.
- Bonnet S, Jouglin M, L'Hostis M, Chauvin A. *Babesia* sp. EU1 from roe deer and transmission within *Ixodes ricinus*. *Emerg Infect Dis*. 2007;13(8):1208–10. <https://doi.org/10.3201/eid1308.061560>.
- Kumar S, Stecher G, Tamura K. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol Biol Evol*. 2016;33(7):1870–4. <https://doi.org/10.1093/molbev/msw054>.
- Hildebrandt A, Hunfeld KP, Baier M, Krumbholz A, Sachse S, Lorenzen T, et al. First confirmed autochthonous case of human *Babesia microti* infection in Europe. *Eur J Clin Microbiol Infect Dis*. 2007;26(8):595–601. <https://doi.org/10.1007/s10096-007-0333-1>.
- Herwaldt BL, Cacciò S, Gherlinzoni F, Aspöck H, Slemenda SB, Piccaluga P, et al. Molecular characterization of a non-*Babesia divergens* organism causing zoonotic babesiosis in Europe. *Emerg Infect Dis*. 2003;9(8):942–8. <https://doi.org/10.3201/eid0908.020748>.
- Mierzejewska EJ, Pawelczyk A, Radkowski M, Welc-Fałęciak R, Bajera A. Pathogens vectored by the tick, *Dermacentor reticulatus*, in endemic regions and zones of expansion in Poland. *Parasites Vectors*. 2015;8:490. <https://doi.org/10.1186/s13071-015-1099-4>.
- Rauter C, Hartung T. Prevalence of *Borrelia burgdorferi* sensu lato genospecies in *Ixodes ricinus* ticks in Europe: a meta-analysis. *Appl Environ Microbiol*. 2005;71(11):7203–16. <https://doi.org/10.1128/AEM.71.11.7203-7216.2005>.
- Mierzejewska EJ, Estrada-Peña A, Bajera A. Spread of *Dermacentor reticulatus* is associated with the loss of forest area. *Exp Appl Acarol*. 2017;72(4):399–413. <https://doi.org/10.1007/s10493-017-0160-8>.
- Kohn M, Krücken J, McKay-Demeler J, Pachnicke S, Krieger K, von Samson-Himmelstjerna G. *Dermacentor reticulatus* in Berlin/Brandenburg (Germany): activity patterns and associated pathogens. *Ticks Tick Borne Dis*. 2019;10(1):191–206. <https://doi.org/10.1016/j.ttbdis.2018.10.003>.
- Estrada-Peña A, Jongejans F. Ticks feeding on humans: a review of records on human-biting Ixodoidea with special reference to pathogen transmission. *Exp Appl Acarol*. 1999;23(9):685–715. <https://doi.org/10.1023/a:1006241108739>.

39. Waindok P, Schicht S, Fingerle V, Strube C. Lyme borreliae prevalence and genospecies distribution in ticks removed from humans. *Ticks Tick Borne Dis.* 2017;8(5):709–14. <https://doi.org/10.1016/j.ttbdis.2017.05.003>.
40. Gałęziowska E, Rzymowska J, Najda N, Kolodziej P, Domżał-Drzewicka R, Rząca M, et al. Prevalence of *Borrelia burgdorferi* in ticks removed from skin of people and circumstances of being bitten—research from the area of Poland, 2012–2014. *Ann Agric Environ Med.* 2018;25(1):31–5. <https://doi.org/10.5604/12321966.1233906>.
41. Lernout T, De Regge N, Tersago K, Fonville M, Suin V, Sprong H. Prevalence of pathogens in ticks collected from humans through citizen science in Belgium. *Parasites Vectors.* 2019;12(1):550. <https://doi.org/10.1186/s13071-019-3806-z>.
42. Zając V, Wójcik-Fatla A, Sawczyn A, Cisak E, Sroka J, Kloc A, et al. Prevalence of infections and co-infections with 6 pathogens in *Dermacentor reticulatus* ticks collected in eastern Poland. *Ann Agric Environ Med.* 2017;24(1):26–32. <https://doi.org/10.5604/12321966.1233893>.
43. Dunaj J, Trzeszczkowski A, Moniuszko-Malinowska A, Rutkowski K, Pancewicz S. Assessment of tick-borne pathogens presence in *Dermacentor reticulatus* ticks in north-eastern Poland. *Adv Med Sci.* 2021;66(1):113–8. <https://doi.org/10.1016/j.advms.2021.01.002>.
44. Robertson JN, Gray JS, Stewart P. Tick bite and Lyme borreliosis risk at a recreational site in England. *Eur J Epidemiol.* 2000;16(7):647–52. <https://doi.org/10.1023/a:1007615109273>.
45. Wilhelmsson P, Lindblom P, Fryland L, Nyman D, Jaenson TG, Forsberg P, et al. *Ixodes ricinus* ticks removed from humans in Northern Europe: seasonal pattern of infestation, attachment sites and duration of feeding. *Parasites Vectors.* 2013;6:362. <https://doi.org/10.1186/1756-3305-6-362>.
46. Tijssen-Klasen E, Jacobs JJ, Swart A, Fonville M, Reimerink JH, Brandenburg AH, et al. Small risk of developing symptomatic tick-borne diseases following a tick bite in The Netherlands. *Parasites Vectors.* 2011;4:17. <https://doi.org/10.1186/1756-3305-4-17>.
47. Springer A, Raulf MK, Fingerle V, Strube C. *Borrelia* prevalence and species distribution in ticks removed from humans in Germany, 2013–2017. *Ticks Tick Borne Dis.* 2020;11(2):101363. <https://doi.org/10.1016/j.ttbdis.2019.101363>.
48. Kalmár Z, Dumitrache MO, D'Amico G, Matei IA, Ionică AM, Gherman CM, et al. Multiple tick-borne pathogens in *Ixodes ricinus* ticks collected from humans in Romania. *Pathogens.* 2020;9(5):390. <https://doi.org/10.3390/pathogens9050390>.
49. Fryland L, Wilhelmsson P, Lindgren PE, Nyman D, Ekerfelt C, Forsberg P. Low risk of developing *Borrelia burgdorferi* infection in the south-east of Sweden after being bitten by a *Borrelia burgdorferi*-infected tick. *Int J Infect Dis.* 2011;15(3):e174–81. <https://doi.org/10.1016/j.ijid.2010.10.006>.
50. Faulde MK, Rutenfranz M, Hepke J, Rogge M, Görner A, Keth A. Human tick infestation pattern, tick-bite rate, and associated *Borrelia burgdorferi* s.l. infection risk during occupational tick exposure at the Seedorf military training area, northwestern Germany. *Ticks Tick Borne Dis.* 2014;5(5):594–9. <https://doi.org/10.1016/j.ttbdis.2014.04.009>.
51. Jahfari S, Hofhuis A, Fonville M, van der Giessen J, van Pelt W, Sprong H. Molecular detection of tick-borne pathogens in humans with tick bites and erythema migrans, in the Netherlands. *PLoS Negl Trop Dis.* 2016;10(10):e0005042. <https://doi.org/10.1371/journal.pntd.0005042>.
52. Briciu VT, Flonta M, Țăulescu DF, Meyer F, Sebah D, Cârștina D, et al. Clinical and serological one-year follow-up of patients after the bite of *Ixodes ricinus* ticks infected with *Borrelia burgdorferi* sensu lato. *Infect Dis (Lond).* 2017;49(4):277–85. <https://doi.org/10.1080/23744235.2016.1258488>.
53. Beltrame A, Laroche M, Degani M, Perandin F, Bisoffi Z, Raoult D, et al. Tick-borne pathogens in removed ticks Veneto, northeastern Italy: a cross-sectional investigation. *Travel Med Infect Dis.* 2018;26:58–61. <https://doi.org/10.1016/j.tmaid.2018.08.008>.
54. Vuong HB, Chiu GS, Smouse PE, Fonseca DM, Brisson D, Morin PJ, et al. Influences of host community characteristics on *Borrelia burgdorferi* infection prevalence in blacklegged ticks. *PLoS ONE.* 2017;12(1):e0167810. <https://doi.org/10.1371/journal.pone.0167810>.
55. Piesman J, Schneider BS, Zeidner NS. Use of quantitative PCR to measure density of *Borrelia burgdorferi* in the midgut and salivary glands of feeding tick vectors. *J Clin Microbiol.* 2001;39(11):4145–8. <https://doi.org/10.1128/JCM.39.11.4145-4148.2001>.
56. Wilhelmsson P, Lindblom P, Fryland L, Ernerudh J, Forsberg P, Lindgren PE. Prevalence, diversity, and load of *Borrelia* species in ticks that have fed on humans in regions of Sweden and Åland Islands, Finland with different Lyme borreliosis incidences. *PLoS ONE.* 2013;8(11):e81433. <https://doi.org/10.1371/journal.pone.0081433>.
57. Stanek G, Wormser GP, Gray J, Strle F. Lyme borreliosis. *Lancet.* 2012;379(9814):461–73. [https://doi.org/10.1016/S0140-6736\(11\)60103-7](https://doi.org/10.1016/S0140-6736(11)60103-7).
58. van Duijvendijk G, Coipan C, Wagemakers A, Fonville M, Ersöz J, Oei A, et al. Larvae of *Ixodes ricinus* transmit *Borrelia afzelii* and *B. miyamotoi* to vertebrate hosts. *Parasites Vectors.* 2016;9:97. <https://doi.org/10.1186/s13071-016-1389-5>.
59. Richter D, Debski A, Hubalek Z, Matuschka FR. Absence of Lyme disease spirochetes in larval *Ixodes ricinus* ticks. *Vector Borne Zoonotic Dis.* 2012;12(1):21–7. <https://doi.org/10.1089/vbz.2011.0668>.
60. Hauck D, Jordan D, Springer A, Schunack B, Pachnicke S, Fingerle V, et al. Transovarial transmission of *Borrelia* spp., *Rickettsia* spp. and *Anaplasma phagocytophilum* in *Ixodes ricinus* under field conditions extrapolated from DNA detection in questing larvae. *Parasit Vectors.* 2020;13(1):176. <https://doi.org/10.1186/s13071-020-04049-7>.
61. Rizzoli A, Haufler H, Carpi G, Vourc HG, Neteler M, Rosa R. Lyme borreliosis in Europe. *Euro Surveill.* 2011;16(27):19906.
62. Ekner A, Dudek K, Sajkowska Z, Majláthová V, Majláth I, Tryjanowski P. Anaplasmataceae and *Borrelia burgdorferi* sensu lato in the sand lizard *Lacerta agilis* and co-infection of these bacteria in hosted *Ixodes ricinus* ticks. *Parasites Vectors.* 2011;4:182. <https://doi.org/10.1186/1756-3305-4-182>.
63. Richter D, Schlee DB, Matuschka FR. Reservoir competence of various rodents for the Lyme disease Spirochete *Borrelia spielmanni*. *Appl Environ Microbiol.* 2011;77(11):3565–70. <https://doi.org/10.1128/AEM.00022-11>.
64. Coipan EC, Jahfari S, Fonville M, Maassen CB, van der Giessen J, Takken W, et al. Spatiotemporal dynamics of emerging pathogens in questing *Ixodes ricinus*. *Front Cell Infect Microbiol.* 2013;3:36. <https://doi.org/10.3389/fcimb.2013.00036>.
65. Okeyo M, Hepner S, Rollins RE, Hartberger C, Straubinger RK, Marosevic D, et al. Longitudinal study of prevalence and spatio-temporal distribution of *Borrelia burgdorferi* sensu lato in ticks from three defined habitats in Latvia, 1999–2010. *Environ Microbiol.* 2020. <https://doi.org/10.1111/1462-2920.15100>.
66. Mechai S, Margos G, Feil EJ, Barairo N, Lindsay LR, Michel P, et al. Evidence for host-genotype associations of *Borrelia burgdorferi* sensu Stricto. *PLoS ONE.* 2016;11(2):e0149345. <https://doi.org/10.1371/journal.pone.0149345>.
67. Wilhelmsson P, Fryland L, Börjesson S, Nordgren J, Bergström S, Ernerudh J, et al. Prevalence and diversity of *Borrelia* species in ticks that have bitten humans in Sweden. *J Clin Microbiol.* 2010;48(11):4169–76. <https://doi.org/10.1128/JCM.01061-10>.
68. Nunes M, Parreira R, Lopes N, Maia C, Carreira T, Sousa C, et al. Molecular Identification of *Borrelia miyamotoi* in *Ixodes ricinus* from Portugal. *Vector Borne Zoonotic Dis.* 2015;15(8):515–7. <https://doi.org/10.1089/vbz.2014.1765>.
69. Răileanu C, Tauchmann O, Vasić A, Wöhnke E, Silaghi C. *Borrelia miyamotoi* and *Borrelia burgdorferi* (sensu lato) identification and survey of tick-borne encephalitis virus in ticks from north-eastern Germany. *Parasites Vectors.* 2020;13(1):106. <https://doi.org/10.1186/s13071-020-3969-7>.
70. Breuner NE, Dolan MC, Replogle AJ, Sexton C, Hojgaard A, Boegler KA, et al. Transmission of *Borrelia miyamotoi* sensu lato relapsing fever group spirochetes in relation to duration of attachment by *Ixodes scapularis* nymphs. *Ticks Tick Borne Dis.* 2017;8(5):677–81. <https://doi.org/10.1016/j.ttbdis.2017.03.008>.
71. Fiecek B, Chmielewski T, Tylewska-Wierzbanska S. *Borrelia miyamotoi*—new etiologic agent of neuroborreliosis? *Przegl Epidemiol.* 2017;71(4):531–8.
72. Herrmann C, Gern L, Voordouw MJ. Species co-occurrence patterns among Lyme borreliosis pathogens in the tick vector *Ixodes ricinus*. *Appl Environ Microbiol.* 2013;79(23):7273–80. <https://doi.org/10.1128/AEM.02158-13>.
73. Kurtenbach K, Sewell HS, Ogden NH, Randolph SE, Nuttall PA. Serum complement sensitivity as a key factor in Lyme disease ecology. *Infect Immun.* 1998;66(3):1248–51. <https://doi.org/10.1128/IAI.66.3.1248-1251>.
74. Kurtenbach K, De Michelis S, Etti S, Schäfer SM, Sewell HS, Brade V, et al. Host association of *Borrelia burgdorferi* sensu lato—the key role of host complement. *Trends Microbiol.* 2002;10(2):74–9. [https://doi.org/10.1016/S0966-842X\(01\)02298-3](https://doi.org/10.1016/S0966-842X(01)02298-3).
75. Lejal E, Marsot M, Chalvet-Monfray K, Cosson JF, Moutailler S, Vayssier-Taussat M, et al. A three-years assessment of *Ixodes ricinus*-borne

- pathogens in a French peri-urban forest. *Parasites Vectors*. 2019;12(1):551. <https://doi.org/10.1186/s13071-019-3799-7>.
76. Wilhelmsson P, Lövmar M, Krogfelt KA, Nielsen HV, Forsberg P, Lindgren PE. Clinical/serological outcome in humans bitten by *Babesia* species positive *Ixodes ricinus* ticks in Sweden and on the Åland Islands. *Ticks Tick Borne Dis*. 2020;11(4):101455. <https://doi.org/10.1016/j.ttbdis.2020.101455>.
  77. Moniuszko A, Dunaj J, Święcicka I, Zambrowski G, Chmielewska-Badora J, Zukiewicz-Sobczak W, et al. Co-infections with *Borrelia* species, *Anaplasma phagocytophilum* and *Babesia* spp. in patients with tick-borne encephalitis. *Eur J Clin Microbiol Infect Dis*. 2014;33(10):1835–41. <https://doi.org/10.1007/s10096-014-2134-7>.
  78. Welc-Falęciak R, Pawełczyk A, Radkowski M, Pancewicz SA, Zajkowska J, Siński E. First report of two asymptomatic cases of human infection with *Babesia microti* (Franca, 1910) in Poland. *Ann Agric Environ Med*. 2015;22(1):51–4. <https://doi.org/10.5604/12321966.1141394>.
  79. Moniuszko-Malinowska A, Święcicka I, Dunaj J, Zajkowska J, Czupryna P, Zambrowski G, et al. Infection with *Babesia microti* in humans with non-specific symptoms in North East Poland. *Infect Dis (Lond)*. 2016;48(7):537–43. <https://doi.org/10.3109/23744235.2016.1164339>.
  80. Dunaj J, Moniuszko-Malinowska A, Święcicka I, Andersson M, Czupryna P, Rutkowski K, et al. Tick-borne infections and co-infections in patients with non-specific symptoms in Poland. *Adv Med Sci*. 2018;63(1):167–72. <https://doi.org/10.1016/j.advms.2017.09.004>.
  81. Skotarczak B, Cichocka A. Isolation and amplification by polymerase chain reaction DNA of *Babesia microti* and *Babesia divergens* in ticks in Poland. *Ann Agric Environ Med*. 2001;8(2):187–9.
  82. Welc-Falęciak R, Bajer A, Paziewska-Harris A, Baumann-Popczyk A, Siński E. Diversity of *Babesia* in *Ixodes ricinus* ticks in Poland. *Adv Med Sci*. 2012;57(2):364–9. <https://doi.org/10.2478/v10039-012-0023-9>.
  83. Wójcik-Fatla A, Zajac V, Sawczyn A, Cisak E, Dutkiewicz J. *Babesia* spp. in questing ticks from eastern Poland: prevalence and species diversity. *Parasitol Res*. 2015;114(8):3111–6. <https://doi.org/10.1007/s00436-015-4529-5>.
  84. Hofhuis A, van de Kasstele J, Sprong H, van den Wijngaard CC, Harms MG, Fonville M, et al. Predicting the risk of Lyme borreliosis after a tick bite, using a structural equation model. *PLoS ONE*. 2017. <https://doi.org/10.1371/journal.pone.0181807>.

### Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

