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# Divergence of *Borrelia burgdorferi* sensu lato spirochetes could be driven by the host: diversity of *Borrelia* strains isolated from ticks feeding on a single bird

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## Abstract

**Background:** The controversy surrounding the potential impact of birds in spirochete transmission dynamics and their capacity to serve as a reservoir has existed for a long time. The majority of analyzed bird species are able to infect larval ticks with *Borrelia*. Dispersal of infected ticks due to bird migration is a key to the establishment of new foci of Lyme borreliosis. The dynamics of infection in birds supports the mixing of different species, the horizontal exchange of genetic information, and appearance of recombinant genotypes.

**Methods:** Four *Borrelia burgdorferi* sensu lato strains were cultured from *Ixodes minor* larvae and four strains were isolated from *Ixodes minor* nymphs collected from a single Carolina Wren (*Thryothorus ludovicianus*). A multilocus sequence analysis that included 16S rRNA, a 5S-23S intergenic spacer region, a 16S-23S internal transcribed spacer, *flagellin*, *p66*, and *ospC* separated 8 strains into 3 distinct groups. Additional multilocus sequence typing of 8 housekeeping genes, *clpA*, *clpX*, *nifS*, *pepX*, *pyrG*, *recG*, *rplB*, and *uvrA* was used to resolve the taxonomic status of bird-associated strains.

**Results:** Results of analysis of 14 genes confirmed that the level of divergence among strains is significantly higher than what would be expected for strains within a single species. The presence of cross-species recombination was revealed: *Borrelia burgdorferi* sensu stricto housekeeping gene *nifS* was incorporated into homologous locus of strain, previously assigned to *B. americana*.

**Conclusions:** Genetically diverse *Borrelia* strains are often found within the same tick or same vertebrate host, presenting a wide opportunity for genetic exchange. We report the cross-species recombination that led to incorporation of a housekeeping gene from the *B. burgdorferi* sensu stricto strain into a homologous locus of another bird-associated strain. Our results support the hypothesis that recombination maintains a majority of sequence polymorphism within *Borrelia* populations because of the re-assortment of pre-existing sequence variants. Even if our findings of broad genetic diversity among 8 strains cultured from ticks that fed on a single bird could be the exception rather than the rule, they support the theory that the diversity and evolution of LB spirochetes is driven mainly by the host.

**Keywords:** *Borrelia burgdorferi* sensu lato, *Ixodes minor*, Bird migration, Bird reservoir host, Multilocus sequence analysis, Multilocus sequence typing, Recombinant genotypes, Southeastern United States

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## Background

The list of hosts for *Ixodid* ticks that serve as reservoirs for *Borrelia* currently includes several hundred vertebrate species comprised of mammals, reptiles and birds [1]. The controversy surrounding the ability of birds to serve as reservoirs, and the impact of birds in spirochete transmission dynamics has existed for quite a long time [2-5]. Recent findings indicate that the majority of analyzed bird species are able to infect larval ticks with *Borrelia* [1,6-8]. However, reservoir capabilities of different bird species vary, as they do in mammals [9]. Infection rates in ticks removed from birds is comparable to those removed from mammals and in some cases reaches as much as 43.5% [9,10]. At least 80 bird species parasitized by *Ixodid* ticks are recognized in North America [6,11]. Additionally, 300 seabird species are involved in a global transmission cycle [12]. The role of birds in the spread of infected ticks is now well documented [12-14]. Earlier estimations have revealed, for example, that birds disperse 50 to 175 million *Ixodes scapularis* ticks across Canada each spring [15]. Today, the reservoir role of various bird species, especially those of ground-nesting and ground-foraging birds, such as thrushes, blackbirds, robins, wrens, nightingales, blue throats and pheasants, is unanimously recognized. They transport infected ticks and the pathogens, and thus serve as efficient amplifying reservoirs of some spirochete species worldwide [10,15-17].

Nineteen named spirochete species from the *Borrelia burgdorferi* sensu lato (sl) complex are recognized around the world [18-34]. The development and wide usage of new techniques such as multi locus sequence analysis (MLSA) [28] and multi locus sequence typing (MLST) [35] resulted in the description of 6 new spirochete species in the past 5 years, *B. yangtze* [20], *B. carolinensis* [31], *B. bavariensis* [26], *B. americana* [32], *B. kurtenbachii* [25] and *B. finlandensis* [34]. Divergences among known *Borrelia* species reflect ecological, evolutionary, epidemiological, adaptive and geographical differences. Certain *Borrelia* species are more common to mammalian, avian or reptilian hosts [36,37], for example. Today, all recognized *B. burgdorferi* sl species can be divided into four ecological groups: a) species adapted to mammals (i.e. *B. afzelii*, *B. carolinensis*); b) species adapted to birds (i.e. *B. garinii*, *B. americana*); c) species adapted to reptilian hosts (i.e. *B. lusitaniae*) or d) species with no specialized hosts (i.e., *B. burgdorferi* ss). Such separation is not, however, absolute. Occasionally, different *Borrelia* species have been detected in ticks found on "inappropriate hosts", whose complement normally eliminates mentioned species during the blood meal.

The key determinant of spirochete/host interaction has been associated with the complement regulator-acquiring surface proteins encoded by members of the *erp* gene family [36,38-40]. The general function of the

*erp* gene family is to bind the host-derived complement control proteins in a species-specific pattern [41]. The *erp* genes represent prophage genomes [42] that are employed in reshuffling of genetic material among *Borrelia* strains, holding the key to the adaptive radiation of *Borrelia* species [43]. The dynamics of infection in host populations will determine the opportunity for mixing of different genotypes, allowing the horizontal gene transfer of genetic material, and triggering genetic changes in the *Borrelia* complex [43].

Kurtenbach and colleagues suggested that the diversity and evolution of LB spirochetes cannot be attributed to tick diversity, but appears to be driven mainly by the host [36]. The diversity of *Borrelia* species associated with rodents, which have migration rates of 200–300 meters per generation, is much lower than in species associated with birds [33]. This conclusion is also supported by the complex population structure of *Borrelia garinii* in subarctic Eurasia. *B. garinii* is thought to be genetically and antigenically the most heterogeneous species among *B. burgdorferi* sl complex because of its association with passerine and marine birds [15]. *Borrelia* has the recombination system needed for genetic exchange. Genetically diverse strains of *Borrelia* are often found within the same tick or same vertebrate host and this gives a wide opportunity for genetic exchange [44-46].

How does the genomic diversity within a *Borrelia* population originate and how is it maintained? Is the appearance of a new species determined by selective pressure from the vertebrate immune response, point mutations or horizontal gene transfer among sympatric genomes? Here we present the analysis of a small group of 8 single-bird-associated *Borrelia* strains primarily assigned to *B. americana*. Combined MLSA and MLST analyses revealed that level of divergence among 8 strains is higher than what would be expected for strains within a single species. We report the cross-species recombination that led to incorporation of the *B. burgdorferi* ss housekeeping gene *nifS* into the homologous locus in strain SCW-30 h. Our results support the hypothesis that recombination maintains a majority of sequence polymorphism within *B. burgdorferi* sl populations [47].

## Methods

### Ticks and *Borrelia*

Eight *Borrelia* strains were isolated from two developmental stages of the hard tick *Ixodes minor* collected from a single Carolina Wren (*Thryothorus ludovicianus*) captured at the Wedge Plantation, Charleston County, South Carolina, USA, in November of 1994. Strains SCW-30a, SCW-30b, SCW-30c and SCW-30d, were isolated each from an individual pool of 3 *I. minor* larvae. Strains SCW-30e, SCW-30f, SCW-30 g and SCW-30 h

were isolated each from an individual *I. minor* nymph. Conditions for *Borrelia* cultivation were described elsewhere [30,31].

#### Primers used in this study

Previously described PCR primer sets were used for amplification of 5S-23S IGS [48], 16S rRNA [49], *flagellin* [50], *p66* [50], *ospC* [51] and 16S-23S ITS [51]. The MLST scheme was used for amplification and analysis of 8 housekeeping genes: *clpA*, *clpX*, *nifS*, *pepX*, *pyrG*, *recG*, *rplB* and *uvrA* [52]. The amplification conditions for all selected genomic loci were strictly as described.

#### General analysis of *Borrelia* isolates

DNA purification, PCR amplification, sequencing and sequence analysis were conducted according to our previously described protocol [31] and MLST scheme developed by Margos *et al.*, 2008 [52]. Total DNA from cultured spirochetes was purified using the DNeasy Blood and Tissue kit (Qiagen, USA). The MasterTaq Kit (Eppendorf, Germany) was used for amplification of selected loci. The purified PCR products were submitted for direct sequencing to the University of Washington High-Throughput Genomic Unit (Seattle, USA). Sequencing was conducted in both directions, using the same primers that were used for amplification of each locus. Sequences determined in this study have been deposited into GenBank.

The restriction fragment length polymorphism (RFLP) analysis of *Borrelia* 5S-23S IGS and *flagellin* gene was done *in silico* using the free software available at <http://insilico.ehu.es> [53] as it was previously described [31]. The 5S-23S IGS was digested with *MseI* and *DraI* restriction endonucleases; *flagellin* gene was digested with *HapII*, *HhaI*, *HincII*, *CelII*, and *DdeI*. Obtained RFLP patterns were compared with patterns already published [13,47,54].

#### Phylogenetic analysis

We sequenced 14 genomic loci for each *Borrelia* strain: i.e., 16S rRNA, a 5S-23S IGS, a 16S-23S ITR, *flagellin*, *p66*, *ospC* and housekeeping genes, *clpA*, *clpX*, *nifS*, *pepX*, *pyrG*, *recG*, *rplB*, and *uvrA*. Twelve loci were ultimately included in the phylogenetic analysis; *ospC* and 16S-23S ITS were excluded due to the high levels of polymorphism and recombination.

Sequences were aligned using Clustal X [55]. Data were evaluated for fit to 24 evolutionary models using MrModeltest [56]. The most-parameterized model that best fits the data at each locus was selected and evaluated by either the likelihood ratio test or Akaike Information Criterion [57]. Phylogenetic analyses were performed using Bayesian reconstruction methods, with the underlying model of evolution set to the chosen model in the program

MrBayes 3.1. Selected models were: GTR + G for *clpA* (579 bp) and *nifS* (564 bp) loci, GTR + I + G for *clpX* (624 bp), *pepX* (570 bp), *pyrG* (603 bp), *recG* (651 bp), *rplB* (624 bp) and *uvrA* (570 bp), HKY + G for 5S-23S IGR (275 bp), HKY + I for *flagellin* gene (487 bp), and GTR + I for *p66* (315 bp) and 16S rRNA (1363 bp) [58,59]. The Markov Chain Monte Carlo (MCMC) analysis was run for  $10 \times 10^6$  generations, sampling trees every 1000 generations, using 4 Markov chains (default heating values). Stationarity of the MCMC was evaluated using the "Are We There Yet" (AWTY) software [60] that plots the cumulative posterior probabilities for each tree. Two to three thousand burn-in trees generated before the point, at which these values stabilized, were discarded. The fifty percent majority rule consensus tree for the estimated posterior distribution of trees (with burn-in trees truncated) was assembled for each locus, using MrBayes [59]. The consensus trees for each of twelve genes (excluding *ospC* and 16S-23S ITS) were not congruent, and thus an overall pattern of relatedness could not be inferred using these gene-trees alone.

The most common approach to inferring relationships across multiple genetic loci is to combine outcomes of individual gene trees into multi-locus analysis. The Bayesian estimation of concordance among gene trees (BUCKy) approach [61], which makes no assumptions about the source of reticulation in gene tree histories was used here. BUCKy uses, as input data, the complete tree files generated by the Bayesian analysis of each individual locus, in the format generated by MrBayes [59]. BUCKy generates a sample of gene trees from the joint distribution of gene trees, from which concordance factors (CFs) are estimated with credibility intervals. The CF ranges from 0.0 to 1.0. BUCKy implements a consensus method based on unrooted quartets and which consistently identifies the species tree [62]. We ran BUCKy at several levels of  $\alpha$  to evaluate how much effect choice of this parameter value would have on the results. The final analysis selected for use was run with an  $\alpha$  of 1, a reasonable intermediate between 0 and infinity [63], using 4 heated chains in the MCMC analysis.

#### Nucleotide sequence accession numbers

Sequences determined in this study have been deposited into GenBank and given the indicated accession numbers (numbers for each isolate are given for sequenced genomic loci in the following order: 5S-23S intergenic spacer, 16S rRNA, *flagellin*, *p66*, 16S-23S ITS, and *ospC*): HM802215, HM802221, HM802227, HM802233, HQ012507 and HM852908 for SCW-30a; HM802216, HM802222, HM802228, HM802234, HQ012502 and HM852909 for SCW-30b; HM802217, HM802223, HM802229, HM802235, HQ012503 and HM852910 for SCW-30c; HM802218, HM802224, HM802230, HM802236, HQ012504 and HM852911 for

SCW-30d; HM140981 and HM146415 for 16S-23S intergenic spacer and *ospC* gene of SCW-30e; HM802219, HM802225, HM802231, HM802237, HQ012505 and HM852912 for SCW-30f; HM140982 and HM146416 for 16S-23S intergenic spacer and *ospC* gene of SCW-30 g and HM802220, HM802226, HM802232, HM802238, HQ012506 and HM852913 for SCW-30 h. Accession numbers for 5S-23S IGS, 16S rRNA, *flagellin* and *p66* of isolates SCW-30e and SCW-30 g were presented earlier in Rudenko et al., 2009 [32]. Sequences of housekeeping genes of all strains from SCW-30 group were submitted as 8 sets to the PopSet database and are accessible in GenBank using Entrez PopSet numbers: 358009204 for *clpA* genes, 358009220 for *clpX* genes, 358009236 for *nifS* genes, 358009252 for *pepX* genes, 358009268 for *pyrG* genes, 358009284 for *recG* genes, 358009300 for *rplB* genes and 358009316 for *uvrA* genes.

#### Control strains of *Borrelia burgdorferi sensu lato* species used in analysis

The sequences of corresponding genomic loci of following control strains available in GenBank were used in the phylogenetic analyses: *B. burgdorferi sensu stricto* B31; *B. afzelii* VS461; *B. americana* SCW41; *B. andersonii* 21133; *B. bavariensis* PBi; *B. bissettii* DN127; *B. californiensis* CA443; *B. carolinensis* SCW22; *B. garinii* 20047; *B. japonica* HO14; *B. kutrenbachii* 25015; *B. lusitanae* PotiB2; *B. sinica* CMN3; *B. spielmanii* A14S; *B. tanukii* HK501; *B. turdi* Ya501; *B. valaisiana* VS116; *B. yangtze* R5 and genomospecies 2 CA-2.

#### Results

The eight strains isolated from *Ixodes minor* larvae and nymphs collected from a single Carolina Wren were analyzed previously at six genomic loci, 5S-23S IGS, 16S rRNA, *flagellin*, *p66*, 16S-23S ITS and *ospC* [30]. At all six loci 8 strains clustered as larvae-associated “b/c/d” group (SCW-30b, SCW-30c, and SCW-30d), nymph-associated “e/f/g” group (SCW-30e, SCW-30f, and SCW-30 g) and standalone nymph-originated SCW-30 h strain. In a previous study, strains SCW-30e and SCW-30 g were assigned to *B. americana* [32]. In the same study strain SCW-30f was also assigned to this species by preliminary analysis of the 16S rRNA gene [32]. The larvae-originated strain SCW-30a alternately clustered with the *B. americana* “e/f/g” and “b/c/d” clusters, depending on the locus analyzed.

PCR of the 5S-23S IGS locus resulted in amplicons of 3 different sizes across the 8 samples: 254 bp for SCW-30a, SCW-30e, SCW-30f and SCW-30 g, 253 bp for SCW-30 h and 225 bp for SCW-30b, SCW-30c and SCW-30d strains. *MseI* and *DraI* RFLP patterns of SCW-30e, SCW-30f, SCW-30 g and SCW-30 h corresponded to those described for *B. americana* subgroup A and B [32]. The RFLP pattern found at this locus in SCW-30b, SCW-30c

and SCW-30d could not be associated with any known *Borrelia* species and represents four *MseI* restriction fragments, 107, 52, 38, and 28 bp, and three *DraI* restriction fragments, 144, 53, and 28 bp. The *DraI* RFLP pattern for SCW-30a was identical to that of *B. americana*, but the *MseI* pattern in that strain was different and contained 5 fragments of 107, 80, 38, 16, and 13 bp instead of the 6 specific to *B. americana* [32].

*In silico* RFLP analysis of the 488 bp *flagellin* gene involved sequence digestion with *HapII*, *HhaI*, *HincII*, *CelII*, and *DdeI* restriction endonucleases. Amplicons of SCW-30e, SCW-30f, SCW-30 g and SCW-30 h revealed the pattern specific for *B. americana* [32]. RFLP patterns in SCW-30a, SCW-30b, SCW-30c and SCW-30d at this locus were identical, but unknown, resulting in two fragments in case of restriction with *DdeI* only (338 and 150 bp).

In the *p66* locus, SCW-30b, SCW-30c and SCW-30d strains showed 99.7-100% similarity among themselves only; SCW-30e, SCW-30f and SCW-30 g strains revealed 99.0-100% similarity to the *B. americana* type strain and to SCW-30 h; strain SCW-30a was 98.1-98.7% similar to both the “b/c/d” and the “e/f/g” groups of strains.

All 8 strains were 99.6-100% similar on the 1,316 bp amplicon of 16S rRNA.

Larvae-originated SCW-30b, SCW-30c and SCW-30d were 100% identical at 16S-23S ITS locus. Previously identified as *B. americana*, SCW-30e, SCW-30f, SCW-30 g, and SCW-30 h showed 99.88 -100% similarity among themselves. Strain SCW-30a showed the lowest similarity, 85.63-88.08%, to the rest of strains at this locus.

Strains SCW-30a, SCW-30b, SCW-30c and SCW-30d were identical at the *ospC* locus and showed no significant similarity to known *ospC* types. The *ospC* genes of SCW-30e, SCW-30f and SCW-30 g were 97-99% similar to those of other *B. americana* strains. SCW-30 h *ospC* was 100% identical to one of *B. americana* type strain SCW-41<sup>T</sup> and revealed the high similarity to *ospC* allele B of *B. burgdorferi* ss strains that are widely distributed in southeastern United States [64].

Results of analysis of 6 genes using MLSA scheme from our previous studies [31,32] was insufficient to resolve the taxonomic status of the strains from SCW-30 group revealing apparent recombination of genetic material among fast evolving genomic loci.

Sequences of 8 housekeeping genes *clpA*, *clpX*, *nifS*, *pepX*, *pyrG*, *recG*, *rplB* and *uvrA* from 8 SCW-30 strains were compared with the allelic profiles using “virtual isolate collections centers” [35] from the online MLST database (www.mlst.net) [65]. The MLST database currently contains data for approximately 1,200 *Borrelia* strains comprising most of the described *B. burgdorferi* sl species from all over the world which have been

resolved into >300 sequence types (ST's) [50]. We found that only strains SCW-30f and SCW-30 g carried the same alleles as the type strain of *B. americana* SCW-41<sup>T</sup>. Strain SCW-30e was broadly variable at the housekeeping loci (Table 1). The similarity of strains SCW-30b, SCW-30c and SCW-30d were below the cut-off value for species assignment at 4 of the 8 loci analyzed, *pepX*, *pyrG*, *recG* and *uvrA* (Table 2). The divergence of the strain SCW-30 h was the highest among the group. Cross-species recombination was detected with this method of comparison, and seemed to be the result of the incorporation of a *B. burgdorferi* ss housekeeping gene, *nifS*, into the homologous locus of strain SCW-30 h. This allele is specific to *B. burgdorferi* ss strains widely distributed in the United States and in Canada (borrelia.mlst.net).

Eight concatenated housekeeping gene sequences from each SCW-30 strain (2,752 bp/strain) were aligned with 18 available in MLST database sequence types (ST) for type strain of known *Borrelia* species and genomospecies 2: ST1 (*B. burgdorferi* ss), ST64 (*B. lusitaniae*), ST70 (*B. afzelii*), ST83 (*B. garinii*), ST84 (*B. bavariensis*), ST95 (*B. valaisiana*), ST152 (*B. yangtze*), ST159 (*B. spielmanii*), ST272 (*B. bissettii*), ST280 (*B. kurtenbachii*), ST388 (*B. andersonii*), ST447 (*B. californiensis*), ST449 (*B. americana*), ST450 (*B. carolinensis*), ST453 (*B. japonica*), ST454 (*B. tanukii*), ST455 (*B. turdii*) (not shown). Genetic distance between SCW-30 group and all known species, except *B. americana*, was significant and far below the cut-off value of 0.170 determined by MLST scheme [52]. Sequence pair distance analysis revealed that strains SCW-30b, SCW-30c and SCW-30d were identical among themselves. Strains SCW-30f and SCW-30 g were identical between themselves and definitely belong to *B. americana* (Table 2). The cut-off value 0.170 for species determination for this scheme was

exactly the one that showed genetic distance between SCW-30b, SCW-30c and SCW-30d and *B. americana* strains. Based on analysis of concatenated sequences strain SCW-30 h represents a species distinct from that of SCW-30a, SCW-30b, SCW-30c and SCW-30d. Analysis based on individual housekeeping genes put SCW-30 h outside of *B. americana* group (Table 1).

Significant incongruence was observed in results of analysis of fast evolving and slow evolving genes or non-coding genomic loci of SCW-30a strain. This could reflect the recent recombination or re-assortment of sequence variations within SCW-30 group or another *Borrelia* population that we did not detect in this study. Analysis of concatenated housekeeping genes assigned SCW-30a to *B. americana* species, though as rather highly divergent member. This conclusion might not be definite yet.

### Phylogenetic analysis

The resulting population and consensus trees from the concordance analysis are shown in Figure 1. This is a combined analysis of fast and slow evolving, plasmid- and chromosomally-located genes and non-coding regions of 8 SCW-30 strains (excluding *ospC* and 16S-23S ITS). The reported sample-wide concordance factors on the tree (CFs) are not comparable to posterior probabilities or bootstrap support values, and thus they are not interpreted as a normal support statistic would be. They are used as information about the status of the clade in question. Values around 0.5 and above indicate that most of the trees in the sample set contain that clade, and that there is no predominant discordant clade configuration.

The type strain of *B. americana* clusters with SCW-30e forming a very well-supported clade, indicating that these are likely very similar or very recently diverged strains. SCW-30f and SCW-30 g cluster together with moderate

**Table 1 Genetic divergence among individual housekeeping genes of SCW-30 strains and estimated allelic profiles**

	<i>clpA</i>	<i>clpX</i>	<i>nifS</i>	<i>pepX</i>	<i>pyrG</i>	<i>recG</i>	<i>rplB</i>	<i>uvrA</i>
SCW-41 <sup>Ta</sup>	176 <sup>c</sup>	141	127	154	159	159	136	150
SCW-33 <sup>b</sup>	175	140	126	153	158	158	135	149
SCW-30a	175 (98.4%)	141 (99.7%)	127 (99.3%)	154 (98.6%)	159 (99.7%)	159 (99.1%)	136 (99.2%)	150 (99%)
SCW-30b	175 (98.4%)	141 (99%)	127 (99.3%)	<b>153 (97.5%)</b>	<b>159 (97.8%)</b>	<b>159 (98.2%)</b>	136 (99.5%)	<b>150 (98%)</b>
SCW-30c	175 (98.4%)	141 (99%)	127 (99.3%)	<b>153 (97.5%)</b>	<b>159 (97.8%)</b>	<b>159 (98.2%)</b>	136 (99.5%)	<b>150 (98%)</b>
SCW-30d	175 (98.4%)	141 (99%)	127 (99.3%)	<b>153 (97.5%)</b>	<b>159 (97.8%)</b>	<b>159 (98.2%)</b>	136 (99.5%)	<b>150 (98%)</b>
SCW-30e	175 (98.4%)	141 (99.8%)	127 (98.6%)	154	<b>159 (98.2%)</b>	159 (98.5%)	136 (99.8%)	<b>150 (98%)</b>
SCW-30f	176 (99.7%)	141	127	154	159	159	136	150
SCW-30 g	176 (99.7%)	141	127	154	159	159	136	150
SCW-30 h	176 (99.7%)	141 (98.4%)	3 (98.4%)	154 (98.9%)	<b>159 (97.5%)</b>	<b>159 (97.5%)</b>	135 (99.5%)	<b>150 (98%)</b>

The percentage of identity of housekeeping genes of SCW-30 strains to the closest alleles available in MLST database are presented in the Table 1.

SCW-41<sup>Ta</sup> is a type strain of *Borrelia americana* and represents subgroup A (ATCC BAA-1877; DSM 22541).

SCW-33<sup>b</sup> represents *B. americana* subgroup B.

176<sup>c</sup> -unique allele numbers of housekeeping genes in MLST database. Strain allelic profile is a composition of 8 numbers presented in a single row. Marked in bold are values that are below or equal to a cut-off value of 0.170 determined for the MLST scheme based on eight chromosomally located housekeeping genes [26].

**Table 2 Genetic divergences among SCW-30 strains based on 8 concatenated housekeeping genes**

	SCW-30a	SCW-30b	SCW-30c	SCW-30d	SCW-30e	SCW-30f	SCW-30 g	SCW-30 h	SCW-41 <sup>T</sup>
SCW-30a		98,9	98,9	98,9	98,5	99,1	99,1	<b>98</b>	99,1
SCW-30b	1,1		100	100	98,5	<b>98,3</b>	<b>98,3</b>	<b>97,7</b>	<b>98,3</b>
SCW-30c	1,1	0		100	98,5	<b>98,3</b>	<b>98,3</b>	<b>97,7</b>	<b>98,3</b>
SCW-30d	1,1	0	0		98,5	<b>98,3</b>	<b>98,3</b>	<b>97,7</b>	<b>98,3</b>
SCW-30e	1,5	1,5	1,5	1,5		98,9	98,9	<b>98,2</b>	98,9
SCW-30f	0,9	1,7	1,7	1,7	1,1		100	98,4	99,9
SCW-30 g	0,9	1,7	1,7	1,7	1,1	0		98,4	99,9
SCW-30 h	2	2,4	2,4	2,4	1,8	1,6	1,6		98,4
SCW-41 <sup>T</sup>	0,9	1,7	1,7	1,7	1,7	0,1	0,1	1,7	

SCW-41<sup>T</sup> is a type strain of *Borrelia americana* (ATCC BAA-1877; DSM 22541).

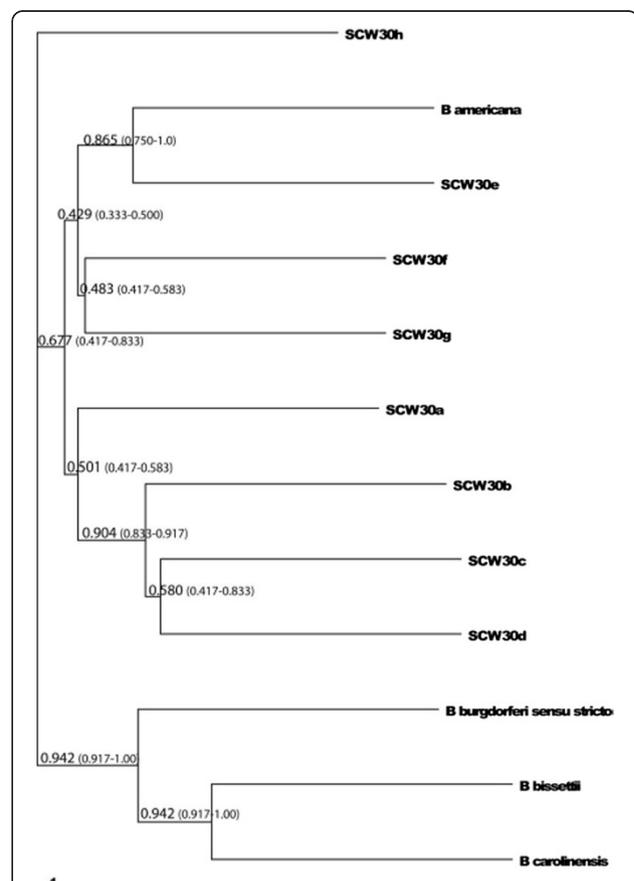
Marked in bold are values that are below or equal to a cut-off value of 0.170 determined for the MLST scheme based on eight chromosomally located housekeeping genes [26].

support, and together with *B. americana* and SCW-30e form a moderately well-supported clade “e/g/f”. Strains SCW-30b, SCW-30c and SCW-30d form an extremely well supported clade, with a concordance factor above 90%. The arrangement of SCW-30c and SCW-30d within the clade is less well supported, but the three strains clearly share histories strongly. SCW-30a clusters with SCW-30b, SCW-30c and SCW-30d, but with only moderate support forming a clade “a/b/c/d”. SCW-30 h is located outside the “a/b/c/d” and “e/g/f” clades and its placement is moderately well supported (>67% but with a wide confidence interval).

### Discussion

Comparison of SCW-30 strains with the control samples makes it clear that either there is ongoing changes among these strains, or that the divergence is very recent. The concordance among gene trees for control samples as separate and distinct clades is extremely high. However, concordance among gene trees for most of the clusters of SCW-30 strains is less clear. The clades that stand out as distinct are *B. americana* + SCW-30e, than SCW-30b, SCW-30c, SCW-30d and, SCW-30 h. Branch lengths make it clear that genetic divergence of these strains is as great as divergence among other clearly designated distinct species. Traditional rules of *Borrelia* taxonomy would support the claim that some of SCW-30 strains could have the status of new species, distinct from *B. americana*. However, the ongoing changes (from whatever source) and incongruence makes it difficult to determine if new species status should be assigned to selected SCW-30 strains, even after extended molecular analysis of 14 genomic loci of different functionality, which is the biggest known attempt other than the whole genome sequencing. The support for any cluster at this point would be ambiguous.

Local *Borrelia* populations show established biogeographic structure, often including a significant number



**Figure 1 Population tree of control *Borrelia* species and strains from SCW-30 group.** Fast and slow evolving plasmid and chromosome located genes and non-coding region were included into the analysis. Bayesian concordance analysis was conducted using BUCKY [63] with a value of 1.0 for  $\alpha$ , the parameter that summarizes prior expectations of the amount of concordance among gene trees. Each split is annotated with the posterior mean sample-wide concordance factor and its 95% credibility interval. The concordance factor shows the proportion of the sample trees for which the split is true, ranging from 0.0-1.0. Branch lengths are in coalescence units.

of distinct genomic groups and are highly diverse, often co-infect a single tick species or vertebrate host, and co-exist in sympatric fashion [44,51,66-68]. MLST and whole genome sequencing of different *Borrelia* species has confirmed that horizontal exchange of genetic information is pervasive across the *Borrelia* genome, occurs frequently between different genospecies and is more frequent than point mutations [47]. The origin of the high diversity within a local *Borrelia* population is an open subject for discussion. Several hypotheses have been proposed and one suggests that the diversity and evolution of LB spirochetes appears to be driven mainly by the host [36], and distinct clonal groups are maintained by host specialization [44,69].

The importance of birds as reservoir hosts in the ecology of LB around the world is clearly recognized today. Birds are capable of transmitting the highest number of known genotypes, albeit at different frequencies [70]. Multiple studies have confirmed the involvement of different bird species in the enzootic maintenance of *B. burgdorferi* s.l. species in endemic areas [1,10,17,40,54,71-78]. Migratory passerine birds have been shown to be responsible for spreading *Borrelia* infected ticks within and between continents, establishing new foci for Lyme borreliosis [13,16,66,77-79].

Passerines are parasitized by both tick larvae and nymphs [80]. Typically, in the northern populations of *I. scapularis* and *I. ricinus*, infected nymphs transmit spirochetes to hosts that subsequently infect larval ticks. Since the discovery of Lyme borreliosis, the evidences of transovarial transmission of *B. burgdorferi* s.l. were presented and it was believed until recently that larval ticks may, infrequently, obtain the LB spirochete by transovarial transmission, with the prevalence of less than 1% [81]. A recent literature review and observations have indicated that the transovarial transmission of the LB spirochete does not exist and was confused for years with the transovarial transmission of the antigenically and phylogenetically related *Borrelia miyamotoi* [82]. This fact leaves the only possibility for larvae to get infected with LB spirochete - the host. The relatively high prevalence of *Borrelia*-infected larvae collected from birds (from 3-3.1% [82,83] to 29% [14,83]) indicates that they became infected while feeding on birds. When the spirochetes persist in a bird for a long time, birds become an amplifier of *B. burgdorferi* s.l., transmitting the pathogen to a greater number of ticks. Larvae maintain infection through the molt, giving rise to a new population of infected host-seeking nymphs. The efficiency of this cycle leads to a high prevalence of *B. burgdorferi* s.l. infection in questing nymphs, and a high public health risk in the region [84].

The transmission of *B. burgdorferi* s.l. between ticks and vertebrate hosts is a complex process. The interaction of

*Borrelia* with the alternative pathway of the host's complement system is considered to be the key determinant of spirochete-host association [36,85-88]. As part of the innate immune system, the alternative pathway can rapidly respond to pathogens before antibodies are generated [36]. A clear pattern of resistance or sensitivity of spirochetes to host complement is correlated with patterns of transmissibility. Spirochetes that are sensitive to the complement of a particular species are lysed by the host complement in the gut of the feeding tick before they are transmitted to the hosts. Selective survival of *B. burgdorferi* s.l. in the tick midgut, depending on the source of serum and the genetic background of the bacteria, can manifest itself in the selective replacement of *Borrelia* strains during the tick life cycle [36]. The high diversity of local spirochete populations has often been connected to host specialization or to the coexistence of multiple genospecies in the region with pervasive recombination among sympatric genomes [11,47]. Multiple-niche polymorphism, a form of balancing selection, can maintain diversity within the population [44].

Analysis of the 8 *Borrelia* strains cultured from *I. minor* larvae and nymphs and reported here showed high heterogeneity among the isolates using different methods of analysis. Previous MLSA of 5 genomic loci of *B. americana* strains separated closely related members into two subgroups, A and B [32]. In this study, combined analysis that involved several methods and 14 genomic loci of SCW-30 strains showed a significant divergence among the strains isolated from 2 developmental stages of *I. minor*, feeding on a single bird. Earlier studies show that if more than one infected tick was collected from a single bird host, all ticks harbour the same spirochete species [83]. Three strains, SCW-30e, SCW-30f and SCW-30 g, cultured from *I. minor* nymphs support this observation, representing diverged strains of the same species. However, analysis of four *Borrelia* strains isolated from *I. minor* larvae indicated that they were distinct from *B. americana* and other known spirochete species at majority of genomic loci analyzed. While strains SCW-30b, SCW-30c and SCW-30d clustered together, localization of SCW-30a was incongruent over the whole spectrum of analyzed loci, indicating a probable high level of horizontal genetic exchange among all 8 strains connected to a single bird reservoir host. Even though the recombination does not often occur in a core region of the *Borrelia* genome [89], a single event of gene conversion was registered in strain SCW-30 h. A *nifS* gene from *B. burgdorferi* s.s., a species that is widely distributed in the United States and Canada, was incorporated into the homologous locus of strain SCW-30 h. This might be possible only in the presence of sympatric genome, either by transmission through a bird host that tick nymphs fed on, or by transmission through the molt stage

of larvae that fed previously on a host infected with *B. burgdorferi* ss.

A laboratory study of *I. minor* indicated that both nymphs and larvae feed for 4 days on average when fed on laboratory white mice (*Mus musculus*). Adults were reluctant to feed on the mice, but readily fed on eastern woodrats (*Neotoma floridana*) [90]. In nature *I. minor* feeds on a variety of mammals and birds including the cotton mouse, house mouse, cotton rat, cottontail rabbit, eastern rice rat, eastern grey squirrel, eastern spotted skunk, eastern woodrat, and the bird species, Carolina wren, house wren, and the eastern towhee see ref. [90], establishing the possibility of harvesting and amplification of multiple spirochete species.

We have cultured a large number of *Borrelia* strains from 8 bird species: Carolina wren (*Thryothorus ludovicianus*), downy woodpecker (*Picoides pubescens*), white-eyed vireo (*Vireo griseus*), Swainson's thrush (*Catharus ustulatus*), American redstart (*Setophaga ruticilla*), northern water thrush (*Parkesia noveboracensis*), pine warbler (*Setophaga pinus*), and northern cardinal (*Cardinalis cardinalis*) [91]. Four *I. minor* nymphs, collected from a Carolina Wren, had a chance to pick up *B. americana* from various hosts during the larval feeding or during their current nymph feeding. *I. minor* larvae collected from the same bird were feeding for the first time, picking up *Borrelia* species from the same bird host. It is not unusual that the vector, the reservoir host or LB patient maintain multiple spirochete species [92-99]. However, it is difficult to explain the unprecedented level of diversity among single-bird host-associated spirochete strains, specifically those isolated from larvae, considering the recent claim that transovarial transmission of *B. burgdorferi* sl does not exist [82].

Our previous studies of LB spirochetes showed very small if any diversity of strains isolated from different vector ticks or rodent hosts, whether it was a group of *B. burgdorferi* ss strains or distinct spirochete species [31,64,91]. *B. burgdorferi* sl species are adapted to hosts and this adaptation is driven by host complement. The dynamics of infection of the host supports the mixing of different genotypes and the horizontal exchange of genetic information. The striking divergence of *Borrelia* strains associated with a single bird reservoir that we present here supports the earlier hypothesis that vertebrate hosts are the key determinants in the diversity of Lyme disease spirochete [36].

## Conclusions

Genetically diverse *Borrelia* strains are often found within the same tick or same vertebrate host and this gives a wide opportunity for genetic exchange. We report a case of cross-species recombination that led to the incorporation of an allele of a housekeeping gene

from a *B. burgdorferi* sensu stricto strain, the primary causative agent of LB, into the homologous locus of bird-associated strain. Our results add to the recent hypothesis that recombination maintains a majority of sequence polymorphism within *Borrelia* populations due to re-assortment of pre-existing sequence variants. Even though our findings of increased diversity among 8 strains cultured from ticks that fed on a single bird could be the exception rather than the rule they support the theory that diversity and evolution of LB spirochetes is driven mainly by the host.

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

NR and MG designed the experimental scheme and carried complete laboratory work: i.e., strains cultivation and maintenance, DNA purification, PCR amplification and analysis, sequence analysis, RFLP analysis, sequence alignments, results evaluation and drafted the manuscript. NMB conducted phylogenetic analysis, evaluated results, drafted the figures and edited the manuscript. LG designed the study, evaluated results and edited the manuscript. JHO isolated bird-associated samples, cultured SCW-30 strains, evaluated results and drafted the manuscript. All authors read and approved the final version of the manuscript.

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## References

1. Gern L: *Borrelia burgdorferi* sensu lato, the agent of Lyme borreliosis: life in the wilds. *Parasite* 2008, **15**:244-247.
2. Anderson JF, Magnarelli LA, Stafford KC 3rd: Bird-feeding ticks transiently transmit *Borrelia burgdorferi* that infect Syrian hamsters. *J Wildl Dis* 1990, **26**:1-10.
3. Manweiler SA, Lane RS, Bloc WM, Morrison ML: Survey of birds and lizards for ixodid ticks (Acari) and spirochetal infection in northern California. *J Med Entomol* 1990, **27**:1011-1015.
4. McLean RG, Ubico SR, Hughes CA, Engstrom SM, Johnson RC: Isolation and characterization of *Borrelia burgdorferi* from the blood of a bird captured in the Saint Croix River Halley, USA [abstract 235]. Arlington, USA: Proceedings of the Vth International Conference on Lyme Borreliosis; 1992.
5. Olsen B, Jaenson TG, Bunikis J, Noppa L, Bergström S: The Lyme borreliosis spirochete found in ticks parasitizing sea birds [abstract 285]. Arlington, USA: Proceedings of the Vth International Conference on Lyme Borreliosis; 1992.
6. Brinkerhoff RJ, Folsom-O'Keefe CM, Tsao K, Diuk-Wasser MA: Do birds affect Lyme disease risk? Range expansion of the vector-borne pathogen *Borrelia burgdorferi*. *Front Ecol Environ* 2011, **9**:103-110.
7. Norte AC, Ramos JA, Gern L, Nuncio MS, Lopes de Carvalho I: Birds as reservoirs for *Borrelia burgdorferi* s.l. in Western Europe: circulation of *B. turdi* and other genospecies in bird-tick cycles in Portugal. *Environ Microbiol* 2013, **15**:386-397.

8. Heylen D, Tjisse E, Fonville M, Matthyssen E, Sprong H: **Transmission dynamics of *Borrelia burgdorferi* s.l. in a bird tick community.** *Environ Microbiol* 2013, **15**:663–673.
9. Ginsberg HS, Buckley PA, Balmforth MG, Zhioua E, Mitra S, Buckley FG: **Reservoir competence of native North American birds for the Lyme disease spirochete, *Borrelia burgdorferi*.** *J Med Entomol* 2005, **42**:445–449.
10. Rand PW, Lacombe EH, Smith RP, Ficker J: **Participation of birds (Aves) in the emergence of Lyme disease in southern Maine.** *J Med Entomol* 1998, **35**:270–276.
11. Hamer SA, Hickling GJ, Sidge JL, Rosen ME, Walker ED, Tsao JI: **Diverse *Borrelia burgdorferi* strains in a bird-tick cryptic cycle.** *Appl Environ Microbiol* 2011, **77**:1999–2007.
12. Olsen B, Duffy DC, Jaenson TG, Gylfe A, Bonnedahl J, Bergström S: **Transhemispheric exchange of Lyme disease spirochetes by seabirds.** *J Clin Microbiol* 1995, **33**:3270–3274.
13. Comstedt P, Bergström S, Olsen B, Garpmo U, Marjavaara L, Mejlon H, Barbour AG, Buniks J: **Migratory passerine birds as reservoirs of Lyme borreliosis in Europe.** *Emerg Infect Dis* 2006, **12**:1087–1095.
14. Poupon MA, Lommano E, Humair PF, Douet V, Rais O, Schaad M, Jenni L, Gern L: **Prevalence of *Borrelia burgdorferi* sensu lato in ticks collected from migratory birds in Switzerland.** *Appl Environ Microbiol* 2006, **72**:976–979.
15. Ogden NH, Lindsay LR, Hanincová K, Barker IK, Bigras-Poulin M, Charron DF, Heagy A, Francis CM, O'Callaghan CJ, Schwartz I, Thompson RA: **Role of migratory birds in introduction and range expansion of *Ixodes scapularis* ticks and of *Borrelia burgdorferi* and *Anaplasma phagocytophilum* in Canada.** *Appl Environ Microbiol* 2008, **74**:1780–1790.
16. Anderson JF, Johnson RC, Magnarelli LA, Hyde FW: **Involvement of birds in the epidemiology of the Lyme disease agent *Borrelia burgdorferi*.** *Infect Immun* 1986, **51**:394–396.
17. Hanincová K, Taragelová V, Koci J, Schäfer SM, Hails R, Ullmann AJ, Piesman J, Labuda M, Kurtenbach K: **Association of *Borrelia garinii* and *B. valaisiana* with songbirds in Slovakia.** *Appl Environ Microbiol* 2003, **69**:2825–2830.
18. Baranton G, Postic D, Saint Girons I, Boerlin P, Piffaretti JC, Assous M, Grimont PA: **Delineation of *Borrelia burgdorferi* sensu stricto, *Borrelia garinii* sp. nov., and group VS461 associated with Lyme borreliosis.** *Int J Syst Bacteriol* 1992, **42**:378–383.
19. Canica MM, Nato F, du Merle L, Mazie JC, Baranton G, Postic D: **Monoclonal antibodies for identification of *Borrelia afzelii* sp. nov. associated with late cutaneous manifestations of Lyme borreliosis.** *Scand J Infect Dis* 1993, **25**:441–448.
20. Chu CY, Liu W, Jiang BG, Wang DM, Jiang WJ, Zhao QM, Zhang PH, Wang ZX, Tang GP, Yang H, Cao WC: **Novel genospecies of *Borrelia burgdorferi* sensu lato from rodents and ticks in southwestern China.** *J Clin Microbiol* 2008, **46**:3130–3133.
21. Fukunaga M, Hamase A, Okada K, Nakao M: ***Borrelia tanukii* sp. nov. and *Borrelia turdae* sp. nov. found from ixodid ticks in Japan: rapid species identification by 16S rRNA gene-targeted PCR analysis.** *Microbiol Immunol* 1996, **40**:877–881.
22. Kawabata H, Masuzawa T, Yanagihara Y: **Genomic analysis of *Borrelia japonica* sp. nov. isolated from *Ixodes ovatus* in Japan.** *Microbiol Immunol* 1993, **37**:843–848.
23. Le Fleche A, Postic D, Girardet K, Peter O, Baranton G: **Characterization of *Borrelia lusitaniae* sp. nov. by 16S ribosomal DNA sequence analysis.** *Int J Syst Bacteriol* 1997, **47**:921–925.
24. Marconi RT, Liveris D, Schwartz I: **Identification of novel insertion elements, restriction fragment length polymorphism patterns, and discontinuous 23S rRNA in Lyme disease spirochetes: phylogenetic analyses of rRNA genes and their intergenic spacers in *Borrelia japonica* sp. nov. and genomic group 21038 (*Borrelia andersonii* sp. nov.) isolates.** *J Clin Microbiol* 1995, **33**:2427–2434.
25. Margos G, Hojgaard A, Lane RS, Cornet M, Fingerle V, Rudenko N, Ogden N, Aanensen DM, Fish D, Piesman J: **Multilocus sequence analysis of *Borrelia bissettii* strains from North America reveals a new *Borrelia* species, *Borrelia kurtenbachii*.** *Ticks Tick Borne Dis* 2010, **1**:151–158.
26. Margos G, Vollmer SA, Kornet M, Garnier M, Fingerle V, Wilske B, Bormane A, Vitorino L, Collares-Pereira M, Drancourt M, Kurtenbach K: **A new *Borrelia* species defined by multilocus sequence analysis of housekeeping genes.** *Appl Environ Microbiol* 2009, **75**:5410–5416.
27. Masuzawa T, Takada N, Kudeken M, Fukui T, Yano Y, Ishiguro F, Kawamura Y, Imai Y, Ezaki T: ***Borrelia sinica* sp. nov., a Lyme disease-related *Borrelia* species isolated in China.** *Int J Syst Evol Microbiol* 2001, **51**:1817–1824.
28. Postic D, Garnier M, Baranton G: **Multilocus sequence analysis of atypical *Borrelia burgdorferi* sensu lato isolates—description of *Borrelia californiensis* sp. nov., and genomospecies 1 and 2.** *Int J Med Microbiol* 2007, **297**:263–271.
29. Postic D, Ras NM, Lane RS, Hendson M, Baranton G: **Expanded diversity among Californian *Borrelia* isolates and description of *Borrelia bissettii* sp. nov. (formerly *Borrelia* group DN127).** *J Clin Microbiol* 1998, **36**:3497–3504.
30. Richter D, Postic D, Sertour N, Livey I, Matuschka FR, Baranton G: **Delineation of *Borrelia burgdorferi* sensu lato species by multilocus sequence analysis and confirmation of the delineation of *Borrelia spielmanii* sp. nov.** *Int J Syst Evol Microbiol* 2006, **56**:873–881.
31. Rudenko N, Golovchenko M, Grubhoffer L, Oliver JH Jr: ***Borrelia carolinensis* sp. nov., a new (14th) member of the *Borrelia burgdorferi* sensu lato complex from the southeastern region of the United States.** *J Clin Microbiol* 2009, **47**:134–141.
32. Rudenko N, Golovchenko M, Lin T, Gao L, Grubhoffer L, Oliver JH Jr: **Delineation of a new species of the *Borrelia burgdorferi* sensu lato complex, *Borrelia americana* sp. nov.** *J Clin Microbiol* 2009, **47**:3875–3880.
33. Wang G, van Dam AP, Le Fleche A, Postic D, Peter O, Baranton G, de Boer R, Spanjaard L, Dankert J: **Genetic and phenotypic analysis of *Borrelia valaisiana* sp. nov. (*Borrelia* genomic groups VS116 and M19).** *Int J Syst Bacteriol* 1997, **47**:926–932.
34. Casjens SR, Fraser-Liggett CM, Mongodin EF, Qiu WG, Dunn JJ, Luft BJ, Schutzer SE: **Whole genome sequence of an unusual *Borrelia burgdorferi* sensu lato isolate.** *J Bacteriol* 2011, **193**:1489–1490.
35. Urwin R, Maiden MC: **Multi-locus sequence typing: a tool for global epidemiology.** *Trends Microbiol* 2003, **11**:479–487.
36. Kurtenbach K, De Michelis S, Etti S, Schäfer SM, Sewell HS, Brade V, Kraiczy P: **Host association of *Borrelia burgdorferi* sensu lato—the key role of host complement.** *Trends Microbiol* 2002, **10**:74–79.
37. Kurtenbach K, De Michelis S, Sewell HS, Etti S, Schäfer SM, Holmes E, Hails R, Collares-Pereira M, Santos-Reis M, Hanincová K, Labuda M, Bormane A, Donaghy M: **The key roles of selection and migration in the ecology of Lyme borreliosis.** *Int J Med Microbiol* 2002, **33**:152–154.
38. Skerka C, Brade V, Zipfel PF: **Further characterization of complement regulator-acquiring surface proteins of *Borrelia burgdorferi*.** *Infect Immun* 2001, **69**:7800–7809.
39. Kraiczy P, Skerka C, Kirschfink M, Zipfel PF, Brade V: **Mechanism of complement resistance of pathogenic *Borrelia burgdorferi* isolates.** *Int Immunopharmacol* 2001, **1**:393–401.
40. Kurtenbach K, Schäfer SM, Sewell HS, Peacey M, Hoodless A, Nuttall PA, Randolph SE: **Differential survival of Lyme borreliosis spirochetes in ticks that feed on birds.** *Infect Immun* 2002, **70**:5893–5895.
41. Stevenson B, El-Hage N, Hines MA, Miller JC, Babb K: **Differential binding of host complement inhibitor factor H by *Borrelia burgdorferi* Erp surface proteins: a possible mechanism underlying the expansive host range of Lyme disease spirochetes.** *Infect Immun* 2002, **70**:491–497.
42. Eggers CH, Kimmel BJ, Bono JL, Elias AF, Rosa P, Samuels DS: **Transduction by phiBB-1, a bacteriophage of *Borrelia burgdorferi*.** *J Bacteriol* 2001, **183**:4771–4778.
43. Kurtenbach K, Hanincová K, Tsao JI, Margos G, Fish D, Ogden NH: **Fundamental processes in the evolutionary ecology of Lyme borreliosis.** *Nat Rev Microbiol* 2006, **4**:660–669.
44. Brisson D, Dykhuizen DE: **ospC diversity in *Borrelia burgdorferi*: different hosts are different niches.** *Genetics* 2004, **168**:713–722.
45. Brisson D, Vandermause MF, Meece JK, Reed KD, Dykhuizen DE: **Evolution of northeastern and midwestern *Borrelia burgdorferi*, United States.** *Emerg Infect Dis* 2010, **16**:911–917.
46. Guttman DS, Wang PW, Wang IN, Bosler EM, Luft BJ, Dykhuizen DE: **Multiple infections of *Ixodes scapularis* ticks by *Borrelia burgdorferi* as revealed by single-strand conformation polymorphism analysis.** *J Clin Microbiol* 1996, **34**:652–656.
47. Haven J, Vargas LC, Mongodin EF, Xue V, Hernandez Y, Pagan P, Fraser-Liggett CM, Schutzer SE, Luft BJ, Casjens SR, Qiu WG: **Pervasive recombination and sympatric genome diversification driven by frequency-dependent selection in *Borrelia burgdorferi*, the Lyme disease bacterium.** *Genetics* 2011, **189**:951–966.
48. Postic D, Assous MV, Grimont PA, Baranton G: **Diversity of *Borrelia burgdorferi* sensu lato evidenced by restriction fragment length polymorphism of rrf (5S)-rrl (23S) intergenic spacer amplicons.** *Int J Syst Bacteriol* 1994, **44**:743–752.

49. Güner ES, Hashimoto N, Takada N, Kaneda K, Omak Y, Masuzawa T: **First isolation and characterization of *Borrelia burgdorferi* sensu lato strains from *Ixodes ricinus* ticks in Turkey.** *J Med Microbiol* 2003, **52**:807–813.
50. Clark K, Hendricks A, Burge D: **Molecular identification and analysis of *Borrelia burgdorferi* sensu lato in lizards in the southeastern United States.** *Appl Environ Microbiol* 2005, **71**:2616–2625.
51. Bunikis J, Garpmo U, Tsao J, Berglund J, Fish D, Barbour AG: **Sequence typing reveals extensive strain diversity of the Lyme borreliosis agents *Borrelia burgdorferi* in North America and *Borrelia afzelii* in Europe.** *Microbiology* 2004, **150**:1741–1755.
52. Margos G, Gatewood AG, Aanensen DM, Hanincová K, Terekhova D, Vollmer SA, Cornet M, Piesman J, Donaghy M, Bormane A, Hurn MA, Feil EJ, Fish D, Casjens S, Wormser GP, Schwartz I, Kurtenbach K: **MLST of housekeeping genes captures geographic population structure and suggests a European origin of *Borrelia burgdorferi*.** *Proc Natl Acad Sci USA* 2008, **105**:8730–8735.
53. Bikandi J, San Millán R, Rementería A, Garaizar J: **In silico analysis of complete bacterial genomes: PCR, AFLP-PCR and endonuclease restriction.** *Bioinformatics* 2004, **20**:798–799.
54. Hoogstraal H: **Birds as tick hosts and as reservoirs and disseminators of tickborne infectious agents.** *Wiad Parazytol* 1972, **18**:703–706.
55. Larkin MA, Blackshields G, Brown NP, Chenna R, McGettigan PA, McWilliam H, Valentin F, Wallace JM, Wilm A, Lopez R, Thompson JD, Gibson TJ, Higgins DG: **Clustal W and Clustal X version 2.0.** *Bioinformatics* 2007, **23**:2947–2948.
56. Nylander JAA: *MrModeltest v2.3. Program distributed by the author.* Uppsala, Sweden: Evolutionary Biology Centre: Uppsala University; 2008.
57. Akaike H: **A new look at the statistical model identification.** *IEEE Trans Autom Contr* 1974, **19**:716–723.
58. Huelsenbeck JP, Ronquist F, Nielsen R, Bollback JP: **Bayesian inference of phylogeny and its impact on evolutionary biology.** *Science* 2001, **294**:2310–2314.
59. Ronquist F, Huelsenbeck JP: **MrBayes 3: Bayesian phylogenetic inference under mixed models.** *Bioinformatics* 2003, **19**:1572–1574.
60. Nylander JAA, Wilgenbusch JC, Warren DL, Swofford DL: **AWTY (are we there yet?): a system for graphical exploration of MCMC convergence in Bayesian phylogenetics.** *Bioinformatics* 2008, **24**:581–583.
61. Larget BR, Kotha SK, Dewey CN, Ané C: **BUCKY: gene tree/species tree reconciliation with Bayesian concordance analysis.** *Bioinformatics* 2010, **26**:2910–2911.
62. Allman ES, Degnan JH, Rhodes JA: **Identifying the rooted species tree from the distribution of unrooted gene trees under the coalescent.** *J Math Biol* 2011, **62**:833–862.
63. Ané C, Larget B, Baum DA, Smith SD, Rokas A: **Bayesian estimation of concordance among gene trees.** *Mol Biol Evol* 2007, **24**:412–426.
64. Rudenko N, Golovchenko M, Hönig V, Mallátová N, Krbková L, Mikulásek P, Fedorova N, Belfiore NM, Grubhoffer L, Lane RS, Oliver JH Jr: **Detection of *Borrelia burgdorferi* sensu stricto ospC alleles associated with human Lyme borreliosis worldwide in non-human-biting tick *Ixodes affinis* and rodent hosts in southeastern United States.** *Appl Environ Microbiol* 2013, **79**:1444–1453.
65. Aanensen DM, Spratt BG: **The multilocus sequence typing network: mlst.net.** *Nucleic Acids Res* 2005, **33**(Web Server issue):W728–W733.
66. Margos G, Vollmer SA, Ogdén NH, Fish D: **Population genetics, taxonomy, phylogeny and evolution of *Borrelia burgdorferi* sensu lato.** *Infect Genet Evol* 2011, **11**:1545–1563.
67. Qiu WG, Dykhuizen DE, Acosta MS, Luft BJ: **Geographic uniformity of the Lyme disease spirochete (*Borrelia burgdorferi*) and its shared history with tick vector (*Ixodes scapularis*) in the northeastern United States.** *Genetics* 2002, **160**:833–849.
68. Wang IN, Dykhuizen DE, Qiu W, Dunn JJ, Bosler EM, Luft BJ: **Genetic diversity of ospC in a local population of *Borrelia burgdorferi* sensu stricto.** *Genetics* 1999, **151**:15–30.
69. Brisson D, Dykhuizen DE: **A modest model explains the distribution and abundance of *Borrelia burgdorferi* strains.** *Am J Trop Med Hyg* 2006, **74**:615–622.
70. Brinkerhoff RJ, Bent SJ, Folsom-O'Keefe CM, Tsao K, Hoen AG, Barbour AG, Diuk-Wasser MA: **Genotypic diversity of *Borrelia burgdorferi* strains detected in *Ixodes scapularis* larvae collected from North American songbirds.** *Appl Environ Microbiol* 2010, **76**:8265–8268.
71. Humair PF, Postic D, Wallich R, Gern L: **An avian reservoir (*Turdus merula*) of the Lyme borreliosis spirochetes.** *Zentralbl Bakteriell* 1998, **287**:521–538.
72. Ishiguro F, Takada N, Masuzawa T, Fukui T: **Prevalence of Lyme disease *Borrelia* spp. in ticks from migratory birds on the Japanese mainland.** *Appl Environ Microbiol* 2000, **66**:982–986.
73. Kurtenbach K, Peacey M, Rijpkema SG, Hoodless AN, Nuttall PA, Randolph SE: **Differential transmission of the genospecies *Borrelia burgdorferi* sensu lato by game birds and small rodents in England.** *Appl Environ Microbiol* 1998, **64**:1169–1174.
74. Miyamoto K, Nakao M, Fujita H, Sato F: **The Ixodid ticks on migratory birds in Japan and the isolation of Lyme disease spirochetes from bird-feeding ticks.** *Jpn J Sanit Zool* 1993, **44**:315–326.
75. Miyamoto K, Sato Y, Okada K, Fukunaga M, Sato F: **Competence of a migratory bird, red-bellied thrush (*Turdus chrysolaus*), as an avian reservoir for the Lyme disease spirochetes in Japan.** *Acta Trop* 1997, **65**:43–51.
76. Nakao M, Miyamoto K, Fukunaga M: **Lyme disease spirochetes in Japan: enzootic transmission cycles in birds, rodents, and *Ixodes persulcatus* ticks.** *J Infect Dis* 1994, **170**:878–882.
77. Smith RP Jr, Rand PW, Lacombe EH, Morris SR, Holme DW, Caporale DA: **Role of bird migration in the long-distance dispersal of *Ixodes dammini*, the vector of Lyme disease.** *J Infect Dis* 1996, **174**:221–224.
78. Weisbrod AR, Johnson RC: **Lyme disease and migrating birds in the Saint Croix River Valley.** *Appl Environ Microbiol* 1989, **55**:1921–1924.
79. Morshed MG, Scott JD, Fernando K, Beati L, Mazerolle DF, Geddes G, Durden LA: **Migratory songbirds disperse ticks across Canada, and first isolation of the Lyme disease spirochete, *Borrelia burgdorferi*, from the avian tick, *Ixodes auritulus*.** *J Parasitol* 2005, **91**:780–790.
80. Humair PF, Turrian N, Aeschlimann A, Gern L: ***Ixodes ricinus* immatures on birds in a focus of Lyme borreliosis.** *Folia Parasitol (Praha)* 1993, **40**:237–242.
81. Olsén B, Jaenson TG, Bergström S: **Prevalence of *Borrelia burgdorferi* sensu lato-infected ticks on migrating birds.** *Appl Environ Microbiol* 1995, **61**:3082–3087.
82. Rollend L, Fish D, Childs JE: **Transovarial transmission of *Borrelia* spirochetes by *Ixodes scapularis*: a summary of the literature and recent observations.** *Ticks Tick Borne Dis* 2013, **4**:46–51.
83. Kipp S, Goedecke A, Dorn W, Wilske B, Fingerle V: **Role of birds in Thuringia, Germany, in the natural cycle of *Borrelia burgdorferi* sensu lato, the Lyme disease spirochaete.** *Int J Med Microbiol* 2006, **40**(296):125–128.
84. Yuval B, Spielman A: **Duration and regulation of the developmental cycle of *Ixodes dammini* (Acari: Ixodidae).** *J Med Entomol* 1990, **27**:196–201.
85. Kuo MM, Lane RS, Giclas PC: **A comparative study of mammalian and reptilian alternative pathway of complement-mediated killing of the Lyme disease spirochete (*Borrelia burgdorferi*).** *J Parasitol* 2000, **86**:1223–1228.
86. Lane RS, Quistad GB: **Borreliacidal factor in the blood of the western fence lizard (*Sceloporus occidentalis*).** *J Parasitol* 1998, **84**:29–34.
87. Nelson DR, Rooney S, Miller NJ, Mather TN: **Complement-mediated killing of *Borrelia burgdorferi* by non-immune sera from sika deer.** *J Parasitol* 2000, **86**:1232–1238.
88. Van Dam AP, Oei A, Jaspars R, Fijen C, Wilske B, Spanjaard L, Dankert J: **Complement-mediated serum sensitivity among spirochetes that cause Lyme disease.** *Infect Immun* 1997, **65**:1228–1236.
89. Morlon H, Kempes BD, Plotkin JB, Brisson D: **Explosive radiation of a bacterial species group.** *Evolution* 2012, **66**:2577–2586.
90. Banks CW, Oliver JH Jr, Phillips JB, Clark KL: **Life cycle of *Ixodes minor* (Acari: Ixodidae) in the laboratory.** *J Med Entomol* 1998, **35**:496–499.
91. Rudenko N, Golovchenko M, Grubhoffer L, Oliver JH Jr: **The rare ospC allele L of *Borrelia burgdorferi* sensu stricto, commonly found among samples collected in a coastal plain area of the southeastern United States, is associated with *Ixodes affinis* ticks and local rodent hosts *Peromyscus gossypinus* and *Sigmodon hispidus*.** *Appl Environ Microbiol* 2013, **79**:1403–1406.
92. Beklemishev AB, Dobrotvorsky AK, Piterina AV, Ivanov ID, Nomokonova NY, Livanova NN: **Detection and typing of *Borrelia burgdorferi* sensu lato genospecies in *Ixodes persulcatus* ticks in West Siberia, Russia.** *FEMS Microbiol Lett* 2003, **227**:157–161.
93. Chu CY, He J, Wang JB, Hasen GW, Zhang PH, Wu XM, Zhao QM, Jiang BG, Gao Y, Cao WC: **Investigation on *Borrelia burgdorferi* sensu lato in ticks and rodents collected in Da Xing-An Mountains Forest areas of China.** *Zhonghua Liu Xing Bing Xue Za Zhi* 2006, **27**:681–684.
94. Demaerschalck I, Ben Messaoud A, De Kesel M, Hoyois B, Lobet Y, Hoet P, Bigaignon G, Bollen A, Godfroid E: **Simultaneous presence of different**

*Borrelia burgdorferi* genospecies in biological fluids of Lyme disease patients. *J Clin Microbiol* 1995, **33**:602–608.

95. Fichet-Calvet E, Jomaa I, Ben Ismail R, Ashford RW: Patterns of infection of haemoparasites in the fat sand rat, *Psammomys obesus*, in Tunisia, and effect on the host. *Ann Trop Med Parasitol* 2000, **94**:55–68.
96. Oksi J, Marjamäki M, Koski K, Nikoskelainen J, Viljanen MK: Bilateral facial palsy and meningitis caused by *Borrelia* double infection. *Lancet* 1995, **345**:1583–1584.
97. Rudenko N, Golovchenko M, Růžek D, Piskunova N, Mallátová N, Grubhoffer L: Molecular detection of *Borrelia bissetii* DNA in serum samples from patients in the Czech Republic with suspected borreliosis. *FEMS Microbiol Lett* 2009, **292**:274–281.
98. Rudenko N, Golovchenko M, Nemeč J, Volkaert J, Mallátová N, Grubhoffer L: Improved method of detection and molecular typing of *Borrelia burgdorferi* sensu lato in clinical samples by polymerase chain reaction without DNA purification. *Folia Microbiol (Praha)* 2005, **50**:31–39.
99. Vennestrøm J, Egholm H, Jensen PM: Occurrence of multiple infections with different *Borrelia burgdorferi* genospecies in Danish *Ixodes ricinus* nymphs. *Parasitol Int* 2008, **57**:32–37.

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