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The impact of different sprayable surfaces on the effectiveness of indoor residual spraying using a micro encapsulated formulation of lambda-cyhalothrin against *Anopheles gambiae s.s.*

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Abstract

Background: The type of sprayable surface impacts on residual efficacy of insecticide used in indoor residual spraying (IRS). However, there is limited data on common types of wall surfaces sprayed in Zanzibar and mainland Tanzania where IRS began in 2006 and 2007 respectively. The study investigated residual efficacy of micro-encapsulated lambda-cyhalothrin sprayed on common surfaces of human dwellings and domestic animal shelters in Zanzibar and mainland Tanzania.

Methods: An experimental hut was constructed with different types of materials simulating common sprayable surfaces in Zanzibar and mainland Tanzania. Surfaces included cement plastered wall, mud-daub, white-wash, wood, palm-thatch, galvanized iron-sheets, burnt-bricks, limestone and oil-paint. The World Health Organization (WHO) procedure for IRS was used to spray lambda-cyhalothrin on surfaces at the dose of 20–25 mg/m². Residual efficacy of insecticide was monitored through cone bioassay using laboratory-reared mosquitoes; Kisumu strain (R-70) of *Anopheles gambiae ss.* Cone bioassay was done every fortnight for a period of 152 days. The WHO Pesticide Evaluation Scheme (WHOPES) threshold (80% mortality) was used as cut-off point for acceptable residual efficacy.

Results: A total of 5,800 mosquitoes were subjected to contact cone bioassay to test residual efficacy of lambda-cyhalothrin. There was a statistically significant variation in residual efficacy between the different types of wall surfaces ($r = 0.24$; $p < 0.001$). Residual efficacy decreased with increasing pH of the substrate ($r = -0.5$; $p < 0.001$). Based on WHOPES standards, shorter residual efficacy (42–56 days) was found in wall substrates made of cement, limestone, mud-daub, oil paint and white wash. Burnt bricks retained the residual efficacy up to 134 days while galvanized iron sheets, palm thatch and wood retained the recommended residual efficacy beyond 152 days.

Conclusion: The study revealed a wide variation in residual efficacy of micro encapsulated formulation of lambda-cyhalothrin across the different types of wall surfaces studied. In areas where malaria transmission is bimodal and wall surfaces with short residual efficacy comprise > 20% of sprayable structures, two rounds of IRS using lambda-cyhalothrin should be considered. Further studies are required to investigate the impact of sprayable surfaces on residual efficacy of other insecticides commonly used for IRS in Zanzibar and mainland Tanzania.

Keywords: Indoor residual spraying, Lambda-cyhalothrin, Wall surfaces, *Anopheles gambiae ss*, Mainland Tanzania, Zanzibar

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Background

Over the last decade, many countries including Tanzania have strengthened malaria control using a combination of both insecticide treated mosquito nets (ITNs) and indoor residual spraying (IRS) [1]. IRS remains a powerful vector control tool for reducing and interrupting malaria transmission in malaria endemic areas. In Africa, more than 77 million people benefited from the intervention in 2011 and the number continue to increase [2]. IRS is one of the core interventions to control malaria vectors both in Zanzibar and mainland Tanzania [3,4]. Between 2006 and 2012, three to six blanket rounds of IRS were implemented, covering 85 – 90% of all eligible structures in Zanzibar and three regions (Kagera, Mwanza and Mara) of mainland Tanzania [5]. It was during this period that the areas where IRS was implemented registered dramatic decline of malaria prevalence from 25% to less than 1% in Zanzibar, 41% to 8% in Kagera, 31.4% to 19% in Mwanza and 30.3% to 25% in Mara [6]. This decline was also attributed to the implementation of long lasting insecticidal nets (LLIN), and malaria case management with malaria rapid diagnostic tests (mRDT) and artemisinin combination therapy (ACTs).

There is strong evidence that supports the efficacy and effectiveness of IRS in malaria control in countries where it was implemented appropriately [2,7]. The intervention has potential to significantly reduce malaria morbidity and mortality and accelerate progress towards global and national malaria control targets [8,9]. Lambda-cyhalothrin (both wettable and micro encapsulated formulations) has been widely used for IRS in Zanzibar and mainland Tanzania. The choice of lambda-cyhalothrin was based on the good profile of the insecticide and baseline studies that supported high susceptibility of *An. Gambiae s.l.* [10]. The other criteria used to select lambda-cyhalothrin were its comparatively low cost compared to other insecticides. In addition, despite its high toxicity to bees and fish, it has low mammalian toxicity [11].

Residual efficacy of an insecticide on the sprayed surfaces is an important factor that influences the effectiveness of IRS [12]. The insecticide should be sufficiently stable to maintain biological efficacy on treated surfaces over time in order to minimize the number of spraying cycles required to cover the targeted malaria transmission seasons [2]. An insecticide for IRS is considered to have adequate residual efficacy when mortality of the exposed mosquitoes is $\geq 80\%$ at 24 hours post-exposure [13]. Factors influencing efficacy include: mosquito susceptibility to insecticide; mosquito behavior (endophilic and endophagic); type of sprayable surfaces; quality of IRS; and community acceptance [14,15]. Other factors have been reported to contribute to the residual efficacy of an insecticide, such as: the type of insecticide; formulation; applied dose; physical and chemical properties of the sprayed surfaces; and weather conditions [16-18].

Varying residual efficacy of lambda-cyhalothrin (ICON® 10CS) has been reported ranging between 2 – 7 months on various surfaces [19-23]. Anecdotal entomological findings suggested that residual efficacy of lambda-cyhalothrin varied with different types of sprayable wall surfaces [5]. In Zanzibar and mainland Tanzania, IRS monitoring data revealed various types of sprayable surfaces. However, there are limited studies on the influence of different wall substrate on the residual efficacy of insecticides commonly used for IRS in Zanzibar and mainland Tanzania. The study investigated residual efficacy of micro-encapsulated lambda-cyhalothrin sprayed on common surfaces of human dwellings and domestic animal shelters in Zanzibar and Mainland Tanzania.

Methods

Study site

The study was conducted at National Institute for Medical Research (NIMR) Mwanza center in Northern Tanzania. An experimental hut was constructed assembling nine different surfaces which are commonly sprayed structures of human dwellings and domestic animal shelters both, in Zanzibar and mainland Tanzania (Figure 1).

Study design

In the experimental study design, treatment and control groups were used. The experimental hut was divided into two rooms and each room comprised 9 substrates plastered in the inside walls (Figure 1A-C). One room contained treated units that were replicated twice and the other room contained one surface of each study unit as the control. The substrates were fixed on the walls of the hut and demarcated to guide the spraying movement (Figure 1D). Side edges were used during spraying to avoid overlap effect (Figure 1E). The treated room also comprised of Whatman paper hinged on wood after being sprayed on tested surfaces (Figure 1F). Units of the study were fabricated surfaces (1.0 m wide and 2.5 m high) of limestone blocks, burnt bricks, white wash, galvanized iron sheet, cement plastered wall, mud daub, wood, palm thatch and oil paint.

Determination of substrates pH

Before spraying, the study substrates were tested for pH which was assumed to be one of the possible causes for accelerating insecticide decay. The corrugated iron sheet was not tested due to methodological limitations. The measurements were done using a hand held pH meter (HK 3C 8818) according to methods developed by Campbell and Bryant [24]. Using a scalpel, the substrate was scraped off from wall surfaces of white wash, burnt bricks, limestone blocks, cement blocks and mud daub. The materials obtained were sieved using a domestic sifter used to sieve flour. For the wood and palm thatch,



Figure 1 Design of the experimental hut and procedure used for spraying. **A.** An experimental hut with two rooms was constructed assembling nine different surfaces. **B.** Surfaces of wood, iron sheet, burnt bricks, limestone blocks and cement plastered walls are shown. **C.** Surfaces of palm thatch, mud daub and oil paint are shown. **D.** The substrates were fixed on the walls of the hut and demarcated to guide the spraying movement. **E.** Spraying on the tested surfaces was done using side edges to avoid overlap effect. **F.** Whatman paper was hinged on wood after being sprayed on tested surfaces.

the experiment used powder obtained while shredding the materials using a hand saw. Each sample was replicated twice. The substrate and distilled water were mixed in a jar with a ratio of 1:5 weights by volume (20 g: 100 ml). The mixture was shaken for five minutes to allow leaching of ions and then left to settle for ten minutes after which pH was measured.

Spraying of the surfaces with lambda-cyhalothrin

The surfaces were sprayed with lambda-cyhalothrin (ICON 10CS®). Before spraying, the surfaces were marked with three lines, one in the middle where the nozzle was pointing from 45 cm off the wall while spraying. The other two lines were 27.5 cm from the middle on either side. The marking identified the area of optimal concentration of the swathe (55 cm), avoiding 10 cm in each side that receives fewer doses [14]. The spraying was done to attain the dose of 20 – 25 mg/m² which is within the recommended range for malaria vector control [25]. The spraying used Hudson® hand compression sprayers with a flat nozzle (SS 8002), which is recommended for the application of public health insecticides [26]. Pressure in the pump ranged between 1.8 – 4 kg/m². Distance from the sprayed wall was maintained at 45 cm, while up and down movement was at 2.2 seconds per meter.

Insecticide residue analysis

Prior to spraying, three pieces of filter papers (Whatman® paper No 1) were pinned on each surface: one on the upper (130 – 250 cm upper); the second in the middle (125 cm middle); and the third on the lower segment (1 – 20 cm). A total of 54 filter paper pieces were pinned (3 pieces per wall × 2 replicates × 9 surfaces). After spraying, 18 pieces of filter paper were randomly selected for

laboratory residual analysis to determine the amount of insecticide that was deposited on the walls through spraying procedure. Only 1/3 of the strips (18/54) were randomly selected and analyzed for the amount of insecticide available, due to high cost of analyzing filter paper strips (40\$ per piece). The remaining 36 were hinged on the wood surface and kept in the experimental hut for cone bioassay to expose them to the same environmental conditions as the tested substrates. The natural decay of insecticide was determined on the filter papers since it exerts minimum physical and chemical influence on the insecticide and was thus used as a positive reference control [27]. Capillary Gas Chromatograph (Agilent 7890A Gas Chromatography Mass Spectrophotometer) was used to determine residues on the sprayed filter papers.

Mosquito rearing

Mosquito rearing was routinely undertaken by a qualified insectary attendant at NIMR, Mwanza. The purpose of rearing mosquitoes was to raise a colony of mosquitoes to cope with the demand of cone bioassays, synchronize their age structure so that experiments would be done with mosquitoes of same age to ensure reliability and reproducibility of data [28].

Cone bioassay

A laboratory-reared Kisumu strain (R-70) of *An. gambiae s.s* was subjected to contact bioassays on different sprayed surfaces. Two-day old, unfed *An. gambiae s.s* females were siphoned out of a cage using mouth-operated aspirators and gently blown into each cone bioassay fixed on the test material with the aid of masking tape and/or elastic bands [29]. Each cone had a maximum of 20 female mosquitoes. After exposing mosquitoes for 30 minutes, they were

siphoned out of the cone and blown into a holding paper cup, fed on 10% glucose and kept at room temperature. Mortality was scored after 1 hour and 24 hours post exposure. Any mortality occurring within 1 hour post exposure was scored as knock-down [28]. Cone bioassay was done every fortnight throughout 152 days using 20 mosquitoes per cone.

Data analysis

Two sets of data (pH of substrates and mosquito mortality) were analyzed using SPSS® (Statistical Package for Social Science version 18) and Microsoft Excel®. Regression analysis model was used to predict the mortality rate of the mosquitoes from day 1 to day 152 for each substrate. Analysis of Variance (ANOVA) was used to test the statistical significance of the differences in mean mosquito mortality rate. Multiple analyses of variance (MANOVA) was used to perform multiple comparisons for the tested surfaces. Whatman® paper was used as a reference substrate against other substrates as it was considered to have very minimal physical and chemical influence of efficacy of lambda-cyhalothrin.

Ethical consideration

The study protocol was approved by the institutional review committee of University of Dar es Salaam. The study did not involve any human subjects or animals.

Results

Concentration of lambda-cyhalothrin

Table 1, shows the estimated dose of insecticide that was deposited on the wall substrates based on quantification from 18 randomly selected filter papers that were pinned on the substrates prior to spraying. The results indicate that, the concentration of insecticide sprayed on the wall substrates was within the recommended range for IRS of 20 – 25 mg/m².

Residual efficacy of lambda-cyhalothrin

The residual efficacy of lambda-cyhalothrin on tested wall substrates was monitored for a period of 152 days. There was variation in residual efficacy between the different types of wall surfaces ($r = 0.24$; $p < 0.001$).

Decay of residual efficacy

Different patterns of residual efficacy decay were observed among the wall substrates tested as shown in Figures 2 and Table 2. The results are presented using regression curves that show the rate of insecticide decay versus number of days post spraying (Figure 2). Based on World Health Organization Pesticide Evaluation Scheme (WHOPES) standards, shorter residual efficacy (42–56 days) was found in wall substrates made of cement, limestone, mud-daub, oil paint and white wash (Figure 2A). However, burnt bricks retained the residual efficacy up to 134 days while walls made of iron sheets, palm thatch and wood, retained the recommended efficacy of $\geq 80\%$ mortality beyond 152 days (Figure 2B). For the untreated control groups, the mosquito mortality was 0% for all tested substrates.

Association of mosquito mortality and pH of surfaces

Table 3 shows the results of pH values on each tested surface except galvanized iron sheet. The pH of the tested surfaces ranged between 5.9 – 11. Increasing pH was associated with decreasing mean mosquito mortality rate ($r = -0.5$; $p < 0.001$).

Discussion

The study investigated residual efficacy of micro encapsulated formulation of lambda-cyhalothrin (at the dose of 20 – 25 mg/m²) on wall substrates comprising mud daub, limestone blocks, cement blocks, white wash and oil paint. Based on WHOPES recommendations, an ideal insecticide should have a minimum effect of $\geq 80\%$ mosquito mortality at 24 hours post exposure on the sprayed

Table 1 Lambda-cyhalothrin concentration levels on the sprayed substrates

Substrate	Sample location*	mg/l	mg/m ²	Substrate	Sample location*	mg/l	mg/m ²
Cement blocks	B	2.22E-02	22	Limestone blocks	F	2.48E-02	25
Cement blocks	E	2.38E-02	24	Limestone blocks	A	2.18E-02	22
Cement blocks	F	2.23E-02	22	Iron sheet	C	2.20E-02	22
Mud daub	A	2.30E-02	23	Burnt bricks	B	2.19E-02	22
Mud daub	E	2.47E-02	25	Palm thatch	A	2.51E-02	25
Wood	B	2.15E-02	22	Palm thatch	D	2.23E-02	22
Wood	D	2.39E-02	24	White wash	C	2.32E-02	23
Oil paint	A	2.00E-02	20	White wash	A	2.36E-02	24
Oil paint	D	2.01E-02	20	White wash	B	2.17E-02	22

*The sample location refers to the level where the whatman paper was located. A = Upper, B = Middle, C = Lower levels of the 1st replicate; D = Upper, E = Middle, F = Lower levels of the 2nd replicate.

Table 3 Relationship between mean mosquito mortality rate and pH of the tested surfaces

Substrate type	Mean pH	No. of mosquitoes exposed over 10 time points*	Mean mortality	Std. deviation
Burnt Bricks	7.64	400	18	±1.631
Cement Blocks	10.3	400	14	±4.989
Oil Paint	10.28	400	14	±4.124
Mud daub	7.71	400	13	±3.829
White wash	11.5	400	14	±3.771
Palm thatch	5.95	400	19	±1.338
Limestone Blocks	10.21	400	13	±5.675
Wood	7.3	400	20	±0.733
Whatman® paper (standard)	7	400	20	±0.0

*2 replicates of 20 mosquitoes per time period.

including limestone blocks that were transported from Pemba to Mwanza. However, the study had a number of potential limitations. It was not possible to monitor residual efficacy beyond 152 days; therefore, the complete period of effective residual efficacy of lambda-cyhalothrin on different wall substrates could not be established. Our study design used 20 mosquitos per cone during the bio-assay. This number is higher than that used in similar studies. It has been argued that too many mosquitoes per cone may increase mortality. However, we do not expect this to have influenced mortality since a similar number of mosquitoes were used for the controls and mortality was 0 for all controls.

Our findings provide important evidence for programmatic decision making. Lambda-cyhalothrin (wetttable powder [WP] and micro encapsulated [CS]) has been widely used for IRS to control malaria vectors in mainland Tanzania and Zanzibar. For these IRS operations, the residual effect of the insecticide was largely considered to be 6–9 months as per the manufacturers recommendations [4]. However, this perception and practice was not ideal, since findings of the study revealed a much shorter residual effect of 2–3 months on most surfaces that are common in mainland Tanzania and Zanzibar. These findings suggest that similar residual efficacy studies should be recommended to investigate if insecticides for IRS provide sufficient protection, especially when new insecticides or formulations are introduced.

In mainland Tanzania (Kagera, Mwanza and Mara regions), mud walls (81%) and cement brick walls (17%) constitute a large proportion of sprayable surfaces [5]. Based on our findings, implementation of two rounds of IRS with micro encapsulated formulation of lambda-cyhalothrin (ICON 10CS) should be considered where mud daub, cement blocks, limestone blocks, white-wash and oil painted walls comprise a significant proportion, and where malaria transmission is bimodal. Furthermore, since pH has significant influence on the residual efficacy of lambda-cyhalothrin, assessment done prior to

IRS should measure pH of a sample of substrates of targeted house structures. These data can help to predict the impact of pH on IRS using lambda-cyhalothrin. Further research on the influence of common sprayable substrates on the effectiveness of IRS using other insecticides registered for use such as carbamates, organophosphates and organochlorines should be considered.

Conclusion

The study revealed a wide variation in residual efficacy of micro encapsulated formulation of lambda-cyhalothrin across the different types of wall surfaces studied. In areas where malaria transmission is bimodal and wall surfaces of short residual efficacy comprise >20% of sprayable structures, two rounds of IRS using lambda-cyhalothrin should be considered. Further studies are required to investigate the impact of sprayable surfaces on residual efficacy of other insecticides commonly used for IRS in Zanzibar and mainland Tanzania.

Abbreviations

ACT: Artemisinin combination therapy; LLIN: Long lasting insecticide nets; IRS: Indoor residual spraying; ITN: Insecticide treated mosquito nets; WP: Wetttable powder; CS: Micro-encapsulated; ANOVA: Analysis of variance; MANOVA: Multiple analysis of variance; WHO: World Health Organization; WHOPES: World Health Organization pesticide evaluation scheme.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

JM, JNI, HBP, FM, MMR, JMK and JGR designed the study. JM, FEM, ECN, JGR and CDM undertook fieldwork. JM, ECN, OM, JMN undertook data analysis. JM, JMN, SMM, IN, RR and NGT drafted the manuscript which all authors reviewed and approved.

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References

- Mboera LEG, Mazigo HD, Rumisha SF, Kramer RA. Towards malaria elimination and its implication for vector control, disease management and livelihoods in Tanzania. *MWJ*. 2013;4(19):1-14.
- World Health Organization. Global Malaria Programme: World malaria report 2012. Geneva: World Health Organization; 2012.
- Ministry of Health, Zanzibar Malaria Control Program. Zanzibar strategic plan for malaria control 2007–2012. Zanzibar; 2007.
- Ministry of Health and Social Welfare, National Malaria Control Program. Medium Strategic Plan 2008-2013. Dar es Salaam; 2008.
- Research Triangle Institute International. Tanzania Vector Control Scale-Up Project: Spray Performance Report Nov 2011-May 2012. North Carolina: RTI International; 2012.
- Tanzania Commission for AIDS (TACAIDS), Zanzibar AIDS Commission (ZAC), National Bureau of Statistics (NBS), Office of the Chief Government Statistician (OCGS), and ICF International. Tanzania HIV/AIDS and Malaria Indicator Survey 2011–12. Dar es Salaam, Tanzania: TACAIDS, ZAC, NBS, OCGS, and ICF International; 2013.
- Pluess B, Tanser FC, Lengeler C, Sharp BL. Indoor residual spraying for preventing malaria. *Cochrane Database Syst Rev*. 2010;4:CD006657.
- Aregawi M, Cibulskis RE, Lynch M, Williams R, World Health Organization, Global Malaria Programme. World malaria report 2011. Geneva: World Health Organization; 2011.
- Kigozi R, Baxi SM, Gasasira A, Sserwanga A, Kakeeto S, Nasr S, et al. Indoor Residual Spraying of Insecticide and Malaria Morbidity in a High Transmission Intensity Area of Uganda. *PLoS One*. 2012;7:e42857.
- Ijumba J. Using Lambda-Cyhalothrin for Indoor Residual Spraying against Malaria Vectors in Zanzibar: A Report on Contact Bioassay. Zanzibar: Zanzibar Malaria Control Program; 2008.
- World Health Organization: Report of the Tenth WHOPES Working Group Meeting, 11 to 14 December 2006, Geneva, Switzerland. WHO/CDS_NTD/WHOPES_2007.1. 2007.
- World Health Organization. Indoor Residual Spraying: An Operational Manual for Indoor Residual Spraying (IRS) for Malaria Transmission Control and Elimination. Geneva: World Health Organization; 2013.
- World Health Organization: Guidelines for testing Mosquito adulticides for indoor residual spraying and treatment of mosquito nets. WHO/CDS/NTD/WHOPES/GCDPP/2006.3. 2006.
- World Health Organization: Pesticides and their application for the control of vectors and pests of public health importance. WHO/CDS_NTD/WHOPES/GCDPP/2006.1. 2006.
- Das S, Garver L, Dimopoulos G. Protocol for Mosquito Rearing (*A. gambiae*). *J Vis Exp*. 2007; 5:22.
- Ferro EA, de Arias AR, Ferreira ME, Simancas LC, Rios LS, Rosner JM. Residual effect of lambda-cyhalothrin on *Triatoma infestans*. *Mem Inst Oswaldo Cruz*. 1995;90:415–9.
- de Arias R, Lehane MJ, Schofield CJ, Fournet A. Comparative evaluation of pyrethroid insecticide formulations against *Triatoma infestans* (Klug): residual efficacy on four substrates. *Mem Inst Oswaldo Cruz*. 2003;98:975–80.
- Rojas de Arias A, Lehane MJ, Schofield CJ, Maldonado M. Pyrethroid insecticide evaluation on different house structures in a Chagas disease endemic area of the Paraguayan Chaco. *Mem Inst Oswaldo Cruz*. 2004;99:657–62.
- Penna R, Oliveira AE, Ferreira MF, Johnson C, Bosworth A, Marsden P. The influence of building materials on the residual action of BHC. *Mem Inst Oswaldo Cruz*. 1985;80:443–5.
- dos Santos RLC, Fayal A da S, Aguiar AEF, Vieira DBR, Póvoa MM. Evaluation of the residual effect of pyrethroids on *Anopheles* in the Brazilian Amazon. *Rev Saude Publica*. 2007;41:276–83.
- Giga DP, Canhao J. Persistence of insecticide spray deposits on different surfaces against *Prostephanus truncatus* (Horn.) and *Sitophilus zeamais* (Motsch.). *Int J Trop Insect Sci*. 2011;13:755–62.
- Fletcher MG, Axtell RC. Susceptibility of the bedbug, *Cimex lectularius*, to selected insecticides and various treated surfaces. *Med Vet Entomol*. 1993;7:69–72.
- Barlow F, Hadaway AB. The insecticidal activity of some synthetic pyrethroids against mosquitoes and flies. *Trop Pest Manag*. 1975;21:233–8.
- Campbell WG, Bryant SA. Determination of pH in Wood. *Nature*. 1941;147:357.
- Africa Fighting Malaria. Keeping Malaria Out of Zanzibar. Africa: Fighting Malaria Occasional Paper; 2008.
- World Health Organization. WHO specifications for hand-operated compression sprayers/Delivery and Management of Vector Control Unit, Division of Vector Biology and Control. WHO/VBC/89.970. Geneva: World Health Organization; 1989.
- Ross Institute Bulletin No 1, Ross Institute Information & Advisory. Insecticides. Ross Institute of Tropical Hygiene London; 1979.
- World Health Organization. Manual on Practical Entomology in Malaria: Methods and Techniques. Division of Malaria and Other Parasitic Diseases, Geneva: World Health Organization; 1975.
- Matola YG, Mwita U, Masoud AE. Malaria in the islands of Zanzibar and Pemba 11 years after the suspension of a malaria eradication programme. *Cent Afr J Med*. 1984;30:91–2.
- Najera JA, Zaim M. Malaria vector control: Decision making criteria and procedure for judicious use of insecticide. WHO/CDS/WHOPES/2002.5.Rev.1. Geneva: World Health Organization; 2003.
- De Silans LNMPD, Dedet J-P, Arias JR. Field monitoring of cypermethrin residual effect on the mortality rates of the Phlebotomine sand fly *Lutzomyia longipalpis* in the State of Paraiba, Brazil. *Mem Inst Oswaldo Cruz*. 1998;93:339–44.

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